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#### (57) Abstract

The present invention provides isolated polypeptides useful in the treatment and prevention of malaria caused by *Plasmodium falciparum* or *P. vivax*. In particular, the polypeptides are derived from the binding domains of the proteins in the DBL family as well as the sialic acid binding protein (SABP) on *P. falciparum* merozoites. The polypeptides may also be derived from the Duffy antigen binding protein (DABP) on *P. vivax* merozoites.

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## BINDING DOMAINS FROM PLASMODIUM VIVAX AND PLASMODIUM FALCIPARUM ERYTHROCYTE BINDING PROTEINS

#### BACKGROUND OF THE INVENTION.

Malaria infects 200 - 400 million people each year causing 1-2 million deaths, thus remaining one of the most important infectious diseases in the world. Approximately 25 percent of all deaths of children in rural Africa between the ages of one and four years are caused by malaria. Due to the importance of the disease as a worldwide health problem, considerable effort is being expended to identify and develop malaria vaccines.

Malaria in humans is caused by four species of the parasite *Plasmodium: P. falciparum, P. vivax,*P. knowlesi and P. malariae. The major cause of malaria in humans is P. falciparum which infects 200 million to 400 million people every year, killing 1 to 4 million.

Duffy Antigen Binding Protein (DABP) and Sialic Acid Binding Protein (SABP) are soluble proteins that appear in the culture supernatant after infected erythrocytes release merozoites. Immunochemical data indicate that DABP and SABP which are the respective ligands for the *P. vivax* and *P. falciparum* Duffy and sialic acid receptors on erythrocytes, possess specificities of binding which are identical either in soluble or membrane bound form.

DABP is a 135 kDa protein which binds specifically to Duffy blood group determinants (Wertheimer et al., Exp. Parasitol. 69: 340-350 (1989); Barnwell, et al., J. Exp. Med. 169: 1795-1802 (1989)). Thus, binding of DABP is specific to human Duffy positive erythrocytes. There are four major Duffy phenotypes for human erythrocytes: Fy(a), Fy(b), Fy(ab) and Fy(negative), as defined by the anti-Fy<sup>a</sup> and anti-Fy<sup>b</sup> sera (Hadley et al., In Red Cell Antigens and Antibodies, G. Garratty, ed. (Arlington, Va.:American Association of Blood Banks) pp. 17-33 (1986)). DABP binds equally to both Fy(a) and Fy(b) erythrocytes which are equally susceptible to invasion by P. vivax; but not to Fy(negative) erythrocytes.

In the case of SABP, a 175kDa protein, binding is specific to the glycophorin sialic acid residues on erythrocytes (Camus and Hadley, *Science* 230:553-556 (1985); Orlandi, *et al.*, *J. Cell Biol.* 116:901-909 (1992)). Thus, neuraminidase treatment (which cleaves off sialic acid residues) render erythrocytes immune to *P. falciparus. invasion*.

The specificities of binding and correlation to invasion by the parasite thus indicate that DABP and SABP are the proteins of *P. vivax* and *P. falciparum* which interact with sialic acids and the Duffy antigen on the erythrocyte. The genes encoding both proteins have been cloned and the DNA and predicted protein sequences have been determined (B. Kim Lee Sim, *et al.*, *J. Cell Biol.* 111: 1877-1884 (1990); Fang, X., *et al.*, *Mol. Biochem Parasitol.* 44: 125-132 (1991)).

Despite considerable research efforts worldwide, because of the complexity of the *Plasmodium* parasite and its interaction with its host, it has not been possible to discover a satisfactory solution for prevention or abatement of the blood stage of malaria. Because malaria is a such a large worldwide health <u>problem</u>, there is a need for methods that abate the impact of this disease. The present invention provides effective preventive and therapeutic measures against *Plasmodium* invasion.

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composition.

#### SUMMARY OF THE INVENTION

The present invention provides compositions comprising an isolated DABP binding domain polypeptides and/or isolated SABP binding domain polypeptides. The DABP binding domain polypeptides preferably comprise between about 200 and about 300 amino acid residues while the SABP binding domain polypeptides preferably comprises between about 200 and about 600 amino acid residues. A preferred DABP binding domain polypeptide has about 325 residues of the amino acid sequence found in SEQ ID NO:2. A preferred SABP binding domain polypeptide has about 616 residues of the amino acid sequence of SEQ ID NO:4, encoded by the DNA sequence of SEQ ID NO: 3. The preferred DABP binding domain and SABP binding domain include the cysteine-rich portions of the proteins shown in Figure 1.

The present invention also includes pharmaceutical compositions comprising a pharmaceutically acceptable carrier and an isolated DABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium vivax* merozoites in an organism. In addition, isolated SABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium falciparum* may be added to the pharmaceutical composition.

Also provided are pharmaceutical compositions comprising a pharmaceutically acceptable carrier and an isolated SABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium falciparum* merozoites in an organism. In addition, isolated DABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium vivax* may be added to the pharmaceutical

Isolated polynucleotides which encode a DABP binding domain polypeptides or SABP binding domain polypeptides are also disclosed. In addition, the present invention includes a recombinant cell comprising the polynucleotide encoding the DABP binding domain polypeptide.

The current invention further includes methods of inducing a protective immune response to Plasmodium merozoites in a patient. The methods comprise administering to the patient an immunologically effective amount of a pharmaceutical composition comprising a pharmaceutically acceptable carrier and an isolated DABP binding domain polypeptide, an SABP binding domain polypeptide or a combination thereof.

The present disclosure also provides DNA sequences from additional *P. falciparum* genes in the Duffy-binding like (*DBL*) family that have regions conserved with the *P. falciparum* 175 kD and *P. vivax* 135 kD binding proteins.

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#### DEFINITIONS

As used herein a "DABP binding domain polypeptide" or a "SABP binding domain polypeptide" are polypeptides substantially identical (as defined below) to a sequence from the cysteine-rich, amino-terminal region of the Duffy antigen binding protein (DABP) or sialic acid binding protein (SABP), respectively. Such polypeptides are capable of binding either the Duffy antigen or sialic acid residues on glycophorin. In particular, DABP binding domain polypeptides consist of amino acid residues substantially similar to a sequence of SABP within a binding domain

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containing the cysteine-rich sequence shown in Figure 1. SABP binding domain polypeptides consist of residues substantially similar to a sequence of DABP within a binding domain containing the cysteine-rich sequence shown in Figure 1.

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The binding domain polypeptides encoded by the genes of the *DBL* family consist of those residues substantially identical to the sequence of the binding domains of DABP and SABP as defined above. The DBL family comprises sequences with substantial similarity to the conserved regions of the DABP and SABP. These include those sequences reported here as *ebl-1* (SEQ ID NO:5 and SEQ ID NO:6), E31a (SEQ ID NO:7 and SEQ ID NO:8), *var-7* (SEQ. ID. NO:13 and SEQ. ID. NO:14, GenBank Accession No. L42636) and *var-1* (SEQ. ID. NO:15 and SEQ ID NO:16, GenBank Accession No. L40608). The sequence *ebl-2*, (SEQ ID NO:9 and SEQ ID NO:10) represents the binding domains of *var-7*, and Proj3 (SEQ ID NO:11 and SEQ ID NO:12) is the binding domain of *var-1*. The DBL family also includes two other members *var-2* and *var-3* (GenBank Accession No. L40609).

The polypeptides of the invention can consist of the full length binding domain or a fragment thereof. Typically DABP binding domain polypeptides will consist of from about 50 to about 325 residues, preferably between about 75 and 300, more preferably between about 100 and about 250 residues. SABP binding domain polypeptides will consist of from about 50 to about 616 residues, preferably between about 75 and 300, more preferably between about 100 and about 250 residues.

Particularly preferred polypeptides of the invention are those within the binding domain that are conserved between SABP and the *DBL* family. Residues within these conserved domains are shown in Figure 1, below.

Two polynucleotides or polypeptides are said to be "identical" if the sequence of nucleotides or amino acid residues in the two sequences is the same when aligned for maximum correspondence. Optimal alignment of sequences for comparison may be conducted by the local homology algorithm of Smith and Waterman Adv. Appl. Math. 2: 482 (1981), by the homology alignment algorithm of Needleman and Wunsch J. Mol. Biol. 48:443 (1970), by the search for similarity method of Pearson and Lipman Proc. Natl. Acad. Sci. (U.S.A.) 85: 2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by inspection. term "substantial identity" means that a polypeptide comprises a sequence that has at least 80% sequence identity, preferably 90%, more preferably 95% or more, compared to a reference sequence over a comparison window of about 20 residues to about 600 residues- typically about 50 to about 500 residues usually about 250 to 300 residues. The values of percent identity are determined using the programs above. Particularly preferred peptides of the present invention comprise a sequence in which at least 70% of the cysteine residues conserved in DABP and SABP are present. Additionally, the peptide will comprise a sequence in which at least 50% of the tryptophan residues conserved in DABP and SABP are present. The term substantial similarity is also specifically defined here with respect to those amino acid residues found to be conserved between UABP, SABP and the sequences of the DBL family. These conserved amino acids consist prominently of tryptophan and cysteine residues conserved among all sequences reported here. In addition the conserved amino acid residues include phenylalanine residues which may

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be substituted with tyrosine. These amino acid residues may be determined to be conserved after the sequences have been aligned using methods outlined above by someone skilled in the art.

Another indication that polypeptide sequences are substantially identical is if one protein is immunologically reactive with antibodies raised against the other protein. Thus, the polypeptides of the invention include polypeptides immunologically reactive with antibodies raised against the SABP binding domain, the DABP binding domain or raised against the conserved regions of the *DBL* family.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions. Stringent conditions are sequence dependent and will be different in different circumstances. Generally, stringent conditions are selected to be about 5° C lower than the thermal melting point (Tm) for the specific sequence at a defined ionic strength and pH. The Tm is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Typically, stringent conditions will be those in which the salt concentration is about 0.02 molar at pH 7 and the temperature is at least about 60°C.

Nucleotide sequences are also substaintially identical for purposes of this application when the polypeptides which they encode are substantially identical. Thus, where one nucleic acid sequence encodes essentially the same polypeptide as a second nucleic acid sequence, the two nucleic acid sequences are substantially identical, even if they would not hybridize under stringent conditions due to silent substitutions permitted by the genetic code (see, Darnell et al. (1990) Molecular Cell Biology, Second Edition Scientific American Books, W.H. Freeman and Company, New York, NY, for an explanation of codon degeneracy and the genetic code).

The phrases "isolated" or "biologically pure" refer to material which is substantially or essentially free from components which normally accompany it as found in its native state. Thus, the binding domain polypeptides of this invention do not contain materials normally associated with their in situ environment, e.g., other proteins from a merozoite membrane. Typically, isolated proteins of the invention are at least about 80% pure, usually at least about 90%, and preferrably at least about 95% as measured by band intensity on a silver stained gel.

Protein purity or homogeneity may be indicated by a number of means well known in the art, such as polyacrylamide gel electrophoresis of a protein sample, followed by visualization upon staining. For certain purposes high resolution will be needed and HPLC or a similar means for purification utilized.

The term "residue" refers to an amino acid (D or L) or amino acid mimetic incorporated in a oligopeptide by an amide bond or amide bond mimetic. An amide bond mimetic of the invention includes peptide backbone modifications well known to those skilled in the art.

#### BRIEF DESCRIPTION OF THE DRAWINGS

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Figure 1 represents an alignment of the predicted amino acid sequences of the DABP binding domain (Vivax) (SEQ ID NO:25), the two homologous SABP domains (SABP F1 (SEQ ID NO:26) and SABP F2 (SEQ ID NO:27)) and the sequenced members of the *DBL* gene family (ebl-1 (SEQ ID NO:38), E31a (SEQ ID NO:39), EBL-2 (SEQ ID NO:30)) and the three homologous Proj3 domains (F1 (SEQ ID NO:31), F2 (SEQ ID NO:32) and F3 (SEQ ID NO:33)).

Figure 2 represents a schematic of the pRE4 cloning vector.

Figure 3 shows primers useful for isolating sequences encoding the conserved motifs of the invention. Primers UNIEBP5 (SEQ ID NO:35) and UNIEBP5A (SEQ ID NO:36) encode the amino acid sequence of SEQ ID NO:34; primers UNIEBP5B (SEQ ID NO:38) and UNIEBP5C (SEQ ID NO:39) encode the amino acid sequence of SEQ ID NO:37; primers UNIEBP3 (SEQ ID NO:41) and UNIEBP3A (SEQ ID NO:42) encode the amino acid sequence of SEQ ID NO:40; and primers UNIEBP3B (SEQ ID NO:44) and UNIEBP3C (SEQ ID NO:45) encode the amino acid sequence of SEQ ID NO:43.

Figure 4 shows the relative position of the E31a ORF on chromosome 7.

Figure 5 shows a map of a *var* gene cluster on chromosome 7. Relative positions of four YACs (PfYEF2, PfYFE6, PfYKF8, PfYED9) are indicated under the chromosome 7 line at the top of the figure. YACs PfYFE6 and PfYKF8 lie entirely within a segment linked to CQR in a genetic cross, whereas YACs PfYED9 and PfYEF2 extend beyond sites (identified by pE53a and pH270.5) that are dissociated from the chloroquine response. The *var* cluster extends over a region of 100-150 kb in PfYED9. Exons and introns of the *var-1*, *var-2* and *var-3* genes within the sequenced 40 kb segment are represented by solid and dotted lines, respectively; arrows show the coding direction. Two more *var* elements outside of the sequenced region, identified by conserved restriction sites and cross-hybridization, are indicated by dashed-lines (*var-2c* and *var-3c*). Bold letters mark repeated restriction sites that suggest a duplication in the *var-2/var-3* and *var-2c/var-3c* segments. Enzyme recognition sites: A, *Apal*; B, *Bgh*; C, *Clal*; D, *Hind*III; E, *Hae*III; K, *Kprh*; M, *Bam*HI; P, *Hpal*; S, *Smal*. *Hind*IIII and *Hae*III sites outside of the sequenced region were not mapped. Positions and sizes of inserts from the Dd2 subsegment library are indicated: a, pE280b; b, pB20.3; c, pB600; d, pE21b; e, pB20.24; f, pE32b; h, pE241a; i, pE240a/51d; j, pE33a; k, pB20.23; l, *AL*17BA6; m, pB20.26; n, pB20SU.27; o, p15J2J3. Inserts from the PfYED9 34 kb *Apal-Smal* fragment library: r, pB3; s, p3G11; t, pJVs; u, p2E10; v, pIG3; w, p2E3; x, p2B6; y, PE10; z, pJYr; α, pC5; β, p1A3; γ, p1F6; δ, p3C3; ε, pA2; ζ, p2A9; η, p3C4; θ, pJZn; κ, p3D8.

#### **DESCRIPTION OF THE PREFERRED EMBODIMENT**

The binding of merozoites and schizonts to erythrocytes is mediated by specific binding proteins on the surface of the merozoite or schizont and is necessary for erythrocyte invasion. In the case of *P. falciparum*, this binding involves specific interaction between sialic acid glycophorin residues on the erythrocyte and the sialic acid binding protein (SABP) on the surface of the merozoite or schizont. The ability of purified SABP to bind erythrocytes with chemically or enzymatically altered sialic acid residues paralleled the ability of *P. falciparum* to invade these erythrocytes. Furthermore, sialic acid deficient erythrocytes neither bind SABP nor support invasion by *P. falciparum*. The DNA encoding SABP from *P. falciparum* has also been cloned and sequenced.

In *P. vivax*, specific binding to the erythrocytes involves interaction between the Duffy blood group antigen on the erythrocyte and the Duffy antigen binding protein (DABP) on the merozoite. Duffy binding proteins were defined biologically as those soluble proteins that appear in the culture supernatant after the infected erythrocytes release merozoites which bind to human Duffy positive, but not to human Duffy negative erythrocytes. It has been shown that binding of the *P. vivax* DABP protein to Duffy positive erythrocytes is blocked by antisera to the Duffy blood group determinants. Purified Duffy blood group antigens also block the binding to erythrocytes. DABP has also been shown to bind Duffy blood group determinants on Western blots.

Duffy positive blood group determinants on human erythrocytes are essential for invasion of human erythrocytes by *Plasmodium vivax*. Both attachment and reorientation of *P. vivax* merozoites occur equally well on Duffy positive and negative erythrocytes. A junction then forms between the apical end of the merozoite and the Duffy-positive erythrocyte, followed by vacuole formation and entry of the merozoite into the vacuole. Junction formation and merozoite entry into the erythrocyte do not occur on Duffy negative cells, suggesting that the receptor specific for the Duffy determinant is involved in apical junction formation but not initial attachment. The DNA sequences encoding the DABP from *P. vivax* and *P. knowlesi* have been cloned and sequenced.

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P. vivax red cell invasion has an absolute requirement for the Duffy blood group antigen. Isolates of P. falciparum, however, vary in their dependency on sialic acid for invasion. Certain P. falciparum clones have been developed which invade sialic acid deficient erythrocytes at normal rates. This suggests that certain strains of P. falciparum can interact with other ligands on the erythrocyte and so may possess multiple erythrocyte binding proteins with differing specificities.

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A basis for the present invention is the discovery of the binding domains in both DABP and SABP. Comparison of the predicted protein sequences of DABP and SABP reveals an amino-terminal, cysteine-rich region in both proteins with a high degree of similarity between the two proteins. The amino-terminal, cysteine-rich region of DABP contains about 325 amino acids, whereas the amino-terminal, cysteine-rich region of SABP contains about 616 amino acids. This is due to an apparent duplication of the amino-terminal, cysteine-rich region in the SABP protein. The cysteine residues are conserved between the two regions of SABP and DABP, as are the amino acids surrounding the cysteine residues and a number of aromatic amino acid residues in this region. The amino-terminal cysteine rich region and another cysteine-rich region near the carboxyl-terminus show the most similarity between the DABP and SABP proteins. The region of the amino acid sequence between these two cysteine-rich regions show only limited similarity between DABP and SABP.

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Other *P. falciparum* open reading frames and genes with regions that have substantial identity to binding domains of SABP and DABP have been identified. Multiple copies of these sequences exist in the parasite genome, indicating their important activity in host-parasite interactions. A family of these sequences (the *DBL* family) have been cloned from chromosome 7 subsegment libraries that were constructed during genetic studies of the chloroquine resistance locus (Wellems *et. al.*, *PNAS* 88: 3382-3386 (1991)). Certain of these transcripts are known to be from the *var* family of genes that modulate cytoadherence and antigenic variation of *P. falciparum*- infected erythrocytes (*see*, Example 3, below).

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Genes of the *P. falciparum var* family encode 200-350 kD variant surface molecules that determine antigenic and adhesive properties of parasitized erythrocytes. The large repertoire of *var* genes (50-150 copies, having sufficient DNA to account for 2-6% of the haploid genome), the dramatic sequence variation among the gene copies, their variable expression in different parasite lines, the ready detection of DNA rearrangements, and the receptor binding features of the encoded extracellular domains all implicate *var* genes as the major determinants of antigenic variation and cytoadherence in *P. falciparum* malaria.

A second class of *DBL*-encoding transcripts includes single-copy genes such as *ebl-1*. Genetic linkage studies have placed this gene within a region of chromosome 13 that affects invasion of malarial parasites in human red blood cells (Wellems *et al.*, *Cell* 49:633-642 (1987)). Both SABP and *ebl-1* show restriction patterns that are well conserved among different parasite isolates. This conservation of gene structure and the sequence relationships between the *ebl-1* and SABP domains suggest that *ebl-1* encodes a novel erythrocyte binding molecule having receptor properties distinct from those of SABP.

Southern hybridization experiments using probes from these open reading frames have indicated that additional copies of these conserved sequences are located elsewhere in the genome. The largest of the open reading frames on chromosome 7 is 8 kilobases and contains four tandem repeats homologous to the N-terminal, cysteine-rich unit of SABP and DABP.

Figure 1 represents an alignment of the DBL family with the DABP binding domain and two homologous regions of SABP (F<sub>1</sub> and F<sub>2</sub>). The DBL family is divided into two sub-families to achieve optimal alignment. Conserved cysteine residues are shown in bold face and conserved aromatic residues are underlined.

The polypeptides of the invention can be used to raise monoclonal antibodies specific for the binding domains of SABP, DABP or the conserved regions in the *DBL* gene family. The antibodies can be used for diagnosis of malarial infection or as therapeutic agents to inhibit binding of merozoites to erythrocytes. The production of monoclonal antibodies against a desired antigen is well known to those of skill in the art and is not reviewed in detail here.

The multitude of techniques available to those skilled in the art for production and manipulation of various immunoglobulin molecules can thus be readily applied to inhibit binding. As used herein, the terms "immunoglobulin" and "antibody" refer to a protein consisting of one or more polypeptides substantially encoded by immunoglobulin genes. Immunoglobulins may exist in a variety of forms besides antibodies, including for example, Fv, Fab, and F(ab)<sub>2</sub>, as well as in single chains. For a general review of immunoglobulin structure and function see, Fundamental Immunology, 2d Ed., W.E. Paul ed., Ravens Press, N.Y., (1989).

Antibodies which bind polypeptides of the invention may be produced by a variety of means. The production of non-human monoclonal antibodies, e.g., murine, lagomorpha, equine, etc., is well known and may be accomplished by, for example, immunizing the animal with a preparation containing the polypeptide. Antibody-producing cells obtained from the immunized animals are immortalized and screened, or screened first for the production of antibody which inhibits binding between and meroxoites and erythrocytes and then immortalized.

For a discussion of general procedures of monoclonal antibody production see Harlow and Lane, *Antibodies, A Laboratory Manual* Cold Spring Harbor Publications, N.Y. (1988).

Thus, the present invention allows targeting of protective immune responses or monoclonal antibodies to sequences in the binding domains that are conserved between SABP, DABP and encoded regions of the DBL family. Identification of the binding regions of these proteins facilitates vaccine development because it allows for a focus of effort upon the functional elements of the large molecules. The particular sequences within the binding regions refine the target to critical regions that have been conserved during evolution, and are thus preferred for use as vaccines against the parasite.

The genes of the *DBL* family (which have not previously been sequenced) can be used as markers to detect the presence of the *P. falciparum* parasite in patients. This can be accomplished by means well known to practitioners in the art using tissue or blood from symptomatic patients in PCR reactions with oligonucleotides complementary to portions of the genes of the *DBL* family. Furthermore, sequencing the *DBL* family provides a means for skilled practitioners to generate defined probes to be used as genetic markers in a variety of applications.

Additionally, the present invention defines a conserved motif present in, but not restricted to other members of the subphylum Apicomplexa which participates in host parasite interaction. This motif can be identified in Plasmodium species and other parasitic protozoa by the polymerase chain reaction using the synthetic oligonucleotide primers shown in Figure 3. PCR methods are described in detail below. These primers are designed from regions in the conserved motif showing the highest degree of conservation among DABP, SABP and the DBL family. Figure 3 shows these regions and the consensus amino acid sequences derived from them.

#### A. General Methods

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Much of the nomenclature and general laboratory procedures required in this application can be found in Sambrook, et al., Molecular Cloning A Laboratory Manual, 2nd Ed., Vol. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989. The manual is hereinafter referred to as "Sambrook, et al., 1989."

The practice of this invention involves the construction of recombinant nucleic acids and the expression of genes in transfected cells. Molecular cloning techniques to achieve these ends are known in the art. A wide variety of cloning and *in vitro* amplification methods suitable for the construction of recombinant nucleic acids are well-known to persons of skill. Examples of these techniques and instructions sufficient to direct persons of skill through many cloning exercises are found in Berger and Kimmel, *Guide to Molecular Cloning Techniques, Methods in Enzymology* volume 152 Academic Press, Inc., San Diego, CA (Berger); and *Current Protocols in Molecular Biology*, F.M. Ausubel *et al.*, eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc., (1994 Supplement) (Ausubel).

Examples of techniques sufficient to direct persons of skill through *in vitro* amplification methods, including the polymerase chain reaction (PCR) the ligase chain reaction (LCR), O.B.-replicase amplification and other RNA polymerase mediated techniques are found in Berger, Sambrook et al., 1989, and Ausubel, as well as Mullis et al., (1987) U.S. Patent No. 4,683,202; PCR Protocols A Guide to Methods and Applications (Innis et al. eds), Academic Press Inc., San Diego, CA, 1990) ("Innis"); Arnheim & Levinson (October 1, 1990) C&EN 36-47; The

Journal Of NIH Research (1991) 3, 81-94; Kwoh et al. (1989) Proc. Natl. Acad. Sci. USA 86, 1173; Guatelli et al. (1990) Proc. Natl. Acad. Sci. USA 87, 1874; Lomell et al. (1989) J. Clin. Chem 35, 1826; Landegren et al., (1988) Science 241, 1077-1080; Van Brunt (1990) Biotechnology 8, 291-294; Wu and Wallace, (1989) Gene 4, 560; and Barringer et al. (1990) Gene 89, 117. Improved methods of cloning in vitro amplified nucleic acids are described in Wallace et al., U.S. Pat. No. 5,426,039.

The culture of cells used in the present invention, including cell lines and cultured cells from tissue or blood samples is well known in the art. Freshney (*Culture of Animal Cells, a Manual of Basic Technique, third ed.*, Wiley-Liss, New York, NY (1994)) and the references cited therein provides a general guide to the culture of cells.

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DBL genes are optionally bound by antibodies in one of the embodiments of the present invention. Methods of producing polyclonal and monoclonal antibodies are known to those of skill in the art. See, e.g., Coligan (1991) Current Protocols in Immunology Wiley/Greene, NY; and Harlow and Lane (1989) Antibodies: A Laboratory Manual Cold Spring Harbor Press, NY; Stites et al. (eds.) Basic and Clinical Immunology (4th ed.) Lange Medical Publications, Los Altos, CA, and references cited therein; Goding (1986) Monoclonal Antibodies: Principles and Practice (2d ed.) Academic Press, New York, NY; and Kohler and Milstein (1975) Nature 256: 495-497. Other suitable techniques for antibody preparation include selection of libraries of recombinant antibodies in phage or similar vectors. See, Huse et al. (1989) Science 246: 1275-1281; and Ward, et al. (1989) Nature 341: 544-546. Specific Monoclonal and polyclonal antibodies will usually bind with a KD of at least about .1 mM, more usually at least about 1 µM, and most preferably at least about .1 µM or better.

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#### B. Methods for isolating DNA encoding SABP, DABP and DBL binding regions

The nucleic acid compositions of this invention, whether RNA, cDNA, genomic DNA, or a hybrid of the various combinations, may be isolated from natural sources or may be synthesized in vitro. The nucleic acids claimed may be present in transformed or transfected whole cells, in a transformed or transfected cell lysate, or in a partially purified or substantially pure form.

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Techniques for nucleic acid manipulation of genes encoding the binding domains of the invention, such as subcloning nucleic acid sequences encoding polypeptides into expression vectors, labelling probes, DNA hybridization, and the like are described generally in Sambrook *et al.*, 1989.

Recombinant DNA techniques can be used to produce the binding domain polypeptides. In general, the DNA encoding the SABP and DABP binding domains are first cloned or isolated in a form suitable for ligation into an expression vector. After ligation, the vectors containing the DNA fragments or inserts are introduced into a suitable host cell for expression of the recombinant binding domains. The polypeptides are then isolated from the host cells.

There are various methods of isolating the DNA sequences encoding the SABP, DABP and DBL binding domains. Typically, the DNA is isolated from a genomic or cDNA library using labelled oligonucleotide probes specific for sequences in the DNA. Restriction endonuclease digestion of genomic DNA or cDNA containing the appropriate genes can be used to isolate the DNA encoding the binding domains of these proteins. Since the DNA

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sequences of the SABP and DABP genes are known, a panel of restriction endonucleases can be constructed to give cleavage of the DNA in the desired regions. After restriction endonuclease digestion, DNA encoding SABP binding domain or DABP binding domain is identified by its ability to hybridize with nucleic acid probes, for example on Southern blots, and these DNA regions are isolated by standard methods familiar to those of skill in the art. See Sambrook, et al., 1989.

The polymerase chain reaction can also be used to prepare DABP, SABP DBL binding domain DNA. Polymerase chain reaction technology (PCR) is used to amplify nucleic acid sequences of the DABP and SABP binding domains directly from mRNA, from cDNA, and from genomic libraries or cDNA libraries. The primers shown in Figure 3 are particularly preferred for this process.

Appropriate primers and probes for amplifying the SABP and DABP binding region DNA's are generated from analysis of the DNA sequences. In brief, oligonucleotide primers complementary to the two 3' borders of the DNA region to be amplified are synthesized. The polymerase chain reaction is then carried out using the two primers. See *PCR Protocols: A Guide to Methods and Applications*. (Innis, M, Gelfand, D., Sninsky, J. and White, T., (eds.), Academic Press, San Diego, CA (1990). Primers can be selected to amplify the entire DABP regions or to amplify smaller segments of the DABP and SABP binding domains, as desired.

Oligonucleotides for use as probes are chemically synthesized according to the solid phase phosphoramidite triester method first described by Beaucage, S.L. and Caruthers, M.H., 1981, Tetrahedron Letts., 22(20):1859-1862 using an automated synthesizer, as described in Needham-VanDevanter, D.R., et al. 1984, Nucleic Acids Res., 12:6159-6168. Purification of oligonucleotides is by either native acrylamide gel electrophoresis or by anion-exchange HPLC as described in Pearson, J.D. and Regnier, F.E., 1983, J. Chrom., 255:137-149.

The sequence of the synthetic oligonucleotides can be verified using the chemical degradation method of Maxam, A.M. and Gilbert, 1980, in W., Grossman, L. and Moldave, D., eds. Academic Press, New York, NY, *Methods in Enzymology* 65:499-560.

Other methods known to those of skill in the art may also be used to isolate DNA encoding all or part of the SABP or DABP binding domains. See Sambrook, et al., 1989.

#### C. Expression of DABP, SABP and DBL Binding Domain Polypeptides

Once binding domain DNAs are isolated and cloned, one may express the desired polypeptides in a recombinantly engineered cell such as bacteria, yeast, insect (especially employing baculoviral vectors), and mammalian cells. It is expected that those of skill in the art are knowledgeable in the numerous expression systems available for expression of the DNA encoding the DABP and SABP binding domains. No attempt to describe in detail the various methods known for the expression of proteins in prokaryotes or eukaryotes will be made.

In brief summary, the expression of natural or synthetic nucleic acids encoding binding domains will typically be achieved by operably linking the DNA or cDNA to a promoter (which is either constitutive or inducible), followed by incorporation into an expression vector. The vectors can be suitable for replication and integration in either prokaryotes or eukaryotes. Typical expression vectors contain transcription and translation terminators, initiation sequences, and promoters useful for regulation of the expression of the DNA encoding the

binding domains. To obtain high level expression of a cloned gene, it is desirable to construct expression plasmids which contain, at the minimum, a strong promoter to direct transcription, a ribosome binding site for translational initiation, and a transcription/translation terminator.

#### 1. Expression in Prokaryotes

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Examples of regulatory regions suitable for this purpose in *E. coli* are the promoter and operator region of the *E. coli* tryptophan biosynthetic pathway as described by Yanofsky, C., 1984, J. Bacteriol., 158:1018-1024 and the leftward promoter of phage lambda (P<sub>L</sub>) as described by Herskowitz, I. and Hagen, D., 1980, Ann. Rev. Genet., 14:399-445. The inclusion of selection markers in DNA vectors transformed in *E. coli* is also useful. Examples of such markers include genes specifying resistance to ampicillin, tetracycline, or chloramphenicol. See Sambrook *et al.*, 1989, for details concerning selection markers for use in *E. coli*.

The vector is selected to allow introduction into the appropriate host cell. Bacterial vectors are typically of plasmid or phage origin. Appropriate bacterial cells are infected with phage vector particles or transfected with naked phage vector DNA. If a plasmid vector is used, the bacterial cells are transfected with the plasmid vector DNA.

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Expression systems for expressing the DABP and SABP binding domains are available using *E. coli, Bacillus* sp. (Palva, I *et al.*, 1983, Gene 22:229-235; Mosbach, K. *et al.* Nature, 302:543-545 and *Salmonella. E. coli* systems are preferred.

The binding domain polypeptides produced by prokaryote cells may not necessarily fold properly. During purification from *E. coli*, the expressed polypeptides may first be denatured and then renatured. This can be accomplished by solubilizing the bacterially produced proteins in a chaotropic agent such as guanidine HCl and reducing all the cysteine residues with a reducing agent such as beta-mercaptoethanol. The polypeptides are then renatured, either by slow dialysis or by gel filtration. U.S. Patent No. 4,511,503.

Detection of the expressed antigen is achieved by methods known in the art as radioimmunoassays, Western blotting techniques or immunoprecipitation. Purification from *E. coli* can be achieved following procedures described in U.S. Patent No. 4,511,503.

#### 2. Synthesis of SABP, DABP and DBL Binding Domains in Eukaryotes

A variety of eukaryotic expression systems such as yeast, insect cell lines and mammalian cells, are known to those of skill in the art. As explained briefly below, the DABP and SABP binding domains may also be expressed in these eukaryotic systems.

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#### a. Expression in Yeast

Synthesis of heterologous proteins in yeast is well known and described. *Methods in Yeast Genetics*, Sherman, F., *et al.*, Cold Spring Harbor Laboratory, (1982) is a well recognized work describing the various methods available to produce the binding domains in yeast.

Examples of promoters for use in yeast include GAL1,10 (Johnson, M., and Davies, R.W., 1984, Mol. and Cell. Biol., 4:1440-1448) ADH2 (Russell, D., *et al.* 1983, J. Biol. Chem., 258:2674-2682), PH05 (EMBO J. 6:675-680, 1982), and MF*al* (Herskowitz, I. and Oshima, Y., 1982, in The Molecular Biology of the Yeast

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Saccharomyces, (eds. Strathern, J.N. Jones, E.W., and Broach, J.R., Cold Spring Harbor, Lab., Cold Spring Harbor, N.Y., pp. 181-209. A multicopy plasmid with a selective marker such as Leu-2, URA-3, Trp-1, and His-3 is also desirable.

A number of yeast expression plasmids like YEp6, YEp13, YEp4 can be used as vectors. A gene of interest can be fused to any of the promoters in various yeast vectors. The above-mentioned plasmids have been fully described in the literature (Botstein, et al., 1979, Gene, 8:17-24; Broach, et al., 1979, Gene, 8:121-133).

Two procedures are used in transforming yeast cells. In one case, yeast cells are first converted into protoplasts using zymolyase, lyticase or glusulase, followed by addition of DNA and polyethylene glycol (PEG). The PEG-treated protoplasts are then regenerated in a 3% agar medium under selective conditions. Details of this procedure are given in the papers by J.D. Beggs, 1978, Nature (London), 275:104-109; and Hinnen, A., et al., 1978, Proc. Natl. Acad. Sci. USA, 75:1929-1933. The second procedure does not involve removal of the cell wall. Instead the cells are treated with lithium chloride or acetate and PEG and put on selective plates (Ito, H., et al., 1983, J. Bact., 153:163-168).

The binding domains can be isolated from yeast by lysing the cells and applying standard protein isolation techniques to the lysates. The monitoring of the purification process can be accomplished by using Western blot techniques or radioimmunoassays of other standard immunoassay techniques.

#### b. Expression in Mammalian and Insect Cell Cultures

Illustrative of cell cultures useful for the production of the binding domains are cells of insect or mammalian origin. Mammalian cell systems often will be in the form of monolayers of cells although mammalian cell suspensions may also be used. Illustrative examples of mammalian cell lines include VERO and HeLa cells, Chinese hamster overy (CHO) cell lines, W138, BHK, Cos-7 or MDCK cell lines.

As indicated above, the vector, e. g., a plasmid, which is used to transform the host cell, preferably contains DNA sequences to initiate transcription and sequences to control the translation of the antigen gene sequence. These sequences are referred to as expression control sequences. When the host cell is of insect or mammalian origin illustrative expression control sequences are obtained from the SV-40 promoter (Science, 222:524-527, 1983), the CMV I.E. Promoter (Proc. Natl. Acad. Sci. 81:659-663, 1984) or the metallothionein promoter (Nature 296:39-42, 1982). The cloning vector containing the expression control sequences is cleaved using restriction enzymes and adjusted in size as necessary or desirable and ligated with DNA coding for the SABP or DABP polypeptides by means well known in the art.

As with yeast, when higher animal host cells are employed, polyadenlyation or transcription terminator sequences from known mammalian genes need to be incorporated into the vector. An example of a terminator sequence is the polyadenlyation sequence from the bovine growth hormone gene. Sequences for accurate splicing of the transcript may also be included. An example of a splicing sequence is the VPI intron from SV40 (Sprague, J. et al., 1983, J. Virol. 45: 773-781).

Additionally, gene sequences to control replication in the host cell may be incorporated into the vector such as those found in bovine papilloma virus type-vectors. Saveria-Campo, M., 1985, "Bovine Papilloma virus

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DNA a Eukaryotic Cloning Vector\* in DNA Cloning Vol. II a Practical Approach Ed. D.M. Glover, IRL Press, Arlington, Virginia pp. 213-238.

The host cells are competent or rendered competent for transformation by various means. There are several well-known methods of introducing DNA into animal cells. These include: calcium phosphate precipitation, fusion of the recipient cells with bacterial protoplasts containing the DNA, treatment of the recipient cells with liposomes containing the DNA, DEAE dextran, electroporation and micro-injection of the DNA directly into the cells.

The transformed cells are cultured by means well known in the art. <u>Biochemical Methods in Cell Culture and Virology</u>, Kuchler, R.J., Dowden, Hutchinson and Ross, Inc., (1977). The expressed DABP and SABP binding domain polypeptides are isolated from cells grown as suspensions or as monolayers. The latter are recovered by well known mechanical, chemical or enzymatic means.

#### c. Expression in recombinant vaccinia virus- or adenovirus-infected cells

In addition to use in recombinant expression systems, the isolated binding domain DNA sequences can also be used to transform viruses that transfect host cells in the patient. Live attenuated viruses, such as vaccinia or adenovirus, are convenient alternatives for vaccines because they are inexpensive to produce and are easily transported and administered. Vaccinia vectors and methods useful in immunization protocols are described, for example, in U.S. Patent No. 4,722,848.

Suitable viruses for use in the present invention include, but are not limited to, pox viruses, such as canarypox and cowpox viruses, and vaccinia viruses, alpha viruses, adenoviruses, and other animal viruses. The recombinant viruses can be produced by methods well known in the art, for example, using homologous recombination or ligating two plasmids. A recombinant canarypox or cowpox virus can be made, for example, by inserting the DNA's encoding the DABP and SABP binding domain polypeptides into plasmids so that they are flanked by viral sequences on both sides. The DNA's encoding the binding domains are then inserted into the virus genome through homologous recombination.

A recombinant adenovirus can be produced, for example, by ligating together two plasmids each containing about 50% of the viral sequence and the DNA sequence encoding erythrocyte binding domain polypeptide. Recombinant RNA viruses such as the alpha virus can be made via a cDNA intermediate using methods known in the art.

In the case of vaccinia virus (for example, strain WR), the DNA sequence encoding the binding domains can be inserted in the genome by a number of methods including homologous recombination using a transfer vector, pTKgpt-OFIS as described in Kaslow, et al., Science 252:1310-1313 (1991).

Alternately the DNA encoding the SABP and DABP binding domains may be inserted into another plasmid designed for producing recombinant vaccinia, such as pGS62, Langford, C.L., et al., 1986, Mol. Cell. Biol. 6:3191-3199. This plasmid consists of a cloning site for insertion of foreign genes, the P7.5 promoter of vaccinia to direct synthesis of the inserted gene, and the vaccinia TK gene flanking both ends of the foreign gene.

Confirmation of production of recombinant virus can be achieved by DNA hybridization using cDNA encoding the DABP and SABP binding domain polypeptides and by immunodetection techniques using antibodies

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specific for the expressed binding domain polypeptides. Virus stocks may be prepared by infection of cells such as HELA S3 spinner cells and harvesting of virus progeny.

The recombinant virus of the present invention can be used to induce anti-SABP and anti-DABP binding domain antibodies in mammals, such as mice or humans. In addition, the recombinant virus can be used to produce the SABP and DABP binding domains by infecting host cells *in vitro*, which in turn express the polypeptide (see section on expression of SABP and DABP binding domains in eukaryotic cells, above).

The present invention also relates to host cells infected with the recombinant virus. The host cells of the present invention are preferably mammalian, such as BSC-1 cells. Host cells infected with the recombinant virus express the DABP and SABP binding domains on their cell surfaces. In addition, membrane extracts of the infected cells induce protective antibodies when used to inoculate or boost previously inoculated mammals.

#### D. Purification of the SABP, DABP and DBL Binding Domain Polypeptides

The binding domain polypeptides produced by recombinant DNA technology may be purified by standard techniques well known to those of skill in the art. Recombinantly produced binding domain polypeptides can be directly expressed or expressed as a fusion protein. The protein is then purified by a combination of cell lysis (e. g., sonication) and affinity chromatography. For fusion products, subsequent digestion of the fusion protein with an appropriate proteolytic enzyme release the desired SABP and DABP binding domains.

The polypeptides of this invention may be purified to substantial purity by standard techniques well known in the art, including selective precipitation with such substances as ammonium sulfate, column chromatography, immunopurification methods, and others. See, for instance, R. Scopes, *Protein Purification: Principles and Practice*, Springer-Verlag, New York, NY (1982).

#### E. Production of Binding Domains by protein chemistry techniques

The polypeptides of the invention can be synthetically prepared in a wide variety of ways. For instance polypeptides of relatively short size, can be synthesized in solution or on a solid support in accordance with conventional techniques. Various automatic synthesizers are commercially available and can be used in accordance with known protocols. See, for example, Stewart and Young, *Solid Phase Peptide Synthesis*, 2d. ed., Pierce Chemical Co. (1984).

Alternatively, purified and isolated SABP, DABP or DBL family proteins may be treated with proteolytic enzymes in order to produce the binding domain polypeptides. For example, recombinant DABP and SABP proteins may be used for this purpose. The DABP and SABP protein sequence may then be analyzed to select proteolytic enzymes to be used to generate polypeptides containing desired regions of the DABP and SABP binding domain. The desired polypeptides are then purified by using standard techniques for protein and peptide purification. For a review of standard techniques see, *Methods in Enzymology*, "Guide to Protein Purification", M. Deutscher, ed. Vol. 182 (1990), pages 619-626.

#### F. Modification of nucleic acid and polypeptide sequences

The nucleotide sequences used to transfect the host cells used for production of recombinant binding domain polypeptides can be modified according to standard techniques to yield binding domain polypeptides,

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with a variety of desired properties. The binding domain polypeptides of the present invention can be readily designed and manufactured utilizing various recombinant DNA techniques well known to those skilled in the art. For example, the binding domain polypeptides can vary from the naturally-occurring sequence at the primary structure level by amino acid insertions, substitutions, deletions, and the like. These modifications can be used in a number of combinations to produce the final modified protein chain.

The amino acid sequence variants can be prepared with various objectives in mind, including facilitating purification and preparation of the recombinant polypeptides. The modified polypeptides are also useful for modifying plasma half-life, improving therapeutic efficacy, and lessening the severity or occurrence of side effects during therapeutic use. The amino acid sequence variants are usually predetermined variants not found in nature but exhibit the same immunogenic activity as naturally occurring polypeptides. For instance, polypeptide fragments comprising only a portion (usually at least about 60-80%, typically 90-95%) of the primary structure may be produced. For use as vaccines, polypeptide fragments are typically preferred so long as at least one epitope capable of eliciting production of blocking antibodies remains.

In general, modifications of the sequences encoding the binding domain polypeptides may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis (see, Giliman and Smith, *Gene* 8:81-97 (1979) and Roberts, S. *et al.*, *Nature* 328:731-734 (1987)). One of ordinary skill will appreciate that the effect of many mutations is difficult to predict. Thus, most modifications are evaluated by routine screening in a suitable assay for the desired characteristic. For instance, changes in the immunological character of the polypeptide can be detected by an appropriate competitive binding assay. Modifications of other properties such as redox or thermal stability, hydrophobicity, susceptibility to proteolysis, or the tendency to aggregate are all assayed according to standard techniques.

#### G. <u>Diagnostic and Screening Assays</u>

The polypeptides and nucelic acids of the invention can be used in diagnostic applications for the detection of merozoites or nucleic acids in a biological sample. The presence of parasites can be detected using several well recognized specific binding assays based on immunological results. (See U.S. Patents 4,366,241; 4,376,110; 4,517,288; and 4,837,168). For instance, labeled monoclonal antibodies to polypeptides of the invention can be used to detect merozoites in a biological sample. Alternatively, labelled polypeptides of the invention can be used to detect the presence of antibodies to SABP or DABP in a biological sample. For a review of the general procedures in diagnostic immunoassays, see also *Basic and Clinical Immunology* 7th Edition (D. Stites and A. Terr ed.) 1991.

In addition, modified polypeptides, antibodies or other compounds capable of inhibiting the interaction between SABP or DABP and erythrocytes can be assayed for biological activity. For instance, polypeptides can be recombinantly expressed on the surface of cells and the ability of the cells to bind erythrocytes can be measured as described below. Alternatively, peptides or antibodies can tested for the ability to inhibit binding between erythrocytes and merozoites or SABP and DABP.

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Cell-free assays can also be used to measure binding of DABP or SABP polypeptides to isolated Duffy antigen or glycophorin polypeptides. For instance, the erythrocyte proteins can be immobilized on a solid surface and binding of labelled SABP or DABP polypeptides can be measured.

Many assay formats employ labelled assay components. The labelling systems can be in a variety of forms. The label may be coupled directly or indirectly to the desired component of the assay according to methods well known in the art. A wide variety of labels may be used. The component may be labelled by any one of several methods. The most common method of detection is the use of autoradiography with <sup>3</sup>H, <sup>125</sup>l, <sup>35</sup>S, <sup>14</sup>C, or <sup>32</sup>P labelled compounds or the like. Non-radioactive labels include ligands which bind to labelled antibodies, fluorophores, chemiluminescent agents, enzymes, and antibodies which can serve as specific binding pair members for a labelled ligand. The choice of label depends on sensitivity required, ease of conjugation with the compound, stability requirements, and available instrumentation.

In addition, the polypeptides of the invention can be assayed using animal models, well known to those of skill in the art. For *P falciparum* the *in vivo* models include *Aotus sp.* monkeys or chimpanzees; for *P. vivax* the *in vivo* models include *Saimiri* monkeys.

In the case of the use nucleic acids for diagnostic purposes, standard nucleic hybridization techniques can be used to detect the presence of the genes identified here (e.g., members of the DBL family). If desired, nucleic acids in the sample may first be amplified using standard procedures such as PCR. Diagnostic kits comprising the appropriate primers and probes can also be prepared.

#### H. **DBL** Targeted Therepeutics

DBL polypeptides are expressed on the surface of *Plasmodium*-infected erythrocytes. As such, they present ideal targets for therepeutics which target infected erythrocytes. In one preferred embodiement of the present invention, cytotoxic antibodies or antibody fusion proteins with cytotoxic agents are targeted against *DBL* proteins, killing infected erythrocytes and inhibiting the reproduction of *Plasmodium* in an infected host.

The procedure for attaching a cytotoxic agent to an antibody will vary according to the chemical structure of the agent. Antibodies and cytotoxic agents are typically bound together chemically or, where the antibody and cytotoxic agents are both polypeptides, are optionally synthesized recombinantly as a fusion protein. Polypeptides typically contain variety of functional groups; e.g., carboxylic acid (COOH) or free amine (-NH<sub>2</sub>) groups, which are available for reaction with a suitable functional group on either the antibody or the cytotoxic agent.

Alternatively, antibodies or cytotoxic agents are derivitized to attach additional reactive functional groups. The derivatization optionally involves attachment of linker molecules such as those available from Pierce Chemical Company, Rockford Illinois. A "linker", as used herein, is a molecule that is used to join the nucleic acid binding molecule to the receptor ligand. The linker is capable of forming covalent bonds to both the antibody and the cytotoxic agent. Suitable linkers are well known to those of skill in the art and include, but are not limited to, straight or branched-chain carbon linkers, heterocyclic carbon linkers, or peptide linkers. Where the antibody and the cytotoxic agent are polypeptides, the linkers are joined to the constituent amino acids through their side groups (e.g., through a disulfide linkage to cysteine) or to the alpha carbon amino and carboxyl groups of the terminal amino acids.

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A bifunctional linker having one functional group reactive with a group on a particular ligand, and another group reactive with a nucleic acid binding molecule, can be used to form the desired conjugate. Alternatively, derivatization can proceed through chemical treatment of the ligand or nucleic acid binding molecule, e.g., glycol cleavage of the sugar moiety of a glycoprotein with periodate to generate free aldehyde groups. The free aldehyde groups on the glycoprotein may be reacted with free amine or hydrazine groups on an agent to bind the agent thereto (See, e.g., U.S. Patent No. 4,671,958). Procedures for generation of free sulfhydryl groups on polypeptides, are known (See, e.g., U.S. Pat. No. 4,659,839).

Many procedures and linker molecules for attachment of various compounds to proteins are known. See, for example, European Patent Application No. 188,256; U.S. Patent Nos. 4,671,958, 4,659,839, 4,414,148, 4,699,784; 4,680,338; 4,569,789; and 4,589,071; and Borlinghaus *et al. Cancer Res.* 47: 4071-4075 (1987). In particular, production of various antibody conjugates is well-known within the art and can be found, for example in Thorpe *et al., Monoclonal Antibodies in Clinical Medicine,* Academic Press, pp. 168-190 (1982), Waldmann, *Science*, 252: 1657 (1991), and U.S. Patent Nos. 4,545,985 and 4,894,443.

A number of antibodies which bind cell surface receptors have been converted to form suitable for incorporation into fusion proteins, and similar strategies are used to create fusion-protein antibodies which bind DBR polypeptides. see Batra et al., Mol. Cell. Biol., 11: 2200-2205 (1991); Batra et al., Proc. Natl. Acad. Sci. USA, 89: 5867-5871 (1992); Brinkmann, et al. Proc. Natl. Acad. Sci. USA, 88: 8616-8620 (1991); Brinkmann et al., Proc. Natl. Acad. Sci. USA, 87: 1066-1070 (1990); Friedman et al., Cancer Res. 53: 334-339 (1993); Kreitman et al., J. Immunol., 149: 2810-2815 (1992); Nicholls et al., J. Biol. Chem., 268: 5302-5308 (1993); and Wells, et al., Cancer Res., 52: 6310-6317 (1992), respectively).

#### **B.** Production of Fusion Proteins

Where the antibody fragment and/or the cytotoxic agents are relatively short polypeptides (i.e., less than about 50 amino acids) they are often synthesized using standard chemical peptide synthesis techniques. Where both molecules are relatively short, a chimeric molecule is optionally synthesized as a single contiguous polypeptide. Alternatively, the ligand and the nucleic acid binding molecule can be synthesized separately and then fused chemically.

Solid phase synthesis in which the C-terminal amino acid of the sequence is attached to an insoluble support followed by sequential addition of the remaining amino acids in the sequence is a preferred method for the chemical synthesis of the ligands of this invention. Techniques for solid phase synthesis are described by Barany and Merrifield, Solid-Phase Peptide Synthesis; pp. 3-284 in The Peptides: Analysis, Synthesis, Biology. Vol. 2: Special Methods in Peptide Synthesis, Part A., Merrifield, et al., J. Am. Chem. Soc., 85: 2149-2156 (1963), and Stewart et al., Solid Phase Peptide Synthesis, 2nd ed. Pierce Chem. Co., Rockford, Ill. (1984).

In a preferred embodiment, the fusion molecules of the invention are synthesized using recombinant nucleic acid methodology. Generally this involves creating a nucleic acid sequence that encodes the receptor-targeted fusion molecule, placing the nucleic acid in an expression cassette under the control of a particular promoter, expressing the protein in a host, isolating the expressed protein and, if required, renaturing the protein. Techniques

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sufficient to guide one of skill through such procedures are found in, e.g., Berger, Sambrook, Ausubel, Innis, and Freshney (all supra).

While the two molecules are often joined directly together, one of skill will appreciate that the molecules may be separated by a peptide spacer consisting of one or more amino acids. Generally the spacer will have no specific biological activity other than to join the proteins or to preserve some minimum distance or other spatial relationship between them. However, the constituent amino acids of the spacer may be selected to influence some property of the molecule such as the folding, net charge, or hydrophobicity.

Once expressed, recombinant fusion proteins can be purified according to standard procedures, including ammonium sulfate precipitation, affinity columns, column chromatography, gel electrophoresis and the like (see, generally, R. Scopes, *Protein Purification*, Springer-Verlag, N.Y. (1982), Deutscher, *Methods in Enzymology Vol.* 182: Guide to Protein Purification., Academic Press, Inc. N.Y. (1990)). Substantially pure compositions of about 50 to 95% homogeneity are preferred, and 80 to 95% or greater homogeneity are most preferred for use as therepeutic agents.

One of skill in the art will recognize that after chemical synthesis, biological expression, or purification, the fusion molecule may possess a conformation substantially different than the native conformations of the constituent polypeptides. In this case, it is often necessary to denature and reduce the polypeptide and then to cause the polypeptide to re-fold into the preferred conformation. Methods of reducing and denaturing proteins and inducing re-folding are well known to those of skill in the art (See, Debinski *et al. J. Biol. Chem.*, 268: 14065-14070 (1993); Kreitman and Pastan, *Bioconjug. Chem.*, 4: 581-585 (1993); and Buchner, *et al.*, *Anal. Biochem.*, 205: 263-270 (1992).

### I. Pharmaceutical compositions comprising binding domain polypeptides

The polypeptides of the invention are useful in therapeutic and prophylactic applications for the treatment of malaria. Pharmaceutical compositions of the invention are suitable for use in a variety of drug delivery systems. Suitable formulations for use in the present invention are found in *Remington's Pharmaceutical Sciences*, Mack Publishing Company, Philadelphia, PA, 17th ed. (1985). For a brief review of methods for drug delivery, see, Langer, *Science* 249:1 527-1533 (1990).

The polypeptides of the present invention can be used in pharmaceutical and vaccine compositions that are useful for administration to mammals, particularly humans. The polypeptides can be administered together in certain circumstances, e.g. where infection by both P. falciparum and P. vivax is likely. Thus, a single pharmaceutical composition can be used for the treatment or prophylaxis of malaria caused by both parasites.

The compositions are suitable for single administrations or a series of administrations. When given as a series, inoculations subsequent to the initial administration are given to boost the immune response and are typically referred to as booster inoculations.

The pharmaceutical compositions of the invention are intended for parenteral, topical, oral or local administration. Preferably, the pharmaceutical compositions are administered parenterally, e.g., intravenously, subcutaneously, intradermally, or intramuscularly. Thus, the invention provides compositions for parenteral

administration that comprise a solution of the agents described above dissolved or suspended in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers may be used, e.g., water, buffered water, 0.4% saline, 0.3% glycine, hyaluronic acid and the like. These compositions may be sterilized by conventional, well known sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as is, or hyophilized, the hyophilized preparation being combined with a sterile solution prior to administration. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions, such as pH adjusting and buffering agents, tonicity adjusting agents, wetting agents and the like, for example, sodium acetate, sodium lactate, sodium chloride, potassium chloride, calcium chloride, sorbitan monolaurate, triethanolamine oleate, etc.

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For solid compositions, conventional nontoxic solid carriers may be used which include, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharin, talcum, cellulose, glucose, sucrose, magnesium carbonate, and the like. For oral administration, a pharmaceutically acceptable nontoxic composition is formed by incorporating any of the normally employed excipients, such as those carriers previously listed, and generally 10-95% of active ingredient and more preferably at a concentration of 25%-75%.

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For aerosol administration, the polypeptides are preferably supplied in finely divided form along with a surfactant and propellant. The surfactant must, of course, be nontoxic, and preferably soluble in the propellant. Representative of such agents are the esters or partial esters of fatty acids containing from 6 to 22 carbon atoms, such as caproic, octanoic, lauric, palmitic, stearic, linoleic, linolenic, olesteric and oleic acids with an aliphatic polyhydric alcohol or its cyclic anhydride. Mixed esters, such as mixed or natural glycerides may be employed. A carrier can also be included, as desired, as with, e.g., lecithin for intranasal delivery.

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In certain embodiments patients with malaria may be treated with SABP or DABP polypeptides or other specific blocking agents (e.g. monoclonal antibodies) that prevent binding of *Plasmodium* merozoites and schizonts to the erythrocyte surface.

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The amount administered to the patient will vary depending upon what is being administered, the state of the patient and the manner of administration. In therapeutic applications, compositions are administered to a patient already suffering from malaria in an amount sufficient to inhibit spread of the parasite through erythrocytes and thus cure or at least partially arrest the symptoms of the disease and its complications. An amount adequate to accomplish this is defined as "therapeutically effective dose." Amounts effective for this use will depend on the severity of the disease, the particular composition, and the weight and general state of the patient. Generally, the dose will be in the range of about 1mg to about 5gm per day, preferably about 100 mg per day, for a 70 kg patient.

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Alternatively, the polypeptides of the invention can be used prophylactically as vaccines. The vaccines of the invention contain as an active ingredient an immunogenically effective amount of the binding domain polypeptide or of a recombinant virus as described herein. The immune response may include the generation of antibodies; activation of cytotoxic T lymphocytes (CTL) against cells presenting peptides derived from the peptides encoded by the SABP, DABP or DBL sequences of the present invention, or other mechanisms well known in the art.

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See e.g. Paul Fundamental Immunology, Second Edition (Raven Press, New York, NY) for a description of immune response. Useful carriers are well known in the art, and include, for example, thyroglobulin, albumins such as human serum albumin, tetanus toxoid, polyamino acids such as poly(D-lysine:D-glutamic acid), influenza, hepatitis B virus core protein, hepatitis B virus recombinant vaccine. The vaccines can also contain a physiologically tolerable (acceptable) diluent such as water, phosphate buffered saline, or saline, and further typically include an adjuvant. Adjuvants such as incomplete Freund's adjuvant, aluminum phosphate, aluminum hydroxide, or alum are materials well known in the art.

The DNA or RNA encoding the SABP or DABP binding domains and the DBL gene family motifs may be introduced into patients to obtain an immune response to the polypeptides which the nucleic acid encodes. Wolff et. al., *Science* 247: 1465-1468 (1990) which is describes the use of nucleic acids to produce expression of the genes which the nucleic acids encode.

Vaccine compositions containing the polypeptides, nucleic acids or viruses of the invention are administered to a patient to elicit a protective immune response against the polypeptide. A "protective immune response" is one which prevents or inhibits the spread of the parasite through erythrocytes and thus at least partially prevent the symptoms of the disease and its complications. An amount sufficient to accomplish this is defined as an "immunogenically effective dose." Amounts effective for this use will depend on the composition, the manner of administration, the weight and general state of health of the patient, and the judgment of the prescribing physician. For peptide compositions, the general range for the initial immunization (that is for therapeutic or prophylactic administration) is from about  $100 \mu g$  to about 1 gm of peptide for a 70 kg patient, followed by boosting dosages of from about  $100 \mu g$  to about 1 gm of the polypeptide pursuant to a boosting regimen over weeks to months depending upon the patient's response and condition e.g. by measuring levels of parasite in the patient's blood. For nucleic acids, typically 30-1000ug of nucleic acid is injected into a 70kg patient, more typically about 50-150ug of nucleic acid is injected into a 70kg patient followed by boosting doses as appropriate.

The following examples illustrate preferred embodiments of the invention.

# EXAMPLE 1: <u>Identification of the amino-terminal, cysteine-rich region of SABP and DABP as binding domains for crythrocytes</u>

#### 1. Expression of the SABP binding domain polypeptide on the surface of Cos cells.

To demonstrate that the amino-terminal, cysteine-rich region of the SABP protein is the sialic acid binding region, this region of the protein was expressed on the surface of mammalian Cos cells *in vitro*. This DNA sequence is from position 1 to position 1848 of the SABP DNA sequence (SEQ ID No 3). Polymerase chain reaction technology (PCR) was used to amplify this region of the SABP DNA directly from the cloned gene.

Sequences corresponding to restriction endonuclease sites for Pvull or Apal were incorporated into the oligonucleotide sequence of the probes used in PCR amplification in order to facilitate insertion of the PCR amplified regions into the pHE4 vector (see below). The specific oligonucleotides, 5'-ATCGATCAGCTGGGAAGAATACTTCATCT-3'(SEQID NO:17) and 5'-ATCGATGGGCCCCGAAGTTTGTTCATTATT-3'

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(SEQ ID NO:18) were synthesized. These oligonucleotides were used as primers to PCR-amplify the region of the DNA sequence encoding the cysteine-rich amino terminal region of the SABP protein.

PCR conditions were based on the standard described in Saiki, et al., Science 239: 487-491 (1988). Template DNA was provided from cloned fragments of the gene encoding SABP which had been spliced and re-cloned as a single open-reading frame piece.

The vector, pRE4, used for expression in Cos cells is shown in Figure 2. The vector has an SV40 origin of replication, an ampicillin resistance marker and the Herpes simplex virus glycoprotein D gene (HSV glyd) cloned downstream of the Rous sarcoma virus long terminal repeats (RSV LTR). Part of the extracellular domain of the HSV glyd gene was excised using the Pvull and Apal sites in HSV glyd.

As described above, the PCR oligonucleotide primers contained the Pvull or Apal restriction sites. The PCR-amplified DNA fragments obtained above were digested with the restriction enzymes Pvull and Apal and cloned into the Pvull and Apal sites of the vector pRE4. These constructs were designed to express regions of the SABP protein as chimeric proteins with the signal sequence of HSV glyd at the N-terminal end and the transmembrane and cytoplasmic domain of HSV glyd at the C-terminal end. The signal sequence of HSV glyd targets these chimeric proteins to the surface of Cos cells and the transmembrane segment of HSV glyd anchors these chimeric proteins to the Cos cell surface.

Mammalian Cos cells were transfected with the pRE4 constructs containing the PCR-amplified SABP DNA regions, by calcium phosphate precipitation according to standard techniques.

#### 2. Expression of the DABP binding domain polypeptide on the surface of Cos cells.

To demonstrate that the amino-terminal, cysteine-rich region of the DABP protein is the binding domain, this region was expressed on the surface of Cos cells. This region of the DNA sequence from position 1-975 was first PCR-amplified (SEQ ID No 1).

Sequences corresponding to restriction endonuclease sites for Pvull or Apal were incorporated into the oligonucleotide probes used for PCR amplification in order to facilitate subsequent insertion of the amplified DNA into the pRE4 vector, as described above. The oligonucleotides, 5'-TCTCGTCAGCTGACGATCTCTAGTGCTATT-3' (SEQ ID NO:19) and 5'-ACGAGTGGCCCTGTCACAACTTCCTGAGT-3' (SEQ ID NO:20) were synthesized. These oligonucleotides were used as primers to amplify the region of the DABP DNA sequence encoding the cysteine-rich, amino-terminal region of the DABP protein directly from the cloned DABP gene, using the same conditions described above.

The same pRE4 vector described above in the section on expression of SABP regions in Cos cells was also used as a vector for the DABP DNA regions.

#### 3. <u>Binding studies with erythrocytes.</u>

To demonstrate their ability to bind human erythrocytes, the transfected Cos cells expressing binding domains from DABP and SABP were incubated with erythrocytes for two hours at 37°C in culture media (DMEM/10% FBS). The non-adherent erythrocytes were removed with five washes of phosphate-buffered saline and the bound erythrocytes were observed by light microscopy. Cos cells expressing the amino terminal, cysteine-rich

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SABP polypeptides on their surface bound untreated human erythrocytes, but did not bind neuraminidase treated erythrocytes, that is, erythrocytes which lack sialic acid residues on their surface. Cos cells expressing other regions of the SABP protein on their surface did not bind human erythrocytes. These results identified the amino-terminal, cysteine-rich region of SABP as the erythrocyte binding domain and-indicated that the binding of Cos cells expressing these regions to human erythrocytes is specific. Furthermore, the binding of the expressed region to erythrocytes is identical to the binding pattern seen for the authentic SABP- 175 molecule upon binding to erythrocytes.

Similarly, Cos cells expressing the amino-terminal cysteine-rich region of DABP on their surface bound Duffy-positive human erythrocytes, but did not bind Duffy-negative human erythrocytes, that is erythrocytes which lack the Duffy blood group antigen. Cos cells expressing other regions of the DABP protein on their surface did not bind human erythrocytes. These results identified the amino-terminal cysteine rich region of DABP as the erythrocyte binding domain and indicated that the binding of the Cos cells was specific.

#### **EXAMPLE 2: Isolation of polynucleotide sequences in the DBL family**

P. falciparum clones and cell line used include the following. P. falciparum clones 3D7, D10, LF4/1, Camp/A1, SL/D6, HB3, 7G8, V1/S, T2/C6, KMWII, ItG2F6, FCR3/A2 and Dd2 have been previously tabulated (Dolan, et al. (1993), Mol. Biochem. Parasitol. 61, 137-142). Line Dd2/NM1 was selected from clone Dd2 for invasion via a sialic acid-independent pathway (Dolan, et al. (1990), J. Clin. Invest. 86, 618-624). All parasites were maintained in vitro by standard methods (Trager, et al. (1976), Science 193, 673-675).

DNA and RNA Isolation and Analysis. DNA was extracted as described (Peterson, et al. (1990), Proc. Natl. Acad. Sci. USA 87, 3018-3022). Endonuclease digestion, agarose gel electrophoresis, and filter hybridizations were performed by standard methods (Sambrook, et al., 1989). All hybridizations were at 56°C (Sambrook, et al., 1989). Blots were washed for 2 min. at room temperature in 2x standard saline/phosphate/EDTA (SSPE) with 0.5% SDS, followed by two higher stringency washes at 50°C in 0.3xSSPE with 0.5% SDS. Parasite chromosomes were embedded in agarose blocks and separated by pulsed field gel electrophoresis (Dolan, et al. (1993), Methods. Mol. Biol. 21, 319-332). RNA was isolated from cultured parasites by LiC1 extraction of Catrimox-14-precipitated RNA (Dahle, et al. (1993), BioTechniques 15, 1102-1105). Agarose gel electrophoresis of total RNA and filter hybridizations were performed by standard methods (Sambrook, et al., (1989).

Oligonucleotide Primers and PCR. Primers specific for E31a used in a RT-PCR to test for expression of this sequence were E31aT2 (5'-AGA-CCT-CAA-TTT-CTA-AG-3') (SEQ ID NO:21) and E31aRev1 (5'-AAT-CGC-GAG-CAT-CAT-CTG-3') (SEQ ID NO:22).

Two primers were used to amplify additional sequences from genes encoding *DBL* domains. These were designed from conserved amino acids encoded in the *DBL* domain of the eba-175 and E31a sequences. After adaptation to incorporate the most frequently-used *P. falciparum* codons, forward primer UNIEBP5' [5'-CC(A/G)-AG(G/A)-CAA-(G/A)AA-(C/T)TA-TG-3'] (SEQ ID NO:23), based upon the amino acid sequence PRHUKLU, and reverse primer UNIEBP3' [5'-CUA-(A/T)C(T/G)-(T/G)A(A/G)-(A/G)AA-TTG-(A/T)GG-3'] (SEQ ID NO:24), based upon the amino acid sequence PQFLRW, were synthesized.

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RT-PCR amplifications were performed as described (Kawasaki, et al. (1990), PCR Protocols, A Guide to Methods and Applications, eds. Innis, M.A., Gelfand, D.H., Sninsku, J.J. & White, T.J. (Academic, San Diego), pp. 21-27). In brief, 0.5 to 1 mg of total RNA was treated with RQ1 DNAse (Promega), phenol/chloroform extracted, and ethanol precipitated. The RNA was then annealed with random oligonucleotide primers and extended with Superscript reverse transcriptase (GIBCO/BRL). PCR cycling conditions were 94°C for 10 sec, 45°C for 15 sec, and 72°C for 45 sec, for 30 cycles. All PCRs were performed in an Idaho Technology air thermal cycler using buffer containing 2 mM Mg2+.

PCR amplification products were separated by use of PCR Purity Plus gels and protocols (AT Biochem, Malvern, PA).

DNA Clones and Hybridization Probes. Clone pE31a was isolated from a genomic library prepared from the region of chromosome 7 linked to chloroquine resistance Walker-Jonah, et al. (1992), Mol. Biochem. Parasitol. 51, 313-320. Clone pS31H (GenBank accession no. L38454), containing an insert encompassing that of pE31a, was cloned from a size-selected Hind III restriction digest of Dd2 genomic DNA.

Clone pEBLe1 was cloned from a RT-PCR of Dd2 cDNA after amplification with primers UNIEBP5' (SEQ ID NO:23) and UNIEBP3' (SEQ ID NO:24). Clone pEBP1.2 (GenBank accession no. L38450), containing an insert encompassing that of pEBLe1, was isolated from a Dd2 cDNA library probed with pEBLe1. *DBL*-encoding sequences of *dbl-nm1-4* (GenBank accession no. L38455) and *dbl-nm1-5* (GenBank accession no. L38453) were amplified by RT-PCR from first strand cDNA of line Dd2/NM using primers UNIEBP5' and UNIEBP3'. Sequencing was performed on double stranded DNA templates by standard protocols for the dideoxynucleotide method. (Sequenase; U.S. Biochemicals).

Sequences related to the E31a sequence were detected with the 3005 bp insert of clone pS31H. The *eba-175* gene was detected with a PCR amplified probe consisting of the first 1825 bp of the coding sequence. *ebl-1* sequences were detected with the 2098 bp insert of clone pEBP1.2. All probes were comparable in organization, each containing a region encoding at least one *DBL* domain and varying amounts of flanking sequence.

Homology searches and alignments. Homology searches were performed with BLAST and the Genetics Computer Group program FASTA (Altschul, et al. (1990), J. Mol. Biol. 215, 403-410; Devereux, et al. (1984), Nucleic Acids. Res. 12(1 Pt 1, 387-395). Optimized alignments were produced with MACAW sequence alignment software (Schuler, et al. (1991), Proteins. 9, 180-190).

Multiple P. falciparum sequences encode DBL domains. Positional cloning experiments directed to P. falciparum chromosome 7 identified an ORF (E31a) encoding a DBL domain that is homologous to the domains found in the P. vivax and P. knowlesi DABPs and the P. falciparum SABP. Figure 4 shows the realtive position of the E31a ORF on chromosome 7.

The homology between the *DBL* domains of E31a and the erythrocyte-binding proteins is due to the presence of short motifs of highly conserved amino acids. These well-conserved stretches are separated by non-homologous sequences and by deletions and insertions that vary the size of the domain by greater than 60 aa. The typical *DBL* domain contains 12 or more cysteine residues and has 7 conserved tryptophan residues. Additional

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well conserved amino acids include 4 arginines, 3 aspartates, 9 positions with aliphatic residues (alanine, isoleucine, leucine, or valine) and 4 with aromatic amino acids (tryptophan, phenylalanine, or tyrosine).

Probes spanning the sequence that encodes the E31a *DBL* domain hybridized to multiple fragments within a single restriction digest and yielded bands that varied among parasite lines. The numerous distinct bands from a selection of different parasite DNAs indicated a large number of diverse but related elements. These multiple bands varied among different *P. falciparum* clones, in contrast to the well-conserved, single-copy signal obtained with the *eba-175* probe.

Because of the numerous cross-hybridizing sequences, it seemed likely that many of these related sequences would be on different chromosomes of the parasite. PFG electrophoresis of *P. falciparum* Dd2 chromosomes and hybridization with the E31a probe identified a number of cross-hybridizing sequences on multiple chromosomes. A control hybridization with the *eba-175* probe under identical conditions yielded a single band of hybridization from chromosome 7.

RNA Analysis of *DBL* Elements. Sequences from E31a (pS31H insert) were used to probe RNA blots for corresponding transcripts. No hybridization was detected. Because it was still possible that a message of low abundance was not being detected on the RNA blot, RT-PCR was used as a means of more sensitive detection. For this purpose, cDNA was generated by RT from random primers annealed to DNAse-treated total RNA. E31a-specific oligonucleotides were then used to test for amplification from the cDNA. No amplification of the E31a sequence was obtained, while genomic DNA controls and amplification from cDNA by dihydrofolate reductase/thymidylate synthetase-specific primers yielded the expected bands. A screen of a cDNA library with E31a specific probes also failed to detect any clones hybridizing with the ORF. These results indicate that E31a is either a pseudogene, or is expressed in parasite strains or stages not examined in this work.

A PCR Method to Isolate Sequences Encoding DBL Domains. The identification of short conserved motifs in DBL domains that otherwise have extreme diversity led to a PCR strategy using degenerate oligonucleotide primers designed from conserved amino acid sequences in the DBL domains. Sequences PRRQKLC and PQFLRW were judged most suitable for minimizing degeneracy while allowing amplification of expressed DBL sequences. After these considerations and adjustment for P. falciparum codon usage, primers UNIEBP5' and UNIEBP3' were synthesized.

While some *P. falciparum* lines yielded similar patterns of amplified bands (e. g. Dd2 and MCamp; FCR3/A2 and K-1), no two separate isolates showed identical patterns, reflecting the diversity of the *DBL* domains in the parasite lines. A few bands of the same apparent size were present in many isolates. These included a consistent 490 bp product that was determined to be the *eba-175* gene by its expected size and hybridization to a gene-specific probe. The number of discernible bands probably underestimates the number of amplifiable sequences because of overlapping products of the same size and possible preferential amplification of some sequences over others. Nevertheless, the parasite-specific patterns in the amplified bands may provide a means to quickly type isolates and serves as a measure of parasite diversity in field samples.

To identify *DBL*-encoding sequences in RNA transcripts, the UNIEBP primers were used to amplify first-strand cDNAs generated from DNAse-treated RNA preparations. Amplified products from Dd2, 3D7, HB3 and MCAMP cDNAs had diverse sizes ranging from 400 bp to nearly 1 kb. These included a band at 480-500 bp that was determined to be *eba-175* from its expected size and cross-hybridization to an *eba-175*-specific probe. Other bands were from amplification of different transcripts encoding *DBL* domains. Dd2-NM1 RNA, for example, yielded bands above the *eba-175* product that included two related sequences (*dbl-nm1-4*, *dbl-nm1-5*). These bands were found to be isolate-specific and to have features consistent with the *var* genes described in Example 3, below. Probes that detect *dbl-nm1-4* and *dbl-nm1-5* hybridized to multiple chromosomes and aligned more closely with E31a than with EBA-175 or DABP.

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The RT-PCR amplifications also yielded a consistent band that encoded a novel *DBL* domain distinct from *eba-175*. A cDNA clone corresponding to this product was isolated by screening a *Agt10 Dd2 cDNA* library with a radiolabeled *ebl-1* probe. Sequence from this and additional overlapping cDNA clones confirmed the conserved motifs of the *DBL* domain. The alignment of the predicted amino acid sequences showed that the *DBL* domain of *ebl-1* is more similar to *eba-175* than to the multicopy genes. There was, however, extensive divergence from *eba-175* and other known genes outside of the amplified region.

In contrast to the multicopy hybridization patterns of dbl-nm1-4 and dbl-nm1-5, the ebl-1 sequence, like that of eba-175, was found to have hybridization patterns consistent with a conserved single-copy gene. Probes specific for ebl-1 hybridized only to chromosome 13, and restriction analysis with the enzymes Cla I, EcoRI, HindIII, Hint I, Nsi I, Rsa I, and Spe I, all yielded bands expected from a single copy sequence. RNA blots probed with ebl-1-specific sequences showed several bands of hybridization, however, corresponding to 8-9.5 kb transcripts in mRNA from the Dd2 and 3D7 parasites. The transcripts of different size may result from alternative start and termination points or from incompletely processed species containing introns.

#### **EXAMPLE 3:** Isolation of var genes

Parasite clones, DNA analysis and Chromosome Mapping. Parasite clones were cultivated by the methods of (Trager, et al. (1976), Science 193, 673-675). DNA was extracted from parasite cultures as described (Peterson, et al. (1988), Proc. Natl. Acad. Sci. USA 85, 9114-9118) except that the DNA was as recoverd by ethanol precipitation rather than spooling. Fingerprint analysis with the pC4.H32 probe was used to confirm DNA preparations (Dolan, et al. (1993), Mol. Biochem. Parasitol. 61, 137-142). Southern blotting to Nytran membranes was recommended by the manufacturer (Schleicher & Schuell, Keene, NH). PFG separation of the 14 P. falciparum chromosomes and chromosome mapping were performed as described (Wellems, et al. (1987), Cell 49, 633-642; Sinnis, et al. (1988); Genomics 3, 287-295).

RNA isolation. Parasites from 200 ml mixed stage cultures (5-10% parasitemia) were released by saponin lysis as for DNA preparations except that the procedures were performed with ice-cold solutions. RNA was immediately isolated from the parasite pellet by guanidine thiocyanate/phenol-chloroform methods, recovered and treated with RNAase-free DNAse (Creedon, et al. (1994), J. Biol. Chem. 269, 16364-16370. RNA in H<sub>2</sub>O was combined with 2 vol 100% ETOH, distributed into 2 ml vials and frozen as stock at -70°C. RNA was recovered by

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precipitation with 0.1 vol 3M NaOAc. RNA blots were generated and probed as described (Creedon, et al. (1994), J. Biol. Chem. 269, 16364-16370).

YAC isolation, chromosome-segment libraries and cDNA libraries. Overlapping YACs spanning the 300 kb segment of chromosome 7 that contains the CQR locus were obtained from a YAC library of a CQR FCR3 parasite line de Bruin, et al. (1992), Genomics 14, 332-339) by the procedures of Lanzer, et al. (1993), Nature 361, 654-657. Orientation of the YACs and their overlaps were identified with probes obtained from the YAC ends by inverted PCR.

Attempts to construct cosmid libraries and large insert (~10 kb) A libraries from high molecular weight *P. falciparum* genomic DNA yielded only rearranged clones. An alternative approach was therefore taken in which chromosome-segment libraries were constructed that contained small (0.5-5 kb) inserts in plasmid vectors. Plasmid libraries containing *Alul*, *Hinfl*, *Rsal* and *Sspl* inserts in pCDNAII were constructed from Dd2 chromosome 7 restriction fragments purified by pulsed-field gel (PFG) electrophoresis (Wellems, *et al.* (1991), *Proc. Natl. Acad. Sci. USA* 88, 3382-3386). A plasmid library from a 34 kb *Apal-Smal* restriction fragment of YAC PfYED9 was constructed by the same methods. Inserts in the plasmid libraries were generally 0.5-4 kb.

The Agt10 Dd2 cDNA library was prepared under contract by CloneTech Laboratories Inc. (Palo Alto, CA) from the DNAse-treated, polyA+ fraction of Dd2 RNA. The cDNA was generated in two separate reactions using oligodT primers or random primers. Products of these reactions were combined, processed and cloned into the EcoRI site of Agt10. 1.6 x 10<sup>6</sup> independent recombinants were obtained and amplified.

Isolation of overlapping clones and DNA sequencing. Plasmid clones from the chromosome-segment and YAC-segment libraries were picked at random and their locations were established by restriction mapping. After sequence data from these clones were generated, overlapping clones were isolated in a process of "chromosome walking" by rescreening the libraries with oligonucleotide probes near the ends of sequenced inserts. Sufficient divergence was present among repetitive elements in the sequences to allow distinction of clones and unambiguous assignment of overlaps (generally 50-200 bp).

Sequencing reactions with single-strand M13 DNA (1  $\mu$ g) and double-strand plasmid DNA (2-5  $\mu$ g) were performed in 96-well polyvinyl chloride U-bottom microassay plates using a Sequenase protocol recommended by United States Biochemical Corp. (Cleveland, OH). Reactions were separated by 8M urea-6% polyacrylamide sequencing gels and exposed to Kodak BioMax MR film. Sequence data from some clones were also obtained by use of an ABI 373A automated DNA sequencer (Applied Biosystems Inc., Foster City, CA). Cycle sequencing reactions were performed using the ABI PRISM DyeDeoxy system.

DNA sequence editing, analyses and display were performed with MacVector software (International Biotechnologies Inc., New Haven, CT), BLAST (Altschul, et al. (1990), J. Mol. Biol. 215, 403-410), Genetics Computer Group programs (Devereux, et al. (1984), Nucleic Acids Res. 12, 387-395) and the DNADRAW package (Shapiro, et al. (1986), Nucleic Acids Res. 14, 65-73) maintained at the National Institutes of Health.

resistance. Four overlapping yeast artificial chromosomes from the *P. falciparum* FCR3 line were obtained that span the 300 kb chromosome segment linked to CQR, a segment located 300-600 kb from the telomere of chromosome

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7. Figure 5 shows the positions of these YACs (PfYEF2, PfYFE6, PfYKF8, PfYED9) relative to the chromosome map. In order to define the structure of this 300 kb segment, we performed comparative hybridizations to search for polymorphisms between parasite lines. Clones were randomly picked from chromosome segment-specific plasmid libraries and their inserts were hybridized against restriction digests of the YAC and parasite DNAS. Over thirty inserts were identified that recognized PfYEF2, PfYFE6 or PfYKF8 and showed a predonderance of single copy sequences with few polymorphisms (Alul, Hinfl, Rsal and Sspl digests), consistent with prior findings that chromosome internal regions are largely conserved and contain a preponderance of single copy sequences. However, fifteen other inserts that recognized PfYED9 showed highly polymorphic sets of repetitive elements in the parasite DNAs. Southern analysis indicated that these polymorphic elements were part of a chromosome hypervariable region contained within the PfYED9 clone.

Mapping and DNA sequencing of the hypervariable region spanned by YAC PfYED9. Single copy sequences detected by pE45b and pH270.5 flank the hypervariable region spanned by PfYED9 (Figure 5). The pE45b and pH270.5 probes were therefore used to assign large restriction fragments on the PfYED9 map and establish enzyme recognition sites as reference points. A detailed restriction map of the PfYED9 hypervariable region was then developed. Fifteen overlapping clones ("a"·"f' and "h"·"o" in Figure 5) were isolated by a chromosome walking approach from Dd2 chromosome subsegment libraries (Wellems *et al.*, *supra*) The inserts yielded 19.1 kb of continuous Dd2 sequence having predicted enzyme recognition sites in perfect accord with the PfYED9 restriction map. Such agreement indicates that the Dd2 and FCR3 sequences in this part of the chromosome are very similar, despite differences elsewhere in the genome that are evident by restriction analysis.

We also obtained genomic sequence data from the 34 kb Apal-Smal fragment of PfYED9. Purified PfYED9 DNA was cut with Smal to yield a 110 kb fragment, which was then isolated by PFG electrophoresis and digested with Apal. The resulting 34 kb Apal-Smal band was purified by PFG electrophoresis, digested in four separate reactions by Alul, Hinfl, Rsal or Sspl and incorporated into a plasmid (PCDNAII) library. Cloned inserts from the library were checked for hybridization to the PfYED9 34 kb fragment, assigned to the PfYED9 map and sequenced (Figure 5). Overlapping inserts were obtained by the chromosome walking approach except for three gaps ("t", "z", " $\theta$ " in Figure 5) which were closed by PCR amplification of PfYED9 DNA using primers from flanking sequences. The clones from PfYED9 ("r"-"z"," $\gamma$ ", " $\kappa$ " and " $\alpha$ "+" $\beta$ " in Figure 5) yielded 22.2 kb of continuous DNA sequence that overlaps the Dd2 sequence at the "f"|" $\beta$ " junction and has predicted restriction sites that match the PfYED9 map perfectly. The composite sequence from the Dd2 and PfYED9 segments is 40,171 kb.

Structure of a var gene cluster and comparative analysis of predicted amino acid sequences. The 40,171 bp sequence contains three 10-12 kb regions that have related sequences and structure. Each of these regions harbors a pair of ORFS. The first ORF in each pair begins with a consensus ATG start codon preceded by typical P. falciparum non-coding sequence of abundant A+T content. The ORFs of each pair are separated by an intervening AT-rich and non-coding sequence of 0.9 kb to 1.1 kb. Presence of consensus intron-exon splice junction sequences at either end of these intervening sequences and lack of a consistent translation start site in the 3' ORF indicate that the each pair of ORFs belongs to an individual gene having a two exon structure. This has been verified by

comparison of the genomic sequences to the cDNA sequence of an expressed gene (*var-7*; see subsequent section). The three 10 kb to 12 kb regions thus contain members of a variant gene family which have coding regions of 9.23kb (*var-1*), 7.99 kb (*var-2*) and 9.01 kb (*var-3*). Predicted molecular weights of the encoded proteins are 350 kD, 302 kD and 344 kD. respectively.

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The var genes are flanked by additional members of the var family in PfYED9. Restriction analysis identified two additional genes that are 12-35 kb upstream of the sequenced region and are closely related to var-2 and var-3 (var-2c and Var-3c, Figure 5). The var genes thus have a clustered arrangement in which many individual members are organized in head-to-tail fashion. Between var-1 and var-2 is a 5 kb DNA sequence that harbors a short ORF homologous to that of a repetitive element (rij) suggested to be a transposable element in P. falciparum.

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The deduced protein sequences of the var genes are highly diverse, yet all contain certain conserved motifs and common structural features. Database searches identified 2 to 4 domains within each var sequence that are homologous to cysteine-rich domains of SABP and DABP. In the var sequences, the first domain near the amino-terminus (DBL domain 1) is the most conserved of the DBL domains and has amino acid signatures that differentiate it from subsequent domains (e.g. consensus peptide sequences GAcAp[Y/F]rrL, CTxLARsfadlgdlVrgrdLYLG and VPTYFDYVpqylrwF). Between DBL domains 1 and 2 is another type of conserved domain, a cysteine-rich interdomain region (CIDR) of 300-400 amino acids. The CIDR does not have all the motifs of a DBL domain, but it does have a region at the 3'end which is homologous to the end of the FI DBL domain in SABP. The conservation evident in the sequences of DBL domain I and the CIDR suggest that these regions maintain important structures in the head of the variant molecule.

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DBL domains 2, 3 and 4 (numbering is according to *var-1*, the first sequence completed) have less discriminating signatures than domain 1, and show features of cross-alignment and variation in number that suggest these domains can undergo shuffling and deletion.

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DBL domain 4 is followed by a segment of variable length and a hydrophobic region that is encoded at the end of the first exon (exon 1). In all var sequences this hydrophobic region fits the criteria of a transmembrane segment. The second exon (exon II) encodes a large (45-55 kD) conserved C-terminal sequence that has an acid character (predicted pl = 4.5, vs. 5.9 for the part of the protein upstream of the splice junction) and a cysteine content of < 1% (vs. > 4% upstream). The position of this C-terminal sequence downstream of a single transmembrane segment suggests that it has a cytoplasmic location.

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No consensus signal sequence was detected in the NH<sub>2</sub>-terminal region of the predicted *var* ORFs. We note the presence of several motifs in the protein sequences that are known to act as ligands and receptors in the integrin family. These include RGD (*var-1* codons 886-88, 1992-94) and DGEA (*var-1* codons 2111-14). Not all of these motifs occur in each protein sequence and, when they do occur, their positions vary.

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Identification of var transcripts and chromosome expression sites. To identify transcribed var sequences we screened a Agt10 Dd2 cDNA library with var-containing BssHII restriction fragments that had been purified from PfYED9 and radiolabeled by random hexamer priming. This screening yielded 18 clones with inserts that hybridized back to PfYED9. By cross-hybridization studies and DNA sequence analysis the inserts fell into two groups: group

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I inserts that aligned with sequences of *var* exon I (AT240, AT242, AT244, AT284, AT287, AT288, AT295, AT296); and group II inserts that aligned with sequences of *var* exon II (AT140, AT141, AT142, AT145, AT147, AT148, AT150, AT152).

The full ORF of an expressed var gene (var-7) was determined from \(\lambda\)T242 and overlapping cDNA clones that were obtained by a PCR-based walking strategy. The sequence showed that \(\var-7\) has a 6.6 kb ORF containing two \(\textit{OBL}\) domains, a hydrophobic transmembrane sequence and carboxy-terminal region typical of \(\var\) genes (predicted molecular weight 249 kD). Comparison of \(\var-7\) with the \(\var-1\) sequence demonstrated continuity of the alignments at the predicted splice junction between the ORFs of exons I and II. PCR amplification of Dd2 genomic DNA was also performed with primers derived from the two \(\var-7\) exons. Sequence of this \(\var-7\) PCR product confirmed consensus splice sites and a 1 kb intron typical of the \(\var-7\) genes. Transcription of \(\var-7\) was detected as a 7.5 kb band by RNA blot analysis.

Chromosome mapping experiments with a var-7-specific probe localized the var-7 gene to a region that is 600 kb from one end of Dd2 chromosome 12 (chromosome 12 has a length of 2600 kb). No hybridization of the var-7 probe was detected to any other Dd2 chromosome nor to any chromosomes of the HB3, 3D7 or A4 parasites. Other cDNA inserts from the group I clones were also sequenced and examined for chromosome hybridization signals. The AT240 cDNA insert mapped to the var-1/var-2/var-3 cluster on Dd2 chromosome 7 and its sequence matched that of var-3. The AT244, AT284, AT287, AT288, AT295 and AT296 inserts all showed overlapping sequences and yielded the same hybridization patterns. Chromosome sites recognized by these inserts included regions within two Smal fragments from Dd2 chromosome 7 and another from chromosome 9. We note that loss of a cytoadherence phenotype has been correlated with a chromosome 9 deletion in certain P. falciparum lines.

1.8 kb to 2.4 kb RNA transcripts related to var exon II. In addition to the 7.5 kb var-7 band, a broad 1.8 kb to 2.4 kb band was detected on RNA blots after hybridization with a probe that recognizes var exon II. Sequences of eight group II cDNA inserts homologous to exon II were therefore determined and aligned against the var genes. Comparative analysis of the insert sequences showed that all differed from one another in regions of overlap, indicating that transcription of the corresponding RNAs was from different loci. Three of the cDNA sequences (AT140, AT141 and AT148) aligned downstream of the intron/exon II splice junction. However, five other cDNA inserts (AT142, AT145, AT147, AT150 and AT152) had sequences that aligned upstream of the var intron/exon II splice site and included regions homologous to var intron sequences. In the vicinity of the splice junction, consensus splice sites occurred in three of the cDNA sequences (AT142, AT147, AT150) while a fourth sequence (AT145) showed the required AG dinucleotide but not the expected pyrimidine tract of the splice consensus. The part of the fifth sequence (AT152) that aligned with the var intron extended upstream only to the TAG of the splice sequence. All five sequences lacked a consensus start codon preceded by A+T-rich non-coding DNA that is typical of P. falciparum translation start sites.

<u>Isolate-specific var sequences and evidence for DNA recombination in cultivated parasite clones.</u> The diversity of var forms expressed by *P. falciparum* parasites reflects a tremendous repertoire in the var gene family.

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This repertoire is evident in the patterns of restriction polymorphism detected by *var* probes as well as in the detection of *var*-specific sequences that hybridize to some parasite DNAs but not to others. The *var-7* gene expressed by Dd2, for example, is not present in the HB3, 3D7 or A4 genomes. Such *var* diversity suggests that frequent DNA rearrangements underlie the production of antigenically variant types in different parasite strains.

To test for DNA rearrangements in parasites cultivated *in vitro*, we used *var* sequences to probe restricted DNAs from Dd2 lines adapted to neuraminidase-treated erythrocytes. In one rearrangement a novel 35 kb *Bgl* fragment is seen in NM1 DNA probed with the AT142 (group II) insert. In another rearrangement a deletion of a 20 kb *Pst* band is evident in NM8 DNA probed with a *var-7* sequence. Deletion of this 20 kb band was also detected in the Dd2/R8 subclone obtained before neuraminidase selection, indicating that the DNA rearrangement was

not produced by selection in neuraminidase-treated erythrocytes.

The above examples are provided to illustrate the invention and other variants of the invention encompassed by the claims will be readily apparent to one of ordinary skill in the art.

-31-

## SEQUENCE LISTING

	(1) GENE	RAL INFORMATION:
5	(i) Secretar	APPLICANT: The United States, As Represented by the y, Department of Health and Human Services
10	(ii)	TITLE OF INVENTION: BINDING DOMAINS FROM PLASMODIUM VIVAX AND PLASMODIUM FALCIPARUM ERYTHROCYTE BINDING PROTEINS
10	(iii)	NUMBER OF SEQUENCES: 45
<b>15</b>	(iv)	CORRESPONDENCE ADDRESS:  (A) ADDRESSEE: Knobbe Martens Olson & Bear  (B) STREET: 620 Newport Center Drive 16th Floor  (C) CITY: Newport Beach  (D) STATE: California  (E) COUNTRY: US  (F) ZIP: 92660
20	(v)	COMPUTER READABLE FORM:
25	(*,	(A) MEDIUM TYPE: Floppy disk (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
30	(vi)	CURRENT APPLICATION DATA: (A) APPLICATION NUMBER: (B) FILING DATE: (C) CLASSIFICATION:
35	(vii)	PRIOR APPLICATION DATA (A) APPLICATION NUMBER: US08/487826 (B) FILING DATE: 07-JUN-1996
	(viii)	ATTORNEY/AGENT INFORMATION: (A) NAME: Israelsen, Ned (B) REGISTRATION NUMBER: 29,655 (C) REFERENCE/DOCKET NUMBER: NIH121.001QPC
40	(ix)	TELECOMMUNICATION INFORMATION: (A) TELEPHONE: (619) 235-8550 (B) TELEFAX: (619) 235-0176
45	(2) INFO	RMATION FOR SEQ ID NO:1:
50	(i)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 4084 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear
	(ii)	MOLECULE TYPE: DNA (genomic)
55	(iii)	HYPOTHETICAL: NO
	(vi)	ORIGINAL SOURCE: (A) ORGANISM: Plasmodium vivax
60	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:1:
	AAGCTTTT.	AA AAATAGCAAC AAAATTTCGA AACATTGCCA CAAAAATTTT ATGTTTTACA 60

TATATTTAGA TTCATACAAT TTAGGTGTAC CCTGTTTTTT GATATATGCG CTTAAATTTT 120

TTTTTCGCTC ATATGTTTAG TTATATGTGT AGAACAACTT GCTGAATAAA TTACGTACAC 180 TTTCTGTTCT GAATAATATT ACCACATACA TTTAATTTTA AATACTATGA AAGGAAAAA 240 CCGCTCTTTA TTTGTTCTCC TAGTTTTATT ATTGTTACAC AAGGTATCAT ATAAGGATGA 300 TTTTTCTATC ACACTAATAA ATTATCATGA AGGAAAAAA TATTTAATTA TACTAAAAAG 360 AAAATTAGAA AAAGCTAATA ATCGTGATGT TTGCAATTTT TTTCTTCATT TCTCTCAGGT 420 AAATAATGTA TTATTAGAAC GAACAATTGA AACCCTTCTA GAATGCAAAA ATGAATATGT 480 GAAAGGTGAA AATGGTTATA AATTAGCTAA AGGACACCAC TGTGTTGAGG AAGATAACTT 540 AGAACGATGG TTACAAGGAA CCAATGAAAG AAGAAGTGAG GAAAATATAA AATATAAATA 600 TGGAGTAACG GAACTAAAAA TAAAGTATGC GCAAATGAAT GGAAAAAGAA GCAGCCGCAT 660 10 TTTGAAGGAA TCAATTTACG GGGCGCATAA CTTTGGAGGC AACAGTTACA TGGAGGGAAA 720 AGATGGAGGA GATAAAACTG GGGAGGAAAA AGATGGAGAA CATAAAACTG ATAGTAAAAC 780 TGATAACGGG AAAGGTGCAA ACAATTTGGT AATGTTAGAT TATGAGACAT CTAGCAATGG 840 CCAGCCAGCG GGAACCCTTG ATAATGTTCT TGAATTTGTG ACTGGGCATG AGGGAAATTC 900 TCGTAAAAAT TCCTCGAATG GTGGCAATCC TTACGATATT GATCATAAGA AAACGATCTC 960 TAGTGCTATT ATAAATCATG CITTTCTTCA AAATACTGTA ATGAAAAACT GTAATTATAA 1020 15 GAGAAAACGT CGGGAAAGAG ATTGGGACTG TAACACTAAG AAGGATGTTT GTATACCAGA 1080 TCGAAGATAT CAATTATGTA TGAAGGAACT TACGAATTTG GTAAATAATA CAGACACAA 1140 TTTTCATAGG GATATAACAT TTCGAAAATT ATATTTGAAA AGGAAACTTA TTTATGATGC 1200 TGCAGTAGAG GGCGATTTAT TACTTAAGTT GAATAACTAC AGATATAACA AAGACTTTTG 1260 CAAGGATATA AGATGGAGTT TGGGAGATTT TGGAGATATA ATTATGGGAA CGGATATGGA 1320 20 AGGCATCGGA TATTCCAAAG TAGTGGAAAA TAATTTGCGC AGCATCTTTG GAACTGATGA 1380 AAAGGCCCAA CAGCGTCGTA AACAGTGGTG GAATGAATCT AAAGCACAAA TTTGGACAGC 1440 AATGATGTAC TCAGTTAAAA AAAGATTAAA GGGGAATTTT ATATGGATTT GTAAATTAAA 1500 TGTTGCGGTA AATATAGAAC CGCAGATATA TAGATGGATT CGAGAATGGG GAAGGGATTA 1560 25 CGTGTCAGAA TTGCCCACAG AAGTGCAAAA ACTGAAAGAA AAATGTGATG GAAAAATCAA 1620 TTATACTGAT AAAAAAGTAT GTAAGGTACC ACCATGTCAA AATGCGTGTA AATCATATGA 1680 TCAATGGATA ACCAGAAAAA AAAATCAATG GGATGTTCTG TCAAATAAAT TCATAAGTGT 1740 AAAAAACGCA GAAAAGGTTC AGACGGCAGG TATCGTAACT CCTTATGATA TACTAAAACA 1800 GGAGTTAGAT GAATTTAACG AGGTGGCTTT TGAGAATGAA ATTAACAAAC GTGATGGTGC 1860 30 ATATATTGAG TTATGCGTTT GTTCCGTTGA AGAGGCTAAA AAAAATACTC AGGAAGTTGT 1920 GACAAATGTG GACAATGCTG CTAAATCTCA GGCCACCAAT TCAAATCCGA TAAGTCAGCC 1980 TGTAGATAGT AGTAAAGCGG AGAAGGTTCC AGGAGATTCT ACGCATGGAA ATGTTAACAG 2040 TGGCCAAGAT AGTTCTACCA CAGGTAAAGC TGTTACGGGG GATGGTCAAA ATGGAAATCA 2100 GACACCTGCA GAAAGCGATG TACAGCGAAG TGATATTGCC GAAAGTGTAA GTGCTAAAAA 2160 35 TGTTGATCCG CAGAAATCTG TAAGTAAAAG AAGTGACGAC ACTGCAAGCG TTACAGGTAT 2220 TGCCGAAGCT GGAAAGGAAA ACTTAGGCGC ATCAAATAGT CGACCTTCTG AGTCCACCGT 2280 TGAAGCAAAT AGCCCAGGTG ATGATACTGT GAACAGTGCA TCTATACCTG TAGTGAGTGG 2340 TGAAAACCCA TTGGTAACCC CCTATAATGG TTTGAGGCAT TCGAAAGACA ATAGTGATAG 2400 CGATGGACCT GCGGAATCAA TGGCGAATCC TGATTCAAAT AGTAAAGGTG AGACGGGAAA 2460 GGGGCAAGAT AATGATATGG CGAAGGCTAC TAAAGATAGT AGTAATAGTT CAGATGGTAC 2520 CAGCTCTGCT ACGGGTGATA CTACTGATGC AGTTGATAGG GAAATTAATA AAGGTGTTCC 2580 40 TGAGGATAGG GATAAAACTG TAGGAAGTAA AGATGGAGGG GGGGAAGATA ACTCTGCAAA 2640 TAAGGATGCA GCGACTGTAG TTGGTGAGGA TAGAATTCGT GAGAACAGCG CTGGTGGTAG 2700 CACTAATGAT AGATCAAAAA ATGACACGGA AAAGAACGGG GCCTCTACCC CTGACAGTAA 2760 ACAAAGTGAG GATGCAACTG CGCTAAGTAA AACCGAAAGT TTAGAATCAA CAGAAAGTGG 2820 GGATTTACAA AAGCATGATT TTAAAAGTAA TGATACGCCG AATGAAGAAC CAAATTCTGA 2940 TCAAACTACA GATGCAGAAG GACATGACAG GGATAGCATC AAAAATGATA AAGCAGAAAG 3000 GAGAAAGCAT ATGAATAAAG ATACTTTTAC GAAAAATACA AATAGTCACC ATTTAAATAG 3060 50 TAATAATAAT TTGAGTAATG GAAAATTAGA TATAAAAGAA TACAAATACA GAGATGTCAA 3120 AGCAACAAGG GAAGATATTA TATTAATGTC TTCAGTACGC AAGTGCAACA ATAATATTTC 3180 TTTAGAGTAC TGTAACTCTG TAGAGGACAA AATATCATCG AATACTTGTT CTAGAGAGAA 3240 AAGTAAAAAT TTATGTTGCT CAATATCGGA TTTTTGTTTG AACTATTTTG ACGTGTATTC 3300 TTATGAGTAT CTTAGCTGCA TGAAAAAGGA ATTTGAAGAT CCATCCTACA AGTGCTTTAC 3360 55 GAAAGGGGC TTTAAAGGTA TGCAGAAAAA GATGCTGAAT AGAGAAAGGT GTTGAGTAAA 3420 TTAAAAAGGA ATTAATTTTA GGAATGTTAT AAACATTTTT GTACCCAAAA TTCTTTTTGC 3480 AGACAAGACT TACTTTGCCG CGGCGGAGC GTTGCTGATA CTGCTGTTGT TAATTGCTTC 3540 ANALOTACAN TANCANTTAN ANTANATAN ANTGAGANAT GCCTGTTANT GCACAGTTAN 3660 60 TTCTAACGAT TCCATTTGTG AAGTTTTAAA GAGAGCACAA ATGCATAGTC ATTATGTCCA 3720 TGCATATATA CACATATATG TACGTATATA TAATAAACGC ACACTTTCTT GTTCGTACAG 3780 TTCTGAAGAA GCTACATTTA ATGAGTTTGA AGAATACTGT GATAATATTC ACAGAATCCC 3840 TCTGATGCCT AACAGTAATT CAAATTTCAA GAGCAAAATT CCATTTAAAA AGAAATGTTA 3900 CATCATTTTG CGTTTTTCTT TTTTTCTTT TTTTTCTTT TTTAGATATT GAACACATGC 3960

AGCCATCAAC CCCCCTGGAT TATTCATGAT GCTACTTTGG TAAGTAAAAG CAATTCTGAT 4020 TGTAGTGCTG ATGTAATTTT AGTCATTTTG CTTGCTGCAA TAAACGAGAA AATATATCAA 4080 GCTT 4084

- 5 (2) INFORMATION FOR SEQ ID NO:2:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 1115 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
- 15 (iii) HYPOTHETICAL: NO

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- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Plasmodium vivax
- 20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Lys Gly Lys Asn Arg Ser Leu Phe Val Leu Leu Val Leu Leu Leu 10 Leu His Lys Val Ser Tyr Lys Asp Asp Phe Ser Ile Thr Leu Ile Asn 25 25 Tyr His Glu Gly Lys Lys Tyr Leu Ile Ile Leu Lys Arg Lys Leu Glu 40 45 Lys Ala Asn Asn Arg Asp Val Cys Asn Phe Phe Leu His Phe Ser Gln 30 Val Asn Asn Val Leu Leu Glu Arg Thr Ile Glu Thr Leu Leu Glu Cys 70 75 Lys Asn Glu Tyr Val Lys Gly Glu Asn Gly Tyr Lys Leu Ala Lys Gly 85 90 His His Cys Val Glu Glu Asp Asn Leu Glu Arg Trp Leu Gln Gly Thr 35 100 105 110 Asn Glu Arg Arg Ser Glu Glu Asn Ile Lys Tyr Lys Tyr Gly Val Thr 115 120 125 Glu Leu Lys Ile Lys Tyr Ala Gln Met Asn Gly Lys Arg Ser Ser Arg 135 140 40 . Ile Leu Lys Glu Ser Ile Tyr Gly Ala His Asn Phe Gly Gly Asn Ser 150 155 Tyr Met Glu Gly Lys Asp Gly Gly Asp Lys Thr Gly Glu Glu Lys Asp 165 . 170 175 Gly Glu His Lys Thr Asp Ser Lys Thr Asp Asn Gly Lys Gly Ala Asn 45 180 185 190 Asn Leu Val Met Leu Asp Tyr Glu Thr Ser Ser Asn Gly Gln Pro Ala 200 205 Gly Thr Leu Asp Asn Val Leu Glu Phe Val Thr Gly His Glu Gly Asn 215 220 50 Ser Arg Lys Asn Ser Ser Asn Gly Gly Asn Pro Tyr Asp Ile Asp His 230 235 Lys Lys Thr Ile Ser Ser Ala Ile Ile Asn His Ala Phe Leu Gln Asn 245 250 Thr Val Met Lys Asn Cys Asn Tyr Lys Arg Lys Arg Arg Glu Arg Asp 265 260 Trp Asp Cys Asn Thr Lys Lys Asp Val Cys Ile Pro Asp Arg Arg Tyr 275 280 Gln Leu Cys Met Lys Glu Leu Thr Asn Leu Val Asn Asn Thr Asp Thr 300 295 60 Asn Phe His Arg Asp Ile Thr Phe Arg Lys Leu Tyr Leu Lys Arg Lys 315 310

Leu Ile Tyr Asp Ala Ala Val Glu Gly Asp Leu Leu Leu Lys Leu Asn

Asn Tyr Arg Tyr Asn Lys Asp Phe Cys Lys Asp Ile Arg Trp Ser Leu

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	Gly	Asp	Phe	340 Gly		Ile	Ile	Met	34 Gly		Asp	Met	Glu	35 Gly	Ile	Glv
			355					360	0				36	5	Thr	_
5		370					375	5				38	0	_		_
	Glu 385	Lys	Ala	Gln	Gln	Arg	Arg	Lys	Gln	Trp	Trp 395	Asn	Glu	Ser	Lys	Ala
	Gln	Ile	Trp	Thr	Ala 405		Met	Tyr	Ser	Val	Lys	Lys	Arg	Leu	Lys 41	Gly
10	Asn	Phe	Ile	Trp 420		Cys	Lys	Leu	Asn 42		Ala	Val	Asn	Ile 43	Glu	
			435					44(	0				44	5	Ser	
15		450					455	5			-	46	0 _	•	Lys	
	Asn 465	Tyr	Thr	Asp	Lys	Lys 470		Cys	Lys	Val	Pro 475	Pro	Суз	Gln	Asn	Ala 480
	Суѕ	Lys	Ser	Tyr	Asp 485		Trp	Ile	Thr	Arg 49		Lys	Asn	Gln	Trp	_ =
20				500	_				50	5				<sup>-</sup> 51		
			515					520	)			_	52	5	Ļeu	_
25	Glu	Phe 530	Asn	Glu	Val	Ala	Phe 535		Asn	Glu	Ile	Asn 54		Arg	Asp	Gly
	545					550					555			_	Lys	560
					565	•				57	0				Gln 57	'5
30	Thr	Asn	Ser	Asn 580		Ile	Ser	Gln	Pro 58		Asp	Ser	Ser	Lys 59	Ala 0	Glu
	Lys	Val	Pro 595		Asp	Ser	Thr	His 600		Asn	Val	Asn	Ser 60	_	Gln	Asp
35		610			_	_	615	;		_	_	62	0		Gly	
	Gln 625	Thr	Pro	Ala	Glu	Ser 630	Asp	Val	Gln	Arg	Ser 635	Asp	Ile	Ala	Glu	Ser 640
	Val	Ser	Ala		Asn 645		Asp	Pro	Gln	Lys 65		Val	Ser	Lys	Arg 65	
40		_		660				_	66	5			_	67	-	
		-	675					680	)				68	5	Ala	
45		690	-	_	-		695	;				70	0		Val	
	Gly 705	Glu	Asn	Pro	Leu	Val 710	Thr	Pro	Tyr	Asn	Gly 715	Leu	Arg	His	Ser	Lys 720
	Asp	Asn	Ser	Asp	Ser 725	_	Gly	Pro	Ala	Glu 73		Met	Ala	Asn	Pro	Asp
50	Ser	Asn	Ser	Lys 740	Gly		Thr	Gly	Lys 749	Gly		Asp	Asn	<b>Asp</b> 75	Met	
	Lys	Ala	Thr 755	Lys	Asp	Ser	Ser	Asn 760		Ser	Asp	Gly	Thr 76	Ser	Ser	Ala
55	Thr	Gly 770		Thr	Thr	Asp	Ala 775	Val		Arg	Glu	Ile 78	Asn		Gly	Val
,	Pro 785		Asp	Arg	Asp	Lys 790	–	_	Gly	Ser	Lys 795		-	Gly	Gly	Glu 800
		Asn	Ser	Ala	Asn 805	Lys	Asp	Ala	Ala	Thr	Val	Val	Gly	Glu	Asp	Arg
60	Ile	Arg	Glu	Asn 820	Ser		Gly	Gly	Ser 825	Thr		Asp	Arg	Ser 83	Lys	
	Asp	Thr	Glu 835	١.		Gly	Ala	Ser 840	Thr		Asp	Ser	Lys 84	Gln	Ser	Glu
	Asp	Ala		Ala	Leu	Ser	Lys			Ser	Leu	Glu			Glu	Ser

850 855 Gly Asp Arg Thr Thr Asn Asp Thr Thr Asn Ser Leu Glu Asn Lys Asn 870 875 Gly Gly Lys Glu Lys Asp Leu Gln Lys His Asp Phe Lys Ser Asn Asp 5 885 890 Thr Pro Asn Glu Glu Pro Asn Ser Asp Gln Thr Thr Asp Ala Glu Gly 900 905 His Asp Arg Asp Ser Ile Lys Asn Asp Lys Ala Glu Arg Arg Lys His 915 920 Met Asn Lys Asp Thr Phe Thr Lys Asn Thr Asn Ser His His Leu Asn 10 935 940 Ser Asn Asn Asn Leu Ser Asn Gly Lys Leu Asp Ile Lys Glu Tyr Lys 950 955 Tyr Arg Asp Val Lys Ala Thr Arg Glu Asp Ile Ile Leu Met Ser Ser 15 965 970 Val Arg Lys Cys Asn Asn Asn Ile Ser Leu Glu Tyr Cys Asn Ser Val 980 985 Glu Asp Lys Ile Ser Ser Asn Thr Cys Ser Arg Glu Lys Ser Lys Asn 995 1000 20 Leu Cys Cys Ser Ile Ser Asp Phe Cys Leu Asn Tyr Phe Asp Val Tyr 1010 1015 1020 Ser Tyr Glu Tyr Leu Ser Cys Met Lys Lys Glu Phe Glu Asp Pro Ser 1025 1030 1035 Tyr Lys Cys Phe Thr Lys Gly Gly Phe Lys Ile Asp Lys Thr Tyr Phe 25 1045 1050 Ala Ala Ala Gly Ala Leu Leu Ile Leu Leu Leu Ile Ala Ser Arg Lys 1060 1065 Met Ile Lys Asn Asp Ser Glu Glu Ala Thr Phe Asn Glu Phe Glu Glu 1075 1080 30 Tyr Cys Asp Asn Ile His Arg Ile Pro Leu Met Pro Asn Asn Ile Glu 1090 1095 His Met Gln Pro Ser Thr Pro Leu Asp Tyr Ser 1110 35 (2) INFORMATION FOR SEQ ID NO:3: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 4507 base pairs (B) TYPE: nucleic acid 40 (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) 45 (iii) HYPOTHETICAL: NO (vi) ORIGINAL SOURCE: (A) ORGANISM: Plasmodium falciparum 50 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3: ' TATATATATA TATATATATA GATAATAACA TATAAATATA TTCAATGTGC ATACAATGAA 60 ATGTAATATT AGTATATATT TTTTTGCTTC CTTCTTTGTG TTATATTTTG CAAAAGCTAG 120 GAATGAATAT GATATAAAAG AGAATGAAAA ATTTTTAGAC GTGTATAAAG AAAAATTTAA 180 55 TGAATTAGAT AAAAAGAAAT ATGGAAATGT TCAAAAAACT GATAAGAAAA TATTTACTTT 240 TATAGAAAAT AAATTAGATA TTTTAAATAA TTCAAAATTT AATAAAAGAT GGAAGAGTTA 300 TGGAACTCCA GATAATATAG ATAAAAATAT GTCTTTAATA AATAAACATA ATAATGAAGA 360 AATGTTTAAC AACAATTATC AATCATTTTT ATCGACAAGT TCATTAATAA AGCAAAATAA 420 ATATGTTCCT ATTAACGCTG TACGTGTCTC TAGGATATTA AGTTTCCTGG ATTCTAGAAT 480 TAATAATGGA AGAAATACTT CATCTAATAA CGAAGTTTTA AGTAATTGTA GGGAAAAAAG 540 GAAAGGAATG AAATGGGATT GTAAAAAGAA AAATGATAGA AGCAACTATG TATGTATTCC 600 TGATCGTAGA ATCCAATTAT GCATTGTTAA TCTTAGCATT ATTAAAACAT ATACAAAAGA 660 GACCATGAAG GATCATTTCA TTGAAGCCTC TAAAAAAGAA TCTCAACTTT TGCTTAAAAA 720

AAATGATAAC AAATATAATT CTAAATTTTG TAATGATTTG AAGAATAGTT TTTTAGATTA 780

TGGACATCTT GCTATGGGAA ATGATATGGA TTTTGGAGGT TATTCAACTA AGGCAGAAAA 840 CAAAATTCAA GAAGTTTTTA AAGGGGCTCA TGGGGAAATA AGTGAACATA AAATTAAAAA 900 TTTTAGAAAA GAATGGTGGA ATGAATTTAG AGAGAAACTT TGGGAAGCTA TGTTATCTGA 960 GCATAAAAAT AATATAAATA ATTGTAAAAA TATTCCCCAA GAAGAATTAC AAATTACTCA 1020 ATGGATAAAA GAATGGCATG GAGAATTTTT GCTTGAAAGA GATAATAGAT CAAAATTGCC 1080 AAAAAGTAAA TGTAAAAATA ATACATTATA TGAAGCATGT GAGAAGGAAT GTATTGATCC 1140 ATGTATGAAA TATAGAGATT GGATTATTAG AAGTAAATTT GAATGGCATA CGTTATCGAA 1200 AGAATATGAA ACTCAAAAAG TTCCAAAGGA AAATGCGGAA AATTATTTAA TCAAAATTTC 1260 AGAAAACAAG AATGATGCTA AAGTAAGTTT ATTATTGAAT AATTGTGATG CTGAATATTC 1320 AAAATATTGT GATTGTAAAC ATACTACTAC TCTCGTTAAA AGCGTTTTAA ATGGTAACGA 1380 CAATACAATT AAGGAAAAGC GTGAACATAT TGATTTAGAT GATTTTCTA AATTTGGATG 1440 10 TGATAAAAAT TCCGTTGATA CAAACACAAA GGTGTGGGAA TGTAAAAACC CTTATATATT 1500 ATCCACTAAA GATGTATGTG TACCTCCGAG GAGGCAAGAA TTATGTCTTG GAAACATTGA 1560 TAGAATATAC GATAAAAACC TATTAATGAT AAAAGAGCAT ATTCTTGCTA TTGCAATATA 1620 15 TGAATCAAGA ATATTGAAAC GAAAATATAA GAATAAAGAT GATAAAGAAG TTTGTAAAAT 1680 CATAAATAAA ACTITCGCTG ATATAAGAGA TATTATAGGA GGTACTGATT ATTGGAATGA 1740 TTTGAGCAAT AGAAAATTAG TAGGAAAAAT TAACACAAAT TCAAAATATG TTCACAGGAA 1800 TAAAAAAAT GATAAGCTTT TTCGTGATGA GTGGTGGAAA GTTATTAAAA AAGATGTATG 1860 GAATGTGATA TCATGGGTAT TCAAGGATAA AACTGTTTGT AAAGAAGATG ATATTGAAAA 1920 TATACCACAA TTCTTCAGAT GGTTTAGTGA ATGGGGTGAT GATTATTGCC AGGATAAAAC 1980 AAAAATGATA GAGACTCTGA AGGTTGAATG CAAAGAAAAA CCTTGTGAAG ATGACAATTG 2040 TAAAAGTAAA TGTAATTCAT ATAAAGAATG GATATCAAAA AAAAAAGAAG AGTATAATAA 2100 ACAAGCCAAA CAATACCAAG AATATCAAAA AGGAAATAAT TACAAAATGT ATTCTGAATT 2160 TAAATCTATA AAACCAGAAG TITATTTAAA GAAATACTCG GAAAAATGTT CTAACCTAAA 2220 25 TTTCGAAGAT GAATTTAAGG AAGAATTACA TTCAGATTAT AAAAATAAAT GTACGATGTG 2280 TCCAGAAGTA AAGGATGTAC CAATTTCTAT AATAAGAAAT AATGAACAAA CTTCGCAAGA 2340 AGCAGTTCCT GAGGAAAACA CTGAAATAGC ACACAGAACG GAAACTCCAT CTATCTCTGA 2400 AGGACCAAAA GGAAATGAAC AAAAAGAACG TGATGACGAT AGTTTGAGTA AAATAAGTGT 2460 ATCACCAGAA AATTCAAGAC CTGAAACTGA TGCTAAAGAT ACTTCTAACT TGTTAAAATT 2520 30 AAAAGGAGAT GTTGATATTA GTATGCCTAA AGCAGTTATT GGGAGCAGTC CTAATGATAA 2580 TATAAATGTT ACTGAACAAG GGGATAATAT TTCCGGGGTG AATTCTAAAC CTTTATCTGA 2640 TGATGTACGT CCAGATAAAA AGGAATTAGA AGATCAAAAT AGTGATGAAT CGGAAGAAAC 2700 TGTAGTAAAT CATATATCAA AAAGTCCATC TATAAATAAT GGAGATGATT CAGGCAGTGG 2760 AAGTGCAACA GTGAGTGAAT CTAGTAGTTC AAATACTGGA TTGTCTATTG ATGATGATAG 2820 35 AAATGGTGAT ACATTTGTTC GAACACAAGA TACAGCAAAT ACTGAAGATG TTATTAGAAA 2880 AGAAAATGCT GACAAGGATG AAGATGAAAA AGGCGCAGAT GAAGAAAGAC ATAGTACTTC 2940 TGAAAGCTTA AGTTCACCTG AAGAAAAAAT GTTAACTGAT AATGAAGGAG GAAATAGTTT 3000 AAATCATGAA GAGGTGAAAG AACATACTAG TAATTCTGAT AATGTTCAAC AGTCTGGAGG 3060 AATTGTTAAT ATGAATGTTG AGAAAGAACT AAAAGATACT TTAGAAAATC CTTCTAGTAG 3120 40 CTTGGATGAA GGAAAAGCAC ATGAAGAATT ATCAGAACCA AATCTAAGCA GTGACCAAGA 3180 TATGTCTAAT ACACCTGGAC CTTTGGATAA CACCAGTGAA GAAACTACAG AAAGAATTAG 3240 TAATAATGAA TATAAAGTTA ACGAGAGGGA AGATGAGAGA ACGCTTACTA AGGAATATGA 3300 AGATATTGTT TTGAAAAGTC ATATGAATAG AGAATCAGAC GATGGTGAAT TATATGACGA 3360 AAATTCAGAC TTATCTACTG TAAATGATGA ATCAGAAGAC GCTGAAGCAA AAATGAAAGG 3420 45 AAATGATACA TCTGAAATGT CGCATAATAG TAGTCAACAT ATTGAGAGTG ATCAACAGAA 3480 AAACGATATG AAAACTGTTG GTGATTTGGG AACCACACAT GTACAAAACG AAATTAGTGT 3540 TCCTGTTACA GGAGAAATTG ATGAAAAATT AAGGGAAAGT AAAGAATCAA AAATTCATAA 3600 GGCTGAAGAG GAAAGATTAA GTCATACAGA TATACATAAA ATTAATCCTG AAGATAGAAA 3660 TAGTAATACA TTACATTTAA AAGATATAAG AAATGAGGAA AACGAAAGAC ACTTAACTAA 3720 50 TCAAAACATT AATATTAGTC AAGAAAGGGA TITGCAAAAA CATGGATTCC ATACCATGAA 3780 TAATCTACAT GGAGATGGAG TTTCCGAAAG AAGTCAAATT AATCATAGTC ATCATGGAAA 3840 CAGACAAGAT CGGGGGGAA ATTCTGGGAA TGTTTTAAAT ATGAGATCTA ATAATAATAA 3900 TTTTAATAAT ATTCCAAGTA GATATAATTT ATATGATAAA AAATTAGATT TAGATCITTA 3960 TGAAAACAGA AATGATAGTA CAACAAAAGA ATTAATAAAG AAATTAGCAG AAATAAATAA 4020 55 ATGTGAGAAC GAAATTTCTG TAAAATATTG TGACCATATG ATTCATGAAG AAATCCCATT 4080 AAAAACATGC ACTAAAGAAA AAACAAGAAA TCTGTGTTGT GCAGTATCAG ATTACTGTAT 4140 GAGCTATTTT ACATATGATT CAGAGGAATA TTATAATTGT ACGAAAAGGG AATTTGATGA 4200 TCCATCTTAT ACATGTTTCA GAAAGGAGGC TTTTTCAAGT ATGATATTCA AATTTTTAAT 4260 ABCANATANA ATATATTATT ATTITITATAC TIACAAAACT GCAAAAGTAA CAATAAAAAA 4320 AATTAATTTC TCATTAATTT TTTTTTCTT TTTTTCTTTT TAGGTATGCC ATATTATGCA 4380 60 GGAGCAGGTG TGTTATTTAT TATATTGGTT ATTTTAGGTG CTTCACAAGC CAAATATCAA 4440 AGGTTAGAAA AAATAAATAA AAATAAAATT GAGAAGAATG TAAATTAAAT ATAGAATTCG 4500 AGCTCGG

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#### (2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1435 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Plasmodium falciparum
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Lys Cys Asn Ile Ser Ile Tyr Phe Phe Ala Ser Phe Phe Val Leu 10 20 Tyr Phe Ala Lys Ala Arg Asn Glu Tyr Asp Ile Lys Glu Asn Glu Lys Phe Leu Asp Val Tyr Lys Glu Lys Phe Asn Glu Leu Asp Lys Lys 40 Tyr Gly Asn Val Gln Lys Thr Asp Lys Lys Ile Phe Thr Phe Ile Glu 25 55 Asn Lys Leu Asp Ile Leu Asn Asn Ser Lys Phe Asn Lys Arg Trp Lys 70 75 Ser Tyr Gly Thr Pro Asp Asn Ile Asp Lys Asn Met Ser Leu Ile Asn 90 30 Lys His Asn Asn Glu Glu Met Phe Asn Asn Asn Tyr Gln Ser Phe Leu 105 110 Ser Thr Ser Ser Leu Ile Lys Gln Asn Lys Tyr Val Pro Ile Asn Ala 120 Val Arg Val Ser Arg Ile Leu Ser Phe Leu Asp Ser Arg Ile Asn Asn 35 135 140 Gly Arg Asn Thr Ser Ser Asn Asn Glu Val Leu Ser Asn Cys Arg Glu 150 155 Lys Arg Lys Gly Met Lys Trp Asp Cys Lys Lys Asn Asp Arg Ser 165 170 175 40 Asn Tyr Val Cys Ile Pro Asp Arg Arg Ile Gln Leu Cys Ile Val Asn 180 185 190 Leu Ser Ile Ile Lys Thr Tyr Thr Lys Glu Thr Met Lys Asp His Phe 200 205 Ile Glu Ala Ser Lys Lys Glu Ser Gln Leu Leu Lys Lys Asn Asp 45 215 220 Asn Lys Tyr Asn Ser Lys Phe Cys Asn Asp Leu Lys Asn Ser Phe Leu 230 235 Asp Tyr Gly His Leu Ala Met Gly Asn Asp Met Asp Phe Gly Gly Tyr 245 ·250 255 50 Ser Thr Lys Ala Glu Asn Lys Ile Gln Glu Val Phe Lys Gly Ala His 260 265 Gly Glu Ile Ser Glu His Lys Ile Lys Asn Phe Arg Lys Glu Trp Trp 275 280 Asn Glu Phe Arg Glu Lys Leu Trp Glu Ala Met Leu Ser Glu His Lys 55 295 300 Asn Asn Ile Asn Asn Cys Lys Asn Ile Pro Gln Glu Glu Leu Gln Ile 310 315 Thr Gln Trp Ile Lys Glu Trp His Gly Glu Phe Leu Leu Glu Arg Asp 330 325 60 Asn Arg Ser Lys Leu Pro Lys S r Lys Cys Lys Asn Asn Thr Leu Tyr 345 Glu Ala Cys Glu Lys Glu Cys Ile Asp Pro Cys Met Lys Tyr Arg Asp 360 365

Trp Ile Ile Arg Ser Lys Phe Glu Trp His Thr Leu Ser Lys Glu Tyr

	Cl.	370		T			375					380			_	
	385					Pro 390					395					400
5					405	Asn				410					415	
	Cys	Asp	Ala	Glu 420	Tyr	Ser	Lys	Tyr	Cys 425	Asp	Cys	Lys	His	Thr 430	Thr	Thr
	Leu	Val	Lys 435	Ser	Val	Leu	Asn	Gly			Asn	Thr		Lys	Glu	Lys
10		450	His	Ile		Leu	455	Asp				460			_	_
	Asn 465	Ser	Val	Asp	Thr	Asn 470	Thr	Lys	Val	Trp	Glu 475	Cys	Lys	Asn	Pro	Tyr 480
15	Ile	Leu	Ser	Thr	Lys 485	Asp	Val	Суз	Val	Pro	Pro	Arg	Arg	Gln	Glu 495	Leu
	Cys	Leu	Gly	Asn 500		Asp	Arg	Ile	Tyr 505		Lys	Asn	Leu		Met	Ile
	Lys	Glu	His 515		Leu	Ala	Ile	Ala 520	Ile	Tyr	Glu	Ser	<b>Arg</b> 525	510 Ile	Leu	Lys
20	Arg	Lys 530	Tyr	Lys	Asn	Lys	Asp 535		Lys	Glu	Val	Cys 540		lle	Ile	Asn
	545					Ile 550					555	Gly		_	_	560
25	Asn	Asp	Leu	Ser	Asn 565	Arg	Lys	Leu	Val	Gly 570	Lys	Ile	Asn	Thr	Asn 575	Ser
	Lys	Tyr	Val	His 580	Arg	Asn	Lys	Lys	Asn 585		Lys	Leu	Phe	Arg 590	Asp	Glu
	Trp	Trp	Lys 595	Val	Ile	Lys	Lys	Asp 600	Val	Trp	Asn	Val	Ile 605	Ser	Trp	Val
30	Phe	Lys 610	Asp	Lys	Thr	Val	Cys 615		Glu	Asp	Asp	Ile 620			Ile	Pro
	625					Phe 630					635	Asp				640
35	Lys	Thr	Lys	Met	Ile 645	Glu	Thr	Leu	Lys	Val 650	Glu	Cys	Lys	Glu	Lys 655	Pro
	Cys	Glu	Asp	Asp 660	Asn	Cys	Lys	Ser	Lys 665		Asn	Ser	Tyr	Lys 670	Glu	Trp
-	Ile	Ser	Lys 675	Lys	Lys	Glu	Glu	Tyr 680	Asn	Lys	Gln	Ala	Lys 685	Gln	Tyr	Gln
40	Glu	Tyr 690	Gln	Lys	Gly	Asn	Asn 695	Tyr	Lys	Met	Tyr	Ser 700	Glu	Phe	Lys	Ser
	705					Tyr 710					715					720
45	Leu	Asn	Phe	Glu	Asp 725	Glu	Phe	Lys		Glu 730	Leu	His	Ser	Asp	Tyr 735	Lys
	Asn	Lys	Cys	Thr 740	Met	Cys	Pro	Glu	Val 745	Lys	Asp	Val	Pro	Ile 750	Ser	Ile
	Ile	Arg	Asn 755	Asn	Glu	Gln	Thr	Ser 760	Gln	Glu	Ala	Val	Pro 765	Glu	Glu	Asn
50	Thr	Glu 770	Ile	Ala	His	Arg	Thr 775	Glu	Thr	Pro	Ser	Ile 780	Ser	Glu	Gly	Pro
	Lys 785	Gly	Asn	Glu	Gln	Lys 790	Glu	Arg	Asp	Asp	Asp 795	Ser	Leu	Ser	Lys	Ile 800
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	Ser	Asn	Leu	Leu 820		Leu	Lys	Gly	Asp 825		Asp	Ile	Ser	Met 830		Lys
	Ala	Val	Ile 835		Ser	Ser	Pro	Asn 840		Asn	Ile	Asn	Val 845		Glu	Gln
<b>6</b> 0	Gly	Asp 850		Ile	Ser		Val 855		Ser	Lys	Pro	Leu 860		Asp	Asp	Val
	Arg 865		Asp	Lys				Glu	Asp	Gln	Asn 875	Ser	Asp	Glu	Ser	
	Glu	Thr	Val	Val			Ile	Ser	Lys	Ser	Pro	Ser	Ile	Asn	Asn	880 Gly

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885
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                                920
     Arg Thr Gln Asp Thr Ala Asn Thr Glu Asp Val Ile Arg Lys Glu Asn
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     Ala Asp Lys Asp Glu Asp Glu Lys Gly Ala Asp Glu Glu Arg His Ser
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                                            955
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     Thr Ser Glu Ser Leu Ser Ser Pro Glu Glu Lys Met Leu Thr Asp Asn
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     Glu Gly Gly Asn Ser Leu Asn His Glu Glu Val Lys Glu His Thr Ser
                                     985
                                                        990
     Asn Ser Asp Asn Val Gln Gln Ser Gly Gly Ile Val Asn Met Asn Val
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                               1000
                                                   1005
     Glu Lys Glu Leu Lys Asp Thr Leu Glu Asn Pro Ser Ser Ser Leu Asp
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     Glu Gly Lys Ala His Glu Glu Leu Ser Glu Pro Asn Leu Ser Ser Asp
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     Thr Thr Glu Arg Ile Ser Asn Asn Glu Tyr Lys Val Asn Glu Arg Glu
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                                  1065
                                                      1070
     Asp Glu Arg Thr Leu Thr Lys Glu Tyr Glu Asp Ile Val Leu Lys Ser
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                             1080
                                                  1085
     His Met Asn Arg Glu Ser Asp Asp Gly Glu Leu Tyr Asp Glu Asn Ser
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                                              1100
     Asp Leu Ser Thr Val Asn Asp Glu Ser Glu Asp Ala Glu Ala Lys Met
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                                            1115
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     Lys Gly Asn Asp Thr Ser Glu Met Ser His Asn Ser Ser Gln His Ile
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     Glu Ser Asp Gln Gln Lys Asn Asp Met Lys Thr Val Gly Asp Leu Gly
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                                   1145
                                                       1150
     Thr Thr His Val Gln Asn Glu Ile Ser Val Pro Val Thr Gly Glu Ile
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     Asp Glu Lys Leu Arg Glu Ser Lys Glu Ser Lys Ile His Lys Ala Glu
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     Glu Glu Arg Leu Ser His Thr Asp Ile His Lys Ile Asn Pro Glu Asp
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                                            1195
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     Arg Asn Ser Asn Thr Leu His Leu Lys Asp Ile Arg Asn Glu Glu Asn
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                                      1210
                                                          1215
     Glu Arg His Leu Thr Asn Gln Asn Ile Asn Ile Ser Gln Glu Arg Asp
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                                  1225
                                                       1230
     Leu Gln Lys His Gly Phe His Thr Met Asn Asn Leu His Gly Asp Gly
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                               1240
                                                   1245
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                   1285
                                                          1295
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                                1305
                                                      1310
     Leu Ile Lys Lys Leu Ala Glu Ile Asn Lys Cys Glu Asn Glu Ile Ser
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                               1320
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     Val Lys Tyr Cys Asp His Met Ile His Glu Glu Ile Pro Leu Lys Thr
                           1335
                                              1340
     Cys Thr Lys Glu Lys Thr Arg Asn Leu Cys Cys Ala Val Ser Asp Tyr
                        1350
                                           1355
60
     Cys Met Ser Tyr Phe Thr Tyr Asp Ser Glu Glu Tyr Tyr Asn Cys Thr
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                                    1370
                                                         1375
     Lys Arg Glu Phe Asp Asp Pro Ser Tyr Thr Cys Phe Arg Lys Glu Ala
                                   1385
     Phe Ser Ser Met Ile Phe Lys Phe Leu Ile Thr Asn Lys Ile Tyr Tyr
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GCCGCTCT

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1400
              1395
                                                        1405
       Tyr Phe Tyr Thr Tyr Lys Thr Ala Lys Val Thr Ile Lys Lys Ile Asn
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                                                   1420
       Phe Ser Leu Ile Phe Phe Phe Phe Phe Ser Phe
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                            1430
      (2) INFORMATION FOR SEQ ID NO:5:
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           (i) SEQUENCE CHARACTERISTICS:
                (A) LENGTH: 2288 base pairs
                (B) TYPE: nucleic acid
                (C) STRANDEDNESS: single
15
                (D) TOPOLOGY: linear
          (ii) MOLECULE TYPE: DNA (genomic)
         (iii) HYPOTHETICAL: NO
20
          (vi) ORIGINAL SOURCE:
                (A) ORGANISM: Plasmodium falciparum
          (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:
25
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     GGAAACAGCT ATGACCATGA TTACGCCAAG CTCTAATACG ACTCACTATA GGGAAAGCTG 120
     GTACGCCTGC AGGTCCGGTC CGGAATTCAA TAAAATATTT CCAGAAAGGA ATGTGCAAAT 180
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     TGGTGATAAA ATTTGCAATG CTATATTGGG AAGTTATGCA GATATTGGAG ATATTGTAAG 660
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     TGAAGGAAAG GAAAAAAGTA AAGGTGATCA TTCTTCTCCT GTTCATTCTA AAGATATAAA 1560
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     AAATGAGGAC TCTAATCAAG GGAATAAGGA AACAATAAAT CCTCCTTCTA CAGAAAAAA 1920
     TCTCAAAGAA ATTCATTATA AAACATCTGA TTCTGATGAT CATGGTTCTA AAATTAAAAG 1980
     TCARATTCAR CCARACGAGT TRACGGAGGA ATCACCTCTT ACTGATAGAG AGACTGAGAG 2040
     TGCAGCGATT GGTGATAAAA ATCATGAATC AGTAAAAAGC GCTGATATTT TTCAATCTGA 2100
     GATTCATAAT TCTGATAATA GAGATAGAAT TGTTTCTGAA AGTGTAGTTC AGGATTCTTC 2160
     AGGAAGCTCT ATGAGTACTG AATCTATACG TACTGATAAC AAGGATTTTA AAACAAGTGA 2220
     GGATATTGCA CCTTCTATTA ATGGTCGGAA TTCCCGGGTC GACGAGCTCA CTAGTCGGCG 2280
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WO 96/40766 41. (2) INFORMATION FOR SEQ ID NO:6: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 749 amino acids 5 (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein 10 (iii) HYPOTHETICAL: NO (vi) ORIGINAL SOURCE: (A) ORGANISM: Plasmodium falciparum 15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6: Ala Asp Asn Asn Phe Thr Gln Glu Thr Ala Met Thr Met Ile Thr Pro 10 20 Ser Ser Asn Thr Thr His Tyr Arg Glu Ser Trp Tyr Ala Cys Arg Ser 20 25 Gly Pro Glu Phe Asn Lys Ile Phe Pro Glu Arg Asn Val Gln Ile His Ile Ser Asn Ile Phe Lys Glu Tyr Lys Glu Asn Asn Val Asp Ile Ile 25 55 Phe Gly Thr Leu Asn Tyr Glu Tyr Asn Asn Phe Cys Lys Glu Lys Pro 70 Glu Leu Val Ser Ala Ala Lys Tyr Asn Leu Lys Ala Pro Asn Ala Lys 85 90 100 105 120 130 135 150 155 165 170

30 Ser Pro Arg Ile Tyr Lys Ser Lys Glu His Glu Glu Ser Ser Val Phe Gly Cys Lys Thr Lys Ile Ser Lys Val Lys Lys Lys Trp Asn Cys Tyr 125 Ser Asn Asn Lys Val Thr Lys Pro Glu Gly Val Cys Gly Pro Pro Arg 35 Arg Gln Gln Leu Cys Leu Gly Tyr Ile Phe Leu Ile Arg Asp Gly Asn Glu Glu Gly Leu Lys Asp His Ile Asn Lys Ala Ala Asn Tyr Glu Ala 40 Met His Leu Lys Glu Lys Tyr Glu Asn Ala Gly Gly Asp Lys Ile Cys 185 190 Asn Ala Ile Leu Gly Ser Tyr Ala Asp Ile Gly Asp Ile Val Arg Gly 195 200 205 Leu Asp Val Trp Arg Asp Ile Asn Thr Asn Lys Leu Ser Glu Lys Phe 45 210 215 220 Gln Lys Ile Phe Met Gly Gly Gly Asn Ser Arg Lys Lys Gln Asn Asp 230 235 Asn Asn Glu Arg Asn Lys Trp Trp Glu Lys Gln Arg Asn Leu Ile Trp 245 250 50 Ser Ser Met Val Lys His Ile Pro Lys Gly Lys Thr Cys Lys Arg His 260 265 270 Asn Asn Phe Glu Lys Ile Pro Gln Phe Leu Arg Trp Leu Lys Glu Trp 280 Gly Asp Glu Phe Cys Glu Glu Met Gly Thr Glu Val Lys Gln Leu Glu 55 290 295 300 Lys Ile Cys Glu Asn Lys Asn Cys Ser Glu Lys Lys Cys Lys Asn Ala 310 315 Cys Ser Ser Tyr Glu Lys Trp Ile Lys Glu Arg Lys Asn Glu Tyr Asn 325 330 335 60 Leu Gln Ser Lys Lys Phe Asp Ser Asp Lys Leu Asn Lys Lys Asn 340 345 Asn Leu Tyr Asn Lys Phe Glu Asp Ser Lys Ala Tyr Leu Arg Ser Glu 355 360 Ser Lys Gln Cys Ser Asn Ile Glu Phe Asn Asp Glu Thr Phe Thr Phe

		370					37	5				3	В0			
	Pro 385		Lys	Tyr	Lys	Glu 390	Ala		Met	Val	Cys 395	Glu	Asn	Pro	Ser	Ser
5	Ser	Lys	Ala	Leu	Lys 405		Ile	Lys	Thr	Asn 41	Val	Phe	Pro	Ile		Glu 15
			Lys	420	1				42	Thr	Asp			4	Asn 30	Thr
			Ser 435					44	0				4	Ile 45	Ser	
10		450					45	5				4	60		_	
	465		Val			470					475					480
15			Asn		485	;				49	0				4	95
			Ser	500					50	5				5	10	
			Val 515					52	0				5:	25		
20		530					53	5				54	10	_		
	545		Ser			550					555	•				560
25			Val		565	i				57	0				5	75
			Ile	580					58	5				5	90	_
			Ser 595					60	0				60	)5		
30		610	Ser				619	5				62	20			
	625		Asn			630					635				_	640
35			Ser		645					65	0				6	55
			Pro	660					66	5				6.	70	
			His 675					686	) ·				68	35		
40		690	Ser				695	5				70	0			
	705		Ser			710					715				_	720
<b>4</b> 5			Phe		725					73	0					Arg 35
	Asn	Ser	Arg	Val 740	Asp	Glu	Let	ı Thi	74		g Ar	g Pr	o Le	eu		
	(2) INFOR	MATT	ION I	നേട	SEO	TD N	m · 7 ·	•								

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2606 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:

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- (A) ORGANISM: Plasmodium falciparum
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

```
AGCTCTATTA CGACTCACTA TAGGGAAAGC TGGTACGCCT GCAGGTACCG GTCCGGAATT 60
      CCCGGGTCGA CGAGCTCACT AGTCGGCGGC CGCTCTAGAG GATCCAAGCT TAATAGTGTT 120
      TATACGTCTA TTGGCTTATT TTTAAATAGC TTAAAAAGCG GACCATGTAA AAAGGATAAT 180
      GATAATGCAG AGGATAATAT AGATTTTGGT GATGAAGGTA AAACATTTAA AGAGGCAGAT 240
 5
     AATTGTAAAC CATGTTCTCA ATTTACTGTT GATTGTAAAA ATTGTAATGG TGGTGATACA 300
      AAAGGGAAGT GCAATGGCAG CAATGGCAAA AAGAATGGAA ATGATTATAT TACTGCAAGT 360
     GATATTGAAA ATGGAGGGAA TTCTATTGGA AATATAGATA TGGTTGTTAG TGATAAGGAT 420
     GCAAATGGAT TTAATGGTTT AGACGCTTGT GGAAGTGCAA ATATCTTTAA AGGTATTAGA 480
      AAAGAACAAT GGAAATGTGC TAAAGTATGT GGTTTAGATG TATGTGGTCT TAAAAATGGT 540
10
     AATGGTAGTA TAGATAAAGA TCAAAAACAA ATTATAATTA TTAGAGCATT GCTTAAACGT 600
     TGGGTAGAAT ATTTTTTAGA AGATTATAAT AAAATTAATG CCAAAATTTC ACATTGTACG 660
     AAAAAGGATA ATGAATCCAC ATGTACAAAT GATTGTCCAA ATAAATGTAC ATGTGTAGAA 720
     GAGTGGATAA ATCAGAAAAG GACAGAATGG AAAAATATAA AAAAACATTA CAAAACACAA 780
     AATGAAAATG GTGACAATAA CATGAAATCT TTGGTTACAG ATATTTTGGG TGCCTTGCAA 840
15
     CCCCAAAGTG ATGTTAACAA AGCTATAAAA CCTTGTAGTG GTTTAACTGC GTTCGAGAGT 900
     TTTTGTGGTC TTAATGGCGC TGATAACTCA GAAAAAAAG AAGGTGAAGA TTACGATCTT 960
     GTTCTATGTA TGCTTAAAAA TCTTGAAAAA CAAATTCAGG AGTGCAAAAA GAAACATGGC 1020
     GAAACTAGTG TCGAAAATGG TGGCAAATCA TGTACCCCCC TTGACAACAC CACCCTTGAG 1080
     GAGGAACCCA TAGAAGAGGA AAACCAAGTG GAAGCGCCGA ACATTTGTCC AAAACAAACA 1140
     GTGGAAGATA AAAAAAAAGA GGAAGAAGAA GAAACTTGTA CACCGGCATC ACCAGTACCA 1200
20
     GAAAAACCGG TACCTCATGT GGCACGTTGG CGAACATTTA CACCACCTGA GGTATTCAAG 1260
     ATATGGAGGG GAAGGAGAA TAAAACTACG TGCGAAATAG TGGCAGAAAT GCTTAAAGAT 1320
AAGAATGGAA GGACTACAGT AGGTGAATGT TATAGAAAAG AAACTTATTC TGAATGGACG 1380
     TGTGATGAAA GTAAGATTAA AATGGGACAG CATGGAGCAT GTATTCCTCC AAGAAGACAA 1440
     AAATTATGTT TACATTATTT AGAAAAAATA ATGACAAATA CAAATGAATT GAAATACGCA 1500
25
     TTTATTAAAT GTGCTGCAGC AGAAACTTTT TTGTTATGGC AAAACTACAA AAAAGATAAG 1560
     AATGGTAATG CAGAAGATCT CGATGAAAAA TTAAAAGGTG GTATTATCCC CGAAGATTTT 1620
     AAACGGCAAA TGTTCTATAC GTTTGCAGAT TATAGAGATA TATGTTTGGG TACGGATATA 1680
     TCATCAAAAA AAGATACAAG TAAAGGTGTA GGTAAAGTAA AATGCAATAT TGATGATGTT 1740
30
     TTTTATAAAA TTAGCAATAG TATTCGTTAC CGTAAAAGTT GGTGGGAAAC AAATGGTCCA 1800
     GTTATATGGG AAGGAATGTT ATGCGCTTTA AGTTATGATA CGAGCCTAAA TAATGTTAAT 1860
     CCGGAAACTC ACAAAAAACT TACCGAAGGC AATAACAACT TTGAGAAAGT CATATTTGGT 1920
     AGTGATAGTA GCACTACTTT GTCCAAATTT TCTGAAAGAC CTCAATTTCT AAGATGGTTG 1980
     ACTGAATGGG GAGAAAATTT CTGCAAAGAA CAAAAAAAGG AGTATAAGGT GTTGTTGGCA 2040
35
     AAATGTAAGG ATTGTGATGT TGATGGTGAT GGTAAATGTA ATGGAAAATG TGTTGCGTGC 2100
     AAAGATCAAT GTAAACAATA TCATAGTTGG ATTGGAATAT GGATAGATAA TTATAAAAAA 2160
     CAAAAAGGAA GATATACTGA GGTTAAAAAA ATACCTCTGT ATAAAGAAGA TAAAGACGTG 2220
     AAAAACTCAG ATGATGCTCG CGATTATTTA AAAACACAAT TACAAAATAT GAAATGTGTA 2280
     AATGGAACTA CTGATGAAAA TTGTGAGTAT AAGTGTATGC ATAAAACCTC ATCCACAAAT 2340
40
     AGTGATATGC CCGAATCGTT GGACGAAAAG CCGGAAAAGG TCAAAGACAA GTGTAATTGT 2400
     GTACCTAATG AATGCAATGC ATTGAGTGTA AGTGGTAGCG GTTTTCCTGA TGGTCAAGCT 2460
     TACGTACGCG TGCATGCGAC GTCATAGCTC TTCTATAGTG TCACCTAAAT TCAATTCACT 2520 GGCCGTCGTT TTACAACGTC GTGACTGGGA AAACCTGGCG TTACCCAACT TAATCGCCTT 2580
     GCAGCACATC CCCCTTTCGC CAGCTG
45
     (2) INFORMATION FOR SEO ID NO:8:
           (i) SEQUENCE CHARACTERISTICS:
                (A) LENGTH: 921 amino acids
50
                (B) TYPE: amino acid
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(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

55

60

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Lys Leu Asn Ser Val Tyr Thr Ser Ile Gly Leu Phe Leu Asn Ser Leu

	Lys	Ser	Gly	Pro 20	Cys	Lys	Lys	Asp	Asn 25		Asn	Ala	Glu		Asn 0	Ile
			35					40	Phe	Lys			4	Asn 5	Cys	=
5		50					55		Cys			6	0			
	Thr 65	Lys	Gly	Lys	Сув	Asn 70	Gly	Ser	Asn	Gly	Lys 75	Lys	Asn	Gly	Asn	Asp 80
10					85				Asn	90	)				- 9	Asn 5
				100	) .				Asp 10	5				1	10	
•			115	i				12					1.	25		
15		130					13	5	Leu			14	40		_	
	145					150			Gln		155					160
20					165	5			Tyr	17	0				1	75.
				180	)				Thr 18	5				1	90	
25			195					20	-	•			2	05	-	
25		210					21!	5	Asn			22	20		-	
•	225					230			Met		235					240
30					245	;			Asp	25	0				2	55
				260					Ser 26	5				2'	70	
35			275					28					28	35		
		290					295	5	Ile Gly			30	00	_	_	
	305					310			Ile		315					320
40					325				Thr	33	0				3	35
				340					34! Ala	5				3.	50	
45			355			•		36					36	55	_	
		370					375	5	Lys			38	30			
	385					390			Arg		395					400
50	Arg				405					41	0 -				4	15
	Met			420					42	5				4:	30	
55			435	•				44					44	15		_
	Ala	450	•				455	5				46	0		-	_
	465 Tyr	Ъўā	Lys	Āāÿ	Ьÿā	470 Asii	Gly	Āāñ	Alā		475 Asp	Leu	Asp	Glu		480 Leu
60	Lys				485					49	0				. 4	95
	_	_		500					50! Leu	5				51	LO	
			515		_	_		520		-		-	52			-

		530					53	5				5	40		_	_
	Val 545	Phe	Tyr	Lys	Ile	Ser 550	Asn	Ser	Ile	Arg	Tyr 555	Arg	Lys	Ser	Trp	Trp 560
5	Glu	Thr	Asn	Gly	Pro 565	Val	Ile	Trp	Glu	Gly 57	Met	Leu	Cys	Ala		Ser 575
	Tyr	Asp	Thr	Ser 580	Leu	Asn	Asn	Val	Asn 58	Pro	Glu	Thr	His	_	Lys	Leu
10	Thr	Glu	Gly 595	Asn		Asn	Phe	Glu 60	Lys		Ile	Phe		Ser	90 Asp	Ser
	Ser	Thr 610	Thr		Ser	Lys	Phe 61	Ser		Arg	Pro		Phe 20	05 Leu	Arg	Trp
	Leu 625	Thr	Glu	Trp	Gly	Glu 630			Cys	Lys	Glu 635			Lys	Glu	_
15		Val	Leu	Leu	Ala 645	Lys	Cys	Lys	Asp	Cys 65	Asp	Val	Asp	Gly		640 Gly 55
	Lys	Суѕ	Asn	Gly 660	Lys	-	Val	Ala	Cys 66	Lys		Gln	Cys		Gln 70	Tyr
20	His	Ser	Trp 675	Ile	Gly	Ile	Trp	Ile 68	Asp		Tyr	Lys		Gln 85	Lys	Gly
	Arg	Tyr 690	Thr	Glu	Val	Lys	Lys 69	Ile 5	Pro	Leu	Tyr				Lys	Asp
	705		Asn			710					715					720
25			Lys		725	,				73	0				7	35
			His	740					74	5				7	50	
30			Lys 755					76	0				76	55		
		770	Asn				77	5				78	30		_	
35	785		Gly			790					795				_	800
			Lys		805					81	0				8	15
			Ser	820					82	5				8:	30	
40			B35					840	0				84	15		
		850	Cys				855	5				86	0			
AE	865		Ile			870					875					880
45			Leu		885					89	0				8	95
	Val			900					90	5	Lys	Met	Lys		Met 10	Lys
50	Lys	Met	Lys 915	Lys	Arg	Lys	Lys	920	_	e <sup>.</sup>				•		

# (2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 2101 base pairs
(B) TYPE: nucleic acid

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- 60 (ii) MOLECULE TYPE: DNA (genomic)
  - (iii) HYPOTHETICAL: NO

55

(vi) ORIGINAL SOURCE:

### (A) ORGANISM: Plasmodium falciparum

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

```
5
     GGAACAGGGT GATAATAAAG TAGGAGCCTG TGCTCCGTAT AGACGATTAC ATTTATGTGA 60
     TTATAATTTG GAATCTATAG ACACAACGTC GACGACGCAT AAGTTGTTGT TAGAGGTGTG 120
     TATGGCAGCA AAATACGAAG GAAACTCAAT AAATACACAT TATACACAAC ATCAACGAAC 180
     TAATGAGGAT TCTGCTTCCC AATTATGTAC TGTATTAGCA CGAAGTTTTG CAGATATAGG 240
     TGATATCGTA AGAGGAAAAG ATCTATATCT CGGTTATGAT AATAAAGAAA AAGAACAAAG 300
10
     AAAAAAATTA GAACAGAAAT TGAAAGATAT TTTCAAGAAA ATACATAAGG ACGTGATGAA 360
     GACGAATGGC GCACAAGAAC GCTACATAGA TGATGCCAAA GGAGGAGATT TTTTTCAATT 420
     AAGAGAAGAT TGGTGGACGT CGAATCGAGA AACAGTATGG AAAGCATTAA TATGTCATGC 480
     ACCAAAAGAA GCTAATTATT TTATAAAAAC AGCGTGTAAT GTAGGAAAAG GAACTAATGG 540
     TCAATGCCAT TGCATTGGTG GAGATGTTCC CACATATTTC GATTATGTGC CGCAGTATCT 600
15
     TTTGCAAAAA CAGTGTCGTG ATTACGAACA AAATTTATAT TGTAGTGGTA ATGGCTACGA 720
     TTGCACAAAA ACTATATATA AAAAAGGTAA ACTTGTTATA GGTGAACATT GTACAAACTG 780
     TTCTGTTTGG TGTCGTATGT ATGAAACTTG GATAGATAAC CAGAAAAAAG AATTTCTAAA 840
     ACAAAAAAGA AAATACGAAA CAGAAATATC AGGTGGTGGT AGTGGTAAGA GTCCTAAAAG 900
20
     GACAAAACGG GCTGCACGTA GTAGTAGTAG TAGTGATGAT AATGGGTATG AAAGTAAATT 960
     TTATAAAAAA CTGAAAGAAG TTGGCTACCA AGATGTCGAT AAATTTTTAA AAATATTAAA 1020
     CAAAGAAGGA ATATGTCAAA AACAACCTCA AGTAGGAAAT GAAAAAGCAG ATAATGTTGA 1080
     TTTTACTAAT GAAAAATATG TAAAAACATT TTCTCGTACA GAAATTTGTG AACCGTGCCC 1140
     ATGGTGTGGA TTGGAAAAAG GTGGTCCACC ATGGAAAGTT AAAGGTGACA AAACCTGCGG 1200
25
     AAGTGCAAAA ACAAAGACAT ACGATCCTAA AAATATTACC GATATACCAG TACTCTACCC 1260
     TGATAAATCA CAGCAAAATA TACTAAAAAA ATATAAAAAT TTTTGTGAAA AAGGTGCACC 1320
     TGGTGGTGGT CAAATTAAAA AATGGCAATG TTATTATGAT GAACATAGGC CTAGTAGTAA 1380
     AAATAATAAT AATTGTGTAG AAGGAACATG GGACAAGTTT ACACAAGGTA AACAAACCGT 1440
     TAAGTCCTAT AATGTTTTTT TTTGGGATTG GGTTCATGAT ATGTTACACG ATTCTGTAGA 1500
30
     GTGGAAGACA GAACTTAGTA AGTGTATAAA TAATAACACT AATGGCAACA CATGTAGAAA 1560
     CAATAATAAA TGTAAAACAG ATTGTGGTTG TTTTCAAAAA TGGGTTGAAA AAAAACAACA 1620
     AGAATGGATG GCAATAAAAG ACCATTTTGG AAAGCAAACA GATATTGTCC AACAAAAAGG 1680
     TCTTATCGTA TTTAGTCCCT ATGGAGTTCT TGACCTTGTT TTGAAGGGCG GTAATCTGTT 1740
     GCAAAATATT AAAGATGTTC ATGGAGATAC AGATGACATA AAACACATTA AGAAACTGTT 1800
35
    GGATGAGGAA GACGCAGTAG CAGTTGTTCT TGGTGGCAAG GACAATACCA CAATTGATAA 1860
    ATTACTACAA CACGAAAAAG AACAAGCAGA ACAATGCAAA CAAAAGCAGG AAGAATGCGA 1920
     GAAAAAAGCA CAACAAGAAA GTCGTGGTCG CTCCGCCGAA ACCCGCGAAG ACGAAAGGAC 1980
     ACAACAACCT GCTGATAGTG CCGGCGAAGT CGAAGAAGAA GAAGACGACG ACGACTACGA 2040
    CGAAGACGAC GAAGATGACG ACGTAGTCCA GGACGTAGAT GTAAGTGAAA TAAGAGGTCC 2100
40
                                                                     2101
```

#### (2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 700 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- 50 (ii) MOLECULE TYPE: protein

45

- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
   (A) ORGANISM: Plasmodium falciparum
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:
- Glu Glu Gly Asp Asn Lys Val Gly Ala Cys Ala Pro Tyr Arg Arg Leu

  1 5 10 15

  His Leu Cys Asp Tyr Asn Leu Glu Ser Ile Asp Thr Thr Ser Thr Thr

  20 25 30

  His Lys Leu Leu Leu Glu Val Cys Met Ala Ala Lys Tyr Glu Gly Asn

  35 40

	Ser	Ile 50	Asn	Thr	His	Tyr	Thr 55	Gln	His	Gln	Arg	Thr 60		Glu	Asp	Ser
	Ala 65	Ser	Gln	Leu	Cys	Thr 70	Val	Leu	Ala	Arg	Ser 75			Asp	Ile	Gly 80
5	Asp	Ile	Val	Arg	Gly 85	Lys	Asp	Leu	Tyr	Leu 90	Gly	Tyr	Asp	Asn	Lys 95	
		Glu		100	1				10	5				11	.0	-
10	Lys	Ile	His 115		Asp	Val	Met	Lys 120		Asn	Gly	Ala	Gln 12	. —	Arg	Tyr
		Asp 130				•	135	5				14	0		_	_
	145	Thr				150					155			_	•	160
15		Lys			165	•				17	0				17	<b>'</b> 5
		Thr		180					18	5				. 19	0	_
20		Asp	195					200	)	_			20	5 -		
		Phe 210					215	5	-			22	0		_	
0.5	225	Arg				230				_	235	_		_	_	240
25		Thr			245	5				25	0			_	25	5
		Thr		260					26	5				27	0	
30		Gln	275					280	)				28	5		
		Ser 290					295	5				30	0			
35	305	Arg				310	•				315				_	320
33		Lys			325	;				33	0			_	33	5
		Ile		340					345	5		•		35	0	
40		Glu	355					360	)	•			36	5		_
		Phe 370 Lys		•			375	•				38	0		_	
45	385	Ala				390					395					400
10		Leu			405	;				41	0				41	.5
•		Phe		420					425	5			_	43	0	_
50		Cys	435					440	)		•		44	5		
		450 Val					455	,				46	0			
55	465	Ser				470					475	_	_			480
		Ser			485					49	0				49	5
		Asn		500					505	5	_	_		51	0	
60		Cys	515			_	_	520	)		_	•	52	5	_	-
	Ile	530					535	,		_		54	0	-		
	545	ay o	υoρ		- 11C	550	ay s	JIII	1111	പാവ	555	val	GIII	GIII	пyъ	560

Leu Ile Val Phe Ser Pro Tyr Gly Val Leu Asp Leu Val Leu Lys Gly 565 570 Gly Asn Leu Leu Gln Asn Ile Lys Asp Val His Gly Asp Thr Asp Asp 580 585 5 Ile Lys His Ile Lys Lys Leu Leu Asp Glu Glu Asp Ala Val Ala Val 600 Val Leu Gly Gly Lys Asp Asn Thr Thr Ile Asp Lys Leu Leu Gln His 615 Glu Lys Glu Gln Ala Glu Gln Cys Lys Gln Lys Gln Glu Glu Cys Glu 10 630 635 Lys Lys Ala Gln Gln Glu Ser Arg Gly Arg Ser Ala Glu Thr Arg Glu 645 650 Asp Glu Arg Thr Gln Gln Pro Ala Asp Ser Ala Gly Glu Val Glu Glu 665 Glu Glu Asp Asp Asp Asp Tyr Asp Glu Asp Asp Glu Asp Asp Val 15 680 Val Gln Asp Val Asp Val Ser Glu Ile Arg Gly Pro 690 695 20 (2) INFORMATION FOR SEO ID NO:11: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 8220 base pairs (B) TYPE: nucleic acid 25 (C) STRANDEDNESS: single (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: DNA (genomic)
- 30 (iii) HYPOTHETICAL: NO
  - (vi) ORIGINAL SOURCE: (A) ORGANISM: Plasmodium falciparum
- 35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

AAAAATGGGG CCCAAGGAGG CTGCAGGTGG GGATGATATT GAGGATGAAA GTGCCAAACA 60 TATGTTTGAT AGGATAGGAA AAGATGTGTA CGATAAAGTA AAAGAGGAAG CTAAAGAACG 120 TGGTAAAGGC TTGCAAGGAC GTTTGTCAGA AGCAAAATTT GAGAAAAATG AAAGCGATCC 180 40 ACAAACACCA GAAGATCCAT GCGATCTTGA TCATAAATAT CATACAAATG TAACTACTAA 240 TGTAATTAAT CCGTGCGCTG ATAGATCTGA CGTGCGTTTT TCCGATGAAT ATGGAGGTCA 300 ATGTACACAT AATAGAATAA AAGATAGTCA ACAGGGTGAT AATAAAGGTG CATGTGCTCC 360 ATATAGGCGA TTGCATGTAT GCGATCAAAA TTTAGAACAG ATAGAGCCTA TAAAAATAAC 420 AAATACTCAT AATTTATTGG TAGATGTGTG TATGGCAGCA AAATTTGAAG GACAATCAAT 480 45 AACACAAGAT TATCCAAAAT ATCAAGCAAC ATATGGTGAT TCTCCTTCTC AAATATGTAC 540 TATGCTGGCA CGAAGTTTTG CGGACATAGG GGACATTGTC AGAGGAAGAG ATTTGTATTT 600 AGGTAATCCA CAAGAAATAA AACAAAGACA ACAATTAGAA AATAATTTGA AAACAATTTT 660 CGGGAAAATA TATGAAAAAT TGAATGGCGC AGAAGCACGC TACGGAAATG ATCCGGAATT 720 TTTTAAATTA CGAGAAGATT GGTGGACTGC TAATCGAGAA ACAGTATGGA AAGCCATCAC 780 50 ATGTAACGCT TGGGGTAATA CATATTTTCA TGCAACGTGC AATAGAGGAG AACGAACTAA 840 AGGTTACTGC CGGTGTAACG ACGACCAAGT TCCCACATAT TTTGATTATG TGCCGCAGTA 900 TCTTCGCTGG TTCGAGGAAT GGGCAGAAGA TTTTTGTAGG AAAAAAAAA AAAAAATAAA 960 AGATGTTAAA AGAAATTGTC GTGGAAAAGA TAAAGAGGAT AAGGATCGAT ATTGTAGCCG 1020 TAATGGCTAC GATTGCGAAA AAACTAAACG AGCGATTGGT AAGTTGCGTT ATGGTAAGCA 1080 55 ATGCATTAGC TGTTTGTATG CATGTAATCC TTACGTTGAT TGGATAAATA ACCAAAAAGA 1140 ACAATTTGAC AAACAGAAAA AAAAATATGA TGAAGAAATA AAAAAATATG AAAATGGAGC 1200 ATCAGGTGGT AGTAGGCAAA AACGGGATGC AGGTGGTACA ACTACTACTA ATTATGATGG 1260 ATATGAAAAA AAATTTTATG ACGAACTTAA TAAAAGTGAA TATAGAACCG TTGATAAATT 1320 TITGGAAAAA TTAAGTAATG AAGAAATTG CACAAAAGTT AAAGACGAAG AAGGAGGAAC 1360 60 AATTGATTTT AAAAACGTTA ATAGTGATAG TACTAGTGGT GCTAGTGGCA CTAATGTTGA 1440 AAGTCAAGGA ACATTTTATC GTTCAAAATA TTGCCAACCC TGCCCTTATT GTGGAGTGAA 1500 AAAGGTAAAT AATGGTGGTA GTAGTAATGA ATGGGAAGAG AAAAATAATG GCAAGTGCAA 1560 GAGTGGAAAA CTTTATGAGC CTAAACCCGA CAAAGAAGGT ACTACTATTA CAATCCTTAA 1620 AAGTGGTAAA GGACATGATG ATATTGAAGA AAAATTAAAC AAATTTTGTG ATGAAAAAA 1680

	TGGTGATACA	ATAAATAGTG	GTGGTAGTGG	TACGGGTGGT	AGTGGTGGTG	GTAACAGTGG	1740
	TAGACAGGAA	TTGTATGAAG	AATGGAAATG	TTATAAAGGT	GAAGATGTAG	TGAAAGTTGG	1800
	ACACGATGAG	GATGACGAGG	AGGATTATGA	AAATGTAAAA	<b>AATGCAGGCG</b>	GATTATGTAT	1860
	ATTAAAAAAC	CAAAAAAAGA	ATAAAGAAGA	AGGTGGAAAT	ACGTCTGAAA	AGGAGCCTGA	1920
5	TGAAATCCAA	<b>AAGACATTCA</b>	ATCCTTTTTT	TTACTATTGG	GTTGCACATA	TGTTAAAAGA	1990
	TTCCATACAT	TGGAAAAAA	AACTTCAGAG	ATCTTTACAA	AATGGTAACA	CANTANANTO	2040
	TGGAAACAAT	AAATGTAATA	ATGATTGTGA	ልተርተሞሞተልልል	AGATGGATTA	CACAAAAAA	2100
	AGACGAATGG	GGGAAAATAG	TACAACATTT	TAAAACCCAA	AATATTAAAG	CTACACCACCA	2100
	ТАСТСАСААТ	ACCCCAGAAT	ጥአ አጥር ርር አጥጥ	TOATOACCO	TATGTTCTTC	STAGAGGAGG	<b>7100</b>
10	CCAACAACAA	TOCOCHONN	CCCATTCCCATT	ACACCOMMOC	GAAGAAAAAT	AATACAATTT	2220
	TOTCCATCCA	CACCACCCAC	OCCALICOM	AGACGCTICC	GAAGAAAAT	CCGAAAATAG	2280
	CANTANTONA	CARCCARC	MICCINAA	ACACCTTCGC	GAAATCATTG	AAAGTGAAGA	2340
	CAMIMAICAA	GAAGCATCIG	TIGGIGGIGG	CGTCACTGAA	СААААААТА	TAATGGATAA	2400
	ATTGCTCAAC	TACGAAAAAG	ACGAAGCCGA	TTTATGCCTA	GAAATTCACG	AAGATGAGGA	2460
15	AGAGGAAAAA	GAAAAAGGAG	ACGGAAACGA	ATGTATCGAA	GAGGGCGAAA	ATTTTCGTTA	2520
10	TAATCCATGT	AGTGGCGAAA	GIGGIAACAA	ACGATACCCC	GTTCTTGCGA	ACAAAGTAGC	2580
	GTATCAAATG	CATCACAAGG	CAAAGACACA	ATTGGCTAGT	CGTGCTGGTA	GAAGTGCGTT	2640
	GAGAGGTGAT	ATATCCTTAG	CGCAATTTAA	AAATGGTCGT	AACGGAAGTA	CATTGAAAGG	2700
	ACAAATTTGC	AAAATTAACG	AAAACTATTC	CAATGATAGT	CGTGGTAATA	GTGGTGGACC	2760
	ATGTACAGGC	AAAGATGGAG	ATCACGGAGG	<b>TGTGCGCATG</b>	<b>AGAATAGGAA</b>	CGGAATGGTC	2820
20	AAATATTGAA	<b>GGAAAAAAAC</b>	<b>AAACGTCATA</b>	CAAAAACGTC	TTTTTACCTC	CCCGACGAGA	2880
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	GGCTAGCCAC	<b>TCATTATTGG</b>	GAGATGTTCA	GCTCGCAGCA	AAAACTGATG	CAGCTGAGAT	3000
	AATAAAACGC	TATAAAGATC	AAAATAATAT	ACAACTAACT	GATCCAATAC	AACAAAAACA	3060
	CCAGGAGGCT	ATGTGTCGAG	CTGTACGTTA	TACTTTTCCC	CATTTACCAC	<b>አሮኔምታታታታ</b> ርና	3120
25	AGGAAGAGAT	ATGTGGGATG	AGGATAAGAG	CTCAACAGAC	ATGGAAACAC	CTTTCATAC	3100
	CGTATTTAAA	AACATTAAAG	AAAAACATGA	TCCAATCAAA	CACAACCCTA	A A TATATA COCC	3240
	TGATGAAAGC	AAAAAGCCCG	CATATAAAAA	ATTACCACCA	CATTCCTCC	MAINIACCOG	3240
	ACATCAAGTG	TGGAGAGCCA	TGAAATGCGC	ATTACCACCA	ATTIGGIGG	CHAMAIAG	3300
	AGTTGACGAT	TATATCCCCC	A A COMMITTO COC	CTCCATCACT	WICHINIGIC	LIGGIAIGCC	3360
30	TAAAGCGCAA	TCACAGGAGT	AUCOLLINCA	CIGGAIGACI	GWWIGGGCIG	AAIGGIAIIG	3420
00	GGGTGATGGA	1 CACAGGAGI	WIGWCWWGII	AAAAAAAAA C	TGTGCAGATT	GTATGAGTAA	3480
	TANATATA	CACCAAATAC	MAGGIGATGI	CGAI IGIGGA	AAGIGCAAAG	CAGCATGTGA	3540
	TAAATATAAA	TACCTACA AC	MAMAMIGGAA	TGAACAATGG	AGAAAAATAT	CAGATAAATA	3600
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33	TATTGCCGCA	CGIGITCIIG	TTAAACGTGC	TGCTGGTAGT	CCCACTGAGA	TCGCCGCCGC	3780
	CGCCCCGATC	ACCCCCTACA	GTACTGCTGC	CGGATATATA	CACCAGGAAA	TAGGATATGG	3840
	GGGGTGCCAG	GAACAAACAC	AATTTTGTGA	AAAAAAACAT	GGTGCAACAT	CAACTAGTAC	3900
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	ATGTAATCCA	AAAGAGAGTT	ATCCTGATTG	GGATTGCAAA	AACAATATTG	ACATTAGTCA	4140
	TGATGGTGCT	TGTATGCCTC	CAAGGAGACA	AAAACTATGT	TTATATTATA	TAGCACATGA	4200
	GAGTCAAACA	GAAAATATAA	AAACAGACGA	TAATTTGAAA	GATGCTTTTA	TTAAAACTGC	4260
	AGCAGCAGAA	ACTITICTIT	CATGGCAATA	TTATAAGAGT	AAGAATGATA	GTGAAGCTAA	4320
45	AATATTAGAT	AGAGGCCTTA	TTCCATCCCA	ATTTTTAAGA	TCCATGATGT	ACACGTTTGG	4380
•	AGATTATAGA	GATATATGTT	TGAACACAGA	TATATCTAAA	AAACAAAATG	ATGTAGCTAA	4440
	GGCAAAAGAT	AAAATAGGTA	AATTTTTCTC	AAAAGATGGC	AGCAAATCTC	CTAGTGGCTT	4500
	ATCACGCCAA	GAATGGTGGA	AAACAAATGG	TCCAGAGATT	TGGAAAGGAA	TGTTATGTGC	4560
	CTTAACAAAA	TACGTCACAG	ATACCGATAA	CAAAAGAAAA	ATCAAAAACG	ACTACTCATA	4620
50	CGATAAAGTC	AACCAATCCC	AAAATGGCAA	CCCTTCCCTT	GAAGAGTTTG	CTGCTAAACC	4680
	TCAATTTCTA	CGTTGGATGA	TCGAATGGGG	AGAAGAGTTT	TGTGCTGAAC	GTCAGAAGAA	4740
	GGAAAATATC	ATAAAAGATG	CATGTAATGA	AATAAATTCT	ACACAACAGT	CTAATCATCC	4800
	GAAACATCGT	TGTAATCAAG	CATGTAGAGC	ATATCAAGAA	TATCTTCAAA	AKKKKKKKTK	4000
	AGAATTTTCG	GCACAAACAA	ልጥል ል ርጥጥጥርጥ	ייייייייייייייייייייייייייייייייייייי	እአጥሮጥጥሮአሮሮ	CCCS SCSECC	4000
55	AGAATATAAA	CCATATCAAT	ATAAACIIIGI	CCTACAACCC	AMIGIICAGC	CCCWWGWICC	4920
00	ACTGCAAAAA	TOTAL TOTAL	ATMANGACGG	TO I ACAMCCO	ATACAGGGGA	AIGAGIAIII	4980
	MCIGCHAMMA	IGIGALAAIA	AIMMAIGIIC	COLUMN	GGAAATGTAC	TITCCGTCTC	5040
	TCCAAAAGAA	MAACCI I'I'IG	GAAAATATGC	CCATAAATAT	CCTGAGAAAT	GIGATIGTTA	5100
	TCAAGGAAAA	CATGTACCTA	GCATACCACC	TCCCCCCCA	CCTGTACAAC	CACAACCGGA	5160
20	AGCACCAACA						
60	CAATTTTTCC						
	TATACCAAGT	GACACAAAAA	GTGGTGCTGG	TGCCACCACC	GGCAAAAGTG	GTAGTGATAG	5340
	TGGTAGTATT						
	GGCTACCGCG						
	GCGCAATGCG	TTCATCCAAT	CTGCTGCAAT	AGAGACTTTT	TTCTTATGGG	ATAGATATAA	<b>5520</b> .

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AGAAGAGAAA AAACCACAGG GTGATGGGTC ACAACAAGCA CTATCACAAC TAACCAGTAC 5580
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     CGATITITIG AGATTAATGT TCTATACATT AGGAGATTAT AGGGATATTT TAGTACACGG 5700
     TGGTAACACA AGTGACAGTG GTAACACAAA TGGTAGTAAC AACAACAATA TTGTGCTTGA 5760
     AGCGAGTGGT AACAAGGAGG ACATGCAAAA AATACAAGAG AAAATAGAAC AAATTCTCCC 5820
     AAAAAATGGT GGCACACCTC TTGTCCCAAA ATCTAGTGCC CAAACACCTG ATAAATGGTG 5880
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     GAACCCTGAC ACCAGTGCAA GAGGCGACGA AAACAAAATA GAAAAGGATG ATGAAGTGTA 6000
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     AAGACAATAT AGTGGGGATG GCGAAGCGTG TAATGAGATG CTTCCAAAAA ACGATGGAAC 6360
     TGTTCCGGAT TTAGAAAAGC CGAGTTGTGC CAAACCTTGT AGTTCTTATA GAAAATGGAT 6420
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     GCGTGTCAGT GCTGATAGTA AAAGTGGATT TAATGGTGAT GGTTTAGAGA ATGCTTGTAG 6840
     AGGTGCTGGT ATCTTTGAAG GTATTAGAAA AGATGAATGG AAATGTCGTA ATGTATGTGG 6900
     TTATGTTGTA TGTAAACCGG AAAACGTTAA TGGGGAAGCA AAGGGAAAAC ACATTATACA 6960
25
     AATTAGAGCA CTGGTTAAAC GTTGGGTAGA ATATTTTTTT GAAGATTATA ATAAAATAAA 7020
     ACATAAAATT TCACATCGCA TAAAAAATGG TGAAATATCT CCATGTATAA AAAATTGTGT 7080
     AGAAAAATGG GTAGATCAGA AAAGAAAAGA ATGGAAGGAA ATTACTGAAC GTTTCAAAGA 7140
     TCAATATAAA AATGACAATT CAGATGATGA CAATGTGAGA AGTTTTTTGG AGACCTTGAT 7200
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     TCTAACCGCC CTGGTGACCT CCACCCTCGC CTGGAGCGTT GGCATCGGTT TTGCTACATT 7800
     CACTTATTTT TATCTAAAGG TAAATGGAAG TATATATATG GGGATGTGGA TGTATGTGGA 7860
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     TGTATGTGAA TGTATGTGGA TGTATGTGGA TGTATGTGTAT GGATATGTAT 7920
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     ΑΑΑΑGΑΑΤΑ ΤΑΑΑΑΑCAAA ΤΙΤΑΤΤΑΑΑΑ ΤGAAAAAAG AAAAATGAAA ΤΑΤΑΑΑΑΑΑ 8160
45
     ΑΑΤΤΤΑΤΤΑΑ ΑΑΤΑΑΑΑΑΑΑ ΑΑΑΑΑΑΑΑΑΑ ΑΑΑΑGGAGAA ΑΑΑΤΤΤΤΤΤΑ ΑΑΑΑΑΤΑΑΤΑ 8220
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# (2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2710 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO

50

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60

- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Asn Val Met Val Glu Leu Ala Lys Met Gly Pro Lys Glu Ala Ala Gly

	1				5					10					19	5
				Ile 20					25					30	)	
5	Gly	Lys	Asp 35	Val	Tyr	Asp	Lys	Val	Lys	Glu	Glu	Ala	Lys 45		Arg	Gly
		50		Gln			55					60	)	_		
	Ser 65	Asp	Pro	Gln	Thr	Pro 70	Glu	Asp	Pro	Cys	Asp 75	Leu	Asp	His	Lys	Tyr 80
10	His	Thr	Asn	Val	Thr 85	Thr	Asn	Val	Ile	Asn 90	Pro	Сув	Ala	Asp	Arg	Ser
	Asp	Val	Arg	Phe 100	Ser	<b>A</b> sp	Glu	Tyr	Gly 10		Gln	Сув	Thr	His 11	_	Arg
15	Ile	Lys	Asp 115	Ser	Gln	Gln	Gly	Asp 12		Lys	Gly	Ala	Cys	Ala 5	Pro	Tyr
	Arg	Arg 130	Leu	His	Val	Сув	Asp 135		Asn	Leu	Glu	Gln 14	_	Glu	Pro	Ile
	145			Asn		150					155					160
20				Gly	165	5				17	0				17	'5
				Asp 180					18!	5				19	0	
25			195					200	0				20	5		_
		210		Glu			215	5				22	0			_
20	225			Gly		230					235					240
30				Asp	245	;				25	0		_	_	Ž5	5
				Glu 260					265	5				27	0 ~	
35			275					280	)				28	5	_	_
		290		Cys			295	5				30	0		_	
<b>1</b> 0	Pro 305				-	310					315		_		-	320
<b>70</b> .				Lys	325					336	)				33	5
				Asp 340					345	5				35	0	_
<b>1</b> 5	Glu		355					360	)				36	5		_
		370		Leu			375					386	0			
50	385			Gln		390					395					400
	Lys				405					410	)				41	5
	Ala			420					425	5				43	o ¯	
55	Tyr Glu		435					440	)				44	5		
		450					455					46	0	_		
	Gly 465	Gry	IIII	TIE	wsb	470	пÀг	ASII	vaı		5er 475	Asp	ser	Thr		G1Y 480
50	Ala	Ser	Gly	Thr	Asn 485	Val	Glu	Ser	Gln		Thr	Phe	Tyr	Arg		Lys
	Tyr	Суѕ	Gln	Pro 500			Tyr	Cys	Gly 505	Val		Lys	Val	Asn 51	Asn	
	Gly	Ser	Ser		Glu	Trp	Glu	Glu			Asn	Gly	Lys			Ser

		515		520		525
•	53	Ų	53	s Pro Asp Ly	54(	Thr Thr Ile Thr
5	545		550	• •	p Ile Glu 555	Glu Lys Leu Asn 560
	,	5	65	5	570	Ser Gly Gly Ser
		580		585		Gln Glu Leu Tyr 590
10		595		600		Lys Val Gly His 605
	610	)	61	.5	620	Asn Ala Gly Gly
15	625		630		635	Glu Gly Gly Asn 640
		64	45	6	50	Phe Asn Pro Phe 655
20		660		665		Ile His Trp Lys 670
20		675		680		Ile Lys Cys Gly 685
	690	)	69	5	700	
25	705		710		715	Phe Lys Thr Gln 720
		72	25	7	30	Glu Leu Ile Pro 735
20		740		745		Glu Glu Phe Leu 750
30		755		760		Glu Asn Ser Leu 765
	· 770		77	5	780	Slu Ile Ile Glu
<b>35</b>	785		790		795	Sly Val Thr Glu 800
		80	)5	8	10	ys Asp Glu Ala 815
40	-	820		825		Glu Lys Glu Lys 830
<del>4</del> 0		835		840		Phe Arg Tyr Asn 845
	850		85	5	860	al Leu Ala Asn
45	865		870		875	In Leu Ala Ser 880
		88	35	8:	90	eu Ala Gln Phe 895
En		900		905		le Cys Lys Ile 910
50	•	915		920		ly Gly Pro Cys 925
	930		93	5	940	arg Ile Gly Thr
55	945		950		955	'yr Lys Asn Val 960
		96	55	9'	70	sn Leu Glu Asn 975
60	ьеи Asp	Val Gly Ser	r Val Thr	Lys Asn Asp 985	Lys Ala S	er His Ser Leu 990
60	Leu Gly		ı Leu Ala		Asp Ala A	la Glu Ile Ile
	Lys Arg	995 Tyr Lys Asp	Gln Asn	1000 Asn Ile Glr	Leu Thr A	1005 sp Pro Ile Gln
	101	υ	10:	15	1020	

	<b>~</b> 1	•	•	~~.	~-			_	_							
	102	Lys 5	Asp	Gin	GIU	103		Cys	Arg	Ala	Val 103		Tyr	Ser	Phe	Ala
			Gly	Asp	Ile 104	Ile	Arg	Gly	Arg				Asp	Glu		
5	Ser	Ser	Thr	Asp	Met		Thr	Arg	Leu 10	Ile		Val	Phe		Asn 70	Ile
			107	5			Ile	10	Asp 80	Asn			10	Thr 85	Gly	-
10	Glu	Ser		Lys	Pro	Ala	Tyr		Lys	Leu	Arg			Trp	Trp	Glu
10	Ala 110	Asn		His	Gln	Val	109 Trp		Ala	Met		Cys	00 Ala	Thr	Lys	<del>-</del> -
·			Суз	Pro	Gly 112	Met	Pro	Val	Asp	Asp 11			Pro	Gln		1120 Leu .35
15	Arg	Trp	Met	Thr 114	Glu		Ala	Glu	Trp	Tyr		Lys	Ala		Ser 50	Gln
			115	5			Lys	116	50				11	Ser 65	Lys	_
20		117	0				Gly 117	15				11	80			
	1189	5				1190		•			1195	5				1200
					120	5	Tyr.			12	10				12	15
25				122	0		Arg		122	25				12	30	_
			123	5			Phe	124	10				12	45		
30		125	0				Lys 125	55				12	60			
	1265	5				1270					1275	;		-	-	1280
0.5	His				128	5				129	90				12	95
35	Glu			130	0				130	05				13	10	-
			131	5			Pro	132	20				13	25	_	_
40		1330	0				Thr 133	5				13	40			
	Val 1345	5				1350	)				1355	•			_	1360
45	Gly				136	5				13	70			_	13	75
45		•		138	0		Ile		138	35				13	90	
	Pro		139	5				140	00				14	05		
50	Gln	1410	0				141	.5			•	14	20			
	Lys 1425	5 .				1430	)				1435	,		_	_	1440
	Lys				144	5				145	50				14	55
55	Gln			146	0				146	55				14	70	í
	Cys		147	5				148	30			_	14	85	-	
60	гуѕ	1490	)				149	5				15	ser 00	rys		
	Ser 1505	5				1510	)				1515		_			1520
	Trp	Lys	Gly	Met	Leu 152		Ala	Leu	Thr		Tyr		Thr	Asp	Thr	

•	Asn	Lys	Arg	Lys 154		Lys	Asn		Tyr 15		Tyr	Asp	Lys		Asn 550	Gln
_			155	55				15	60				15	565	Pro	
5		157	0				15	75				15	80		Glu	
	158	5				159	0				159	5			Asn	1600
10					160	)5				16	10				Cys 16	515
				162	20				16	25				16	Gly	
•-			163	5				16	40				16	45	Pro	
15		165	0				165	55				16	60		Gly	
·	166	5				1670	)				1679	5		_	Met	1680
20					168	35				16	90				Lys 16	95
				170	0				17	05				17	His 110	
25			171	5				172	20				17	25	Glu	
		173	0				173	35				17	40		Phe Lys	_
•	1749	5				1750	)				1755	5				1760
30					176	5				17	70		_			75
				178	0				17	85				17	90 Trp	
35			179	5				180	00				18	05	Arg	
		181	0				181	.5				18	20		Thr	
	1825	5				1830	)				1835	5				1840
40					184	5				18	50			-		55
				186	0				186	65				18	70 Pro	
<b>4</b> 5	Phe	Leu	187 Arg	5 Leu	Met	Phe	Tyr	188 Thr	30 Leu			Tyr	18 Arg	85	Ile	
	Val	His		Gly		Thr	Ser			Gly	Asn	19 Thr		Gly	Ser	Asn
50	1905 Asn	-	Asn	Ile				Ala	Ser				Glu	Asp	Met	1920 Gln
50	Lys	Ile	Gln				Glu	Gln				Lys	Asn	_	Gly	35 Thr
	Pro	Leu				Ser	Ser				Pro	Asp	_	Trp	50 Trp	Asn
55	Glu				Ser	Ile				Met	Ile		Ala	65 Leu	Thr	Tyr
				Asn	Pro			_	Ala	Arg				Asn	Lys	
30	1985 Glu		Acp	Asp				Clu	Lys				Ser	Thr	Ala	
J <b>U</b>	Lys	His	Gly				Thr	Pro				Tyr	Lys		Gln	Tyr
	Asp		Glu 203			Lys	Leu					Gly	Ala 20	Lys	30 Thr	Pro
			40.0					40	··				2.13	<b></b>		

	Ser	Ala	Ser	Ser	Asp	Thr			Leu	Ser	Asp			Leu	Arg	Pro
	Pro	205 Tyr		Arg	Tyr	Leu	20! Glu		Trp	Gly		Asn	60 Phe	Cys	Lys	Lys
5	206 Arg		His	Lys	Leu	207 Ala		Ile	Lys			Cys	Lys	Val		2080 Glu
	Asn	Gly	Gly	Gly 210	200 Ser		Arg	Gly	Gly 21	Ile	90 Thr	Arg	Gln		Ser	95 Gly
10	Asp	Gly	Glu 211	Ala		Asn	Glu	Met 21	Leu	Pro	Lys	Asn		Gly .25	10 Thr	Val
	Pro	Asp 213	Leu		Lys	Pro	Ser 213	Cys		Lys	Pro		Ser 40	Ser	Tyr	Arg
	214	5				215	0				Glu 2155	Lys	Gln		_	2160
15					216	55				21					21	.75
				218	0				21	85	Ser		•	21	90	
20			219	5				220	00		Asn		22	05	-	
		221	0				221	L <b>5</b>			Thr	22	20	_	_	
25	222	5				223	0				Asp 2235	;		_		2240
20					224	15				22	Ile 50 Met	_			22	55
				226	.0				22	65				22	70	
30			227	5				228	30		Glu		22	85		
		229	0				229	95			Glu	23	00			
0.5	230	5			•	2310	)				Asn 2315			-		2320
35					232	25				23					23	35
				234	0				234	15	Lys			23	50	
40	Arg		235	5				23€	50				23	65		
		237	0				237	<b>'</b> 5			Lys	23	80			
45	2385	5				2390	)				Asp 2395					2400
					240	5				24:	Thr				24	15
	1			242	0				242	25	Asn			24	30	
50			243	5				244	10		Glu		24	45		
		245	D				245	5			Ile	24	60			_
	2465	5				2470	)				Ser 2475					2480
55					248	5				249					24	95
	Ala	Lys	Lys	Asn 250		Met	Pro	Lys	11e 250		Glu	Asn	Val	Leu 25	-	Thr
60	Alā	Glñ	Gln 251	Glu		Glu		Ğ1у 252	Суs		Pro	AĪa ,	GIu 25:	Asn		Glu
		253	0				Ser 253	Gly 5	Lys		Thr	25	Glu 40	Gln		
	Val 2545		Lys	Pro		Glu 2550		Ala	Val		Glu 2555		Pro	Pro		Pro 2560

30

Pro Gln Glu Lys Ala Pro Ala Pro Ile Pro Gln Pro Gln Pro Pro Thr 2565 2570 Pro Pro Thr Gln Leu Leu Asp Asn Pro His Val Leu Thr Ala Leu Val 2580 2585 2590 Thr Ser Thr Leu Ala Trp Ser Val Gly Ile Gly Phe Ala Thr Phe Thr 5 2595 2600 2605 Tyr Phe Tyr Leu Lys Val Asn Gly Ser Ile Tyr Met Gly Met Trp Met 2610 2615 Tyr Val Asp Val Cys Glu Cys Met Trp Met Tyr Val Asp Val Cys Gly 10 2625 2630 2635 2640 Cys Val Leu Trp Ile Cys Ile Cys Asp Tyr Val Trp Ile Tyr 2645 2650 2655 Ile Tyr Ile Cys Leu Cys Ile Cys Val Phe Gly Tyr Ile Tyr Val Tyr 2660 2665 15 Val Tyr Asp Phe Leu Tyr Met Tyr Leu Trp Val Lys Asp Ile Tyr Ile 2675 2680 Trp Met Tyr Leu Tyr Val Phe Tyr Ile Tyr Ile Leu Tyr Ile Cys Ile 2700 Tyr Ile Lys Lys Glu Ile 20

- (2) INFORMATION FOR SEQ ID NO:13:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 19124 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

35 ATCCTTCTAT TTTCGATTTT TTCATTTTTT TCCAGTATTA ATTTATTTAT TTATTTGTGA 120 TATTTTATAA TATATTATTT AAATGTGTAT TTATATATGT GTTTTATTTT TGTTATTAAT 180 40 TTGAATAATC CGAGCGAAAA AAAATATATA ATCTCATATA AAAATTATTT ATAATACAAT 240 ATTATATAGT TTCCTATTAA AATAAATTAA TATAATATAC AATAATATTT CTTGTTATTT 300 TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTTAT TCCTTTTAA TTTCTTAATT 360 CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAA AAAAAAAAA 420 AAAAAAAAA ATTTATTATA ATATAATAAA AAATATAAAG ACATACGTTC ACTTATTATT 480 45 ATAAATGATT TATTACGATT AAAACATATT GAGATTATAA TAATATAATT TAACATAGAA 540 AGAGTTAAGA ATACATTTTT TTTTTTTTT TGATATGTAA TTCAACATAT ATATATATAT 600 ATATCTTTTT AATTTAATTA AATAAAATTC CTTATTATTC ATATTGTTTC TTTTATCACA 660 TGTGAAATAT TAAAAATAAT TTTCGATTTT ATCGATATAT TTATGTCGTT TATATACTTA 720 TATAGGTCTT TATAACTATT GATTAATAGA AGGTAATAGC CTAATAATAT AAATACTCGT 780 50 ATTTATAAAT TCATTTATAT ATTTCAAATA TATTTCGATG GTTTATTTTC AAATACAATT 840 AATTAGATTT CTTAAATATT TCTTCATTTA TTCATTTTA TAGCATATAC ATGCACATTA 900 TTTCACACAA CATTTAAGTT GTCATAATGT AACACATTAA ATAATATTT ACTTATATAT 1020 ATATAATTAT TAATTATATA TTAAATAAAA ATGTATTATC GCCTGTATTA TCATAGTATA 1080 55 TATAATGTTG TATAACGCTT CAAAATATAT ATAATAATAT AATTAAAAAT ATATATATAG 1140 TAATTAATTA TTTTGTTATG TTATGTAATA ATGCAATTAA TATAAGATAA AATTCTATAG 1200 AATATTATAA TATGTAAATT ATTAATAAAA TATATTTGTA TAACATACAA GACTAAAGAA 1320 AACTATACAA TCTGGTATCT AATAGTATAT ATATATAATA TCTTTTTTAT TTAATTGTTC 1380 GATTTAGTAT TTTAATAATA AATAAATCTT TTAAAAAACT TCAAAACATT TTTGCATAAA 1500 ATAATATTAA TATTAGTAAC CACCTAGATA AATTAGAGAG AAACGTAGAA CATACCAAAA 1560

AAAATTAGAA CAAAAAGAAT ATTACAAAAA ATAATAAAAT TAAATTATTT CTTTACTATT 1620 AATTTAAAGT TTTTTTTCAT ATCATATATT ATGATACACA ATGTTTGTTG TTAAATGTTT 1680

TATATACATG CAATGATATG TTTCTGTTGG AATATGTATT ATATACTTAT ATGTTCTAAT 1740 AAATGTATTG TACACCTTTA GCAACTATTA CTACACACAT TTTTATATAA TTTATAACAG 1800 GAAAATATGT TATATTATTA CAATATCTTA ATGTGTTTTT GCAAAAATAT AAAAAACAAG 1860 AAAATTACAA TIGTAATTAA TCGTATGACA TAAAATTATA TTATATTAGA AATTAAAATT 1920 CAAAATTATA AAAAATATGG AAATGTTTTG TTATATTATT TTTTTAAAAA TTTAATTATT 1980 TTATTTTATT ATTTATTTT TTTTTTTTT GTGTTCTAAA TAAAAAGGCA AATATGATTC 2040 AAGTAAAAA TATATATT TACATAATGG CAAAATAATT GTTTATTATA TTATATGACT 2100 ATAATAATAT TITAGATTAA ACATATGTAA TICATTTAAC AGAATAAAAT AAAATATTAT 2160 ATATATAT TAATTATTAA GTTATAGATT TAATAAAAAT ATATTATACA TATGAGATTA 2220 10 AAAATGAAAG TTCACTACAG TAATATATTA TTATATGTCG TCAATTTAAG TATATTCTTA 2280 ATATCACGTA TGCACTAAAT AATGACAATA ATAATATATA TGTAACATTT TATAATTGAT 2340 GTAAATAAAA AAATATACAT ATATACAAAA ACATATATGA TATTTACATT CTTTTTTATA 2400 GATAAATATC CAGAAGAACT ATTACATCAC TTCACTTCAT ATACCAAACA CGAAAAAAAT 2460 ACAACCACTA GGTTATTATG CGAATGTGAC TTATATACGT CCATTTATGA TAATGACCCG 2520 15 GAAATGATAT TAGTGATGGA AAATTTCAAT AAACAGACAG AAGAAAGGTT TCATGAATAC 2580 AATGAACGCA TGCAAGAAAA ACGAAAAATA TGTAAAGAAC AATGCGAAAA GGATATACAA 2640 AAAATTATTT TAAAAGATAA AATCGAAAAG GAATTAACAG AAAAGTTAGA GGCATTGGAA 2700 ACGAATATAA AGACTGAGGA TATACCTACT TGTGTATGCG AAAAATCAGT AGCAGATAAA 2760 GTGGAAAAAA CGTGTTTGAA ATGTGGAGGT ATATTGGGTG TTGGTGTGAC TCCATCTTTA 2820 GGTTTATTAG GAGAAATAGG TGGACTTGTT ATAAATAATT GGACAAATAC TCCTTTTTAT 2880 AAAGCTTTTC TTACTTTTGC TCAAAAGGAA GGTATAGCTG CCGGTAAAAT TGCTAGTGAT 2940 ACTGCTCGTA TTGATACAGT TATTTAAGGA ATAATATCAA ATTTTGATGT GCACACTATA 3000 AATGGTTCTA CGTTGGGGAA AGTTATTACC GTAGAAGCTC TTAAGGATGA CACTACTCTT 3060 ACTACGGCAC TATATAATGA ATATGTAAGC ATGTGTGTAA ATACGAACCC TGTCGAAGAC 3120 25 AAATTAATTT GTGCTTTTGG GATGAGAGAC GGTCTAGTTG CAGGGCAATA TGCTTCATCG 3180 CGAGACGTTA TAGGATCAAG TGTAAAAGGA ATTATTAGAA AAGCTGCAAA CGCTGCTTCA 3240 CAAGCTGCTG AGACAGCTGC TAACGAAACT ACTTCCGGAA TGATCGAAGC CGAGTTAAGT 3300 AAAATAACAT CTGCAGGTGC TAATTTACAC AGTGCAATTA CTTACTCAGT AACTGCGATA 3360 TTGGTTATAG TTTTGGTTAT GGTAATTATT TATTTAATAT TACGTTATCG TAGAAAAAA 3420 30 AAAATGAAGA AAAAATTGCA ATATATAAAA TTATTAAAGG AATAGATATA CGATGTCGAG 3480 CTATTAGCGG TAATTTAAAG TATTGTGAAT TTTTCATTTA ATATGCTATG ATCATTTGAT 3540 AATTAATTTT TTTTTATAAT ATTATATTTT TTTATACCTT GGATTCTTAC ATTGTTTTAT 3600 TATTATATGA TTATTTAATT ATTATACTTA TATATATATA TATTTTTACA TTAAGATATT 3660 35 TTATTATTAT TAGATGCATA TTAGTGATGA TTATAATAAT AACCTATTGA AGAGAATAGA 3780 ACATAATAAT ATATTAAATT AATAGAACTT CATTTTTATT GTTATATGTA TATAAAAATA 3840 AGAAATTTGA AAAAGTAATT TACACATGAT AATGTATTTT ATTTTATTTG TGTTGTTTTA 3900 TATTTATTTA TAAAAATTGT TTAATATAAG TTGTTATTAT AATTTTTTAA TATGGCACCA 3960 TTAGCTTTCC ATTATACAAA TATATATTC CTCATTAGAA TCTGAATATT TATTGTATTA 4020 40 TAAAAAAAGT ATAATATAAT AAAATATCTA AGATTTTTTC TAATTTGTTT AATTTATAAT 4080 AAATTTTAAT TTTATACGAT AGAATAAATT ATAATCAACA TATATATATG TATTCATCTT 4140 AAGAACCTAT TACAATATAG TAACAACTGG TTCCTTTTTA TTATAAATAA CATAAGAATG 4200 TGTAAAAGGA TAGTTGTTAA AGGCTTTTTT AATATTGATT ATAAATGTTT GTAAGATATA 4260 TATAATAGAT ATCTTAACAT ACAACTTTGC ATAATTGTAA TTAAAAAAAT ATATATAATA 4320 AGAAATATTA TAAATAATAT TATAAAAAAT TAAGCATAAA TGTCACAATA AATTTTTTT 4380 45 TATTAATTTA ATTTTATTTT ATTGTTCTAA AATATATTGA TTATGAGAAT ATTATTTGTG 4440 TCTAATATAA TTAAGATATT TCTAATATTA ATTTATATAT ATATATTTAA AAGTATTTTA 4500 AGAATAATTT TTTACTTATT TATTATAATA TGAAATATGC ATGGAGTATA TATAAATATT 4560 GATGACAAAA AAAAAACTTT TAAAATGGAA AATATGCATA TAATAAAATA CTATATAGTA 4620 50 TAATTGGTGA AATAGTTGTA ACTTATACAA ACATGTTGCA TTCATAATTT AGAGATTATG 4680 TAATATTGTT TATGTATCGT AATATATAT AATATAATTG TTTTTTTAGT ATGTATGGTA 4740 TTCTAATAAT ATATTCATAT GTAGTCATAG TGTCAATGAA TATAAAATAT GGTATATTTA 4800 TATTATTGTA TATATTAAAT AAGTAACACA GAACATTATA TATAGTAATA AATAGAAGAA 4860 ATAATATAT TTTATGTTAT ATATTATAG TTATTATAAA GGGGAAAATT CATAATATTT 4920 55 ATGGAAAGCA TAAAAAATGT TACTGTAATA GGATAAAATA TATTATATAA AATGTTTATT 5040 TTATCTTAAA AAGGTTCCTA TTATAACATT AAAAAAAATT TGTCCCATTT TATAAATAAT 5100 TAACTACATT TACATAATGA AATTTCGATT TTGTGTTTTT TTGATGAATA TTATGGACTA 5160 ATTATTTATA TGTGAATGCG TICTATATAA TAATAATAAT TITATTIAAA AAAATGAAAA 5220 60 ATAAGAAATA AATATCCTGA TTTTGTAGTT CCAATAGCTT AATATAATTA TGGACTCATA 5280 TATATATTAT ATATATCTTT ACAACAAGTA ATAAGTAAAT ATTATTTTAA TCTTAATAAG 5340 GAAAATAAAA ATAATAAAAT AAGAATACTG AATAATAAGT CATATTATAC ATTTTTTAAA 5400 AATGTAACAT AATTACAAAT ACGTAACATG TATTATAGAA ATAATAAGAA TTTAATATTA 5460 

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	ATATTAACAT	GATTAGTTTT	TTGAAAAATA	TTTAAATATC	ATATAATAAT	AATAAATTAG	5580
	TTAAAATAAT	AGTATTTCAT	ACAAAATACT	AACTTATAAG	TATATCATAT	AATATTATAT	5640
	ATATATATAT	TTATGTGTTT	TTGATTGGGT	GTATATAAGG	CTATAAGTAT	ATATGGGTTG	5700
	TTCATTATAT	ATTTATATGT	GAATAGATAC	ΔΤΆΤΑΛΩΤΤΑ	<b>ልጥልጥልጥጥ</b> ልጥ	<b>ምነርጥረጥአጥአጥ</b>	5760 5760
5	Chalchtatany	AGATAGATAT	GCATTACAGT	ጥል <b>እርረርርም</b> አጥ	VCultabelished the VCultabelished to VCultabelis	TIGIGININI	5700
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	GIUCUIUIU	VIVNWWWWIN	GAIAACIAAC	MAINIGCAIA	11ACAAGAA1	AATATTTGTA	5880
	IMMMINIMI	WIWIWIWI	ATATATAAAG	ACATTAAAAC	TATACTAATA	GGTAATTAGT	5940
	TITATIATAT	CATCCTTTTA	TTATTATAAT	TTTTTTTTTT	TTACTTCTTG	TCGTTCTTTT	6000
40	TIGITATIAT	AATATAACAA	ATATAAAACA	ATATCAGTAT	TTGGAATATA	AATAAATTTA	6060
10	TTCTACATAT	ATGCATATAT	ATATATATAT	ATATATATAT	<b>ATATATATAT</b>	ATATATATAT	6120
	ATATGTATGA	TTTTATACTA	TTTTTATAÇA	TGCATTTTTA	TATATTTTAG	TATATACTTT	6180
	AAAGATATTA	TTAATATTTA	TATAGTAGCA	TATATGTATT	TATATTATAA	CAAATATTTT	6240
	CATTTATATA	<b>AATATATAGA</b>	ACATGAACAT	TTTATTAATA	ACTCATATTT	GAATATATAT	6300
	ATTTATAATG	<b>TGTATTTTTA</b>	CTTATTTTTT	TATATTATAC	AATAAAATTT	TGAAATTCAT	6360
15	AAAATGCATG	AAATACATAA	AAAAATACAA	CAAAACAAAT	GATAAAAACA	VALLVALALALA	6420
	TATAATATAA	TATAATATAA	TAATATATTT	TALCALCALATA	ערעתיידעמיים	thinininininininini	6480
	GATCCTATAT	מידע מידע מידע מידע מידע מידע מידע מידע	ATAATAAATT	מית	ACAACAAAAA	ת מיים מיים מיים	6540
	TAATATATTA		AATACAACAA	TACAAACAAT		TIMMIMMIMM	6540
	TAGIAIACIA	TATATATATAT	TGATAGATAA	TACAMAGAMI	AIGIAICIAI	AICAAIIAIA	6600
20	TATATATOMA	INIMIAMAIA	TOMINOMIAM	TATAGATAGA	GAGAAACGAA	GAACATATTT	6660
20	GICICITITE	TIATCICIAA	TATATATATA	TATATAATAA	ATTAAAATAA	AGTCAAAAAA	6720
	AATATACATA	TATTAATGTT	AATAATTAAA	TATATAAACA	CGTTGCATAT	ATACTTTTT	6780
	ATATGTTTGT	ATTTTCGTAT	TTTTTTTTC	TCATITATAA	TTTTACTTAA	TAAATAAAAC	6840
	ATAAAAAAA	TAATATATAT	ATAATTAAAT	AGATAAATAA	AGGAATACAT	TAATATAAA	6900
			TTTGTTAGAA				
25			AATATATAAA				
	TATTATTTTT	TTAACATATA	CATATATTGT	AATATTATAA	TAGTACAACT	ATTAATATAT	7080
	ATATATATAT	<b>ATATACAATA</b>	TTTATATATA	TTGTAATACA	TAAATTATAC	<b>CTTACATATA</b>	7140
	TATATACATT	CACAAAAGTG	TTATTATTCT	TATTCTACCA	TATTATAATA	CTACTGTAAT	7200
•	<b>ATACATATAT</b>	ACATACCCCC	ACGTACGTAC	GAAACACCAC	CAAACCATGT	ATCACGTATG	7260
30	TATGTATGCC	ACGATATAAA	CCACGTACCA	CGTATGACAT	AATGTAATGG	TGGAGTTAGC	7320
	AAAAATGGGG	CCCAAGGAGG	CTGCAGGTGG	GGATGATATT	GAGGATGAAA	GTGCCAAACA	7380
	TATGTTTGAT	AGGATAGGAA	AAGATGTGTA	CCATAAACTA	AAACACCAAC	CTADAGAACG	7440
	TGGTAAAGGC	TTGCAAGGAC	GTTTGTCAGA	ACCANANTTT	CACAAAAAATC	AAAGCCATCC	7500
	ACAAACACCA	CARCATCCAT	CCCATCTTCA	TO CARACITI	CAGAMMAIG	WWWGCGWICC	7500
35	TOTALTOTAL	CCCTCCCCTC	ATAGATCTGA	COMCOOMME	CHINCHMAIG	IMACIACIAA	7500
00							
	ATGTACACAT	WHINGWHIMM	AAGAIAGICA	ACAGGGIGAI	AATAAAGGTG	CATGTGCTCC	7680
	ATATAGGCGA	TIGCATGIAI	GCGATCAAAA	TITAGAACAG	ATAGAGCCTA	TAAAAATAAC	7740
	AAATACTCAT	AATTTATTGG	TAGATGTGTG	TATGGCAGCA	AAATTTGAAG	GACAATCAAT	7800
40	AACACAAGAT	TATCCAAAAT	ATCAAGCAAC	ATATGGTGAT	TCTCCTTCTC	AAATATGTAC	7860
40	TATGCTGGCA	CGAAGTTTTG	CGGACATAGG	GGACATTGTC	AGAGGAAGAG	ATTTGTATTT	7920
	AGGTAATCCA						
	CGGGAAAATA	TATGAAAAAT	TGAATGGCGC	AGAAGCACGC	TACGGAAATG	ATCCGGAATT	8040
	TTTTAAATTA	CGAGAAGATT	GGTGGACTGC	TAATCGAGAA	ACAGTATGGA	AAGCCATCAC	8100
	ATGTAACGCT	TGGGGTAATA	CATATTTTCA	TGCAACGTGC	AATAGAGGAG	AACGAACTAA	8160
45	AGGTTACTGC	CGGTGTAACG	ACGACCAAGT	TCCCACATAT	TTTGATTATG	TGCCGCAGTA	8220
	TCTTCGCTGG	TTCGAGGAAT	GGGCAGAAGA	TTTTTGTAGG	AAAAAAAATA	AAAAAATAAA	8280
	AGATGTTAAA	AGAAATTGTC	GTGGAAAAGA	TAAAGAGGAT	AAGGATCGAT	ATTGTAGCCG	8340
	TAATGGCTAC						
	ATGCATTAGC	THE THE THE THE	CATGTAATCC	ייים רבייים איי	ጥርርልጥልልልጥል	ACCANANAGA	8460
50	ACAATTTGAC	DADCACADAD	AAAAATATGA	TCAACAAATA	TOURINGTE	AAAATCCACC	0400
00	ATCAGGTGGT						
	ATATGAAAAA						
	TTTGGAAAAA						
	AATTGATTTT						
55	AAGTCAAGGA						
	AAAGGTAAAT						
	GAGTGGAAAA						
	AAGTGGTAAA	GGACATGATG	ATATTGAAGA	AAAATTAAAC	AAATTTTGTG	ATGAAAAAA	9000
	TCCTCATACA	ATA A ATACTC	CTCCTACTCC	TACCCCTCCT	ACTECTECTE	CTAACACTCC	2060
60	TAGACAGGAA	TTGTATGAAG	AATGGAAATG	TTATAAAGGT	GAAGATGTAG	TGAAAGTTGG	9120
	ACACGATGAG						
	ATTAAAAAAC						
	TGAAATCCAA						
	TTCCATACAT						

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- (2) INFORMATION FOR SEQ ID NO:14:
- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 3060 amino acids
    - (B) TYPE: amino acid

45

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Val Glu Leu Ala Lys Met Gly Pro Lys Glu Ala Ala Gly Gly Asp 10 Asp Ile Glu Asp Glu Ser Ala Lys His Met Phe Asp Arg Ile Gly Lys 50 25 Asp Val Tyr Asp Lys Val Lys Glu Glu Ala Lys Glu Arg Gly Lys Gly Leu Gln Gly Arg Leu Ser Glu Ala Lys Phe Glu Lys Asn Glu Ser Asp 55 55 Pro Gln Thr Pro Glu Asp Pro Cys Asp Leu Asp His Lys Tyr His Thr 70 Asn Val Thr Thr Asn Val Ile Asn Pro Cys Ala Asp Arg Ser Asp Val 90 60 Arg Phe Ser Asp Glu Tyr Gly Gly Gln Cys Thr His Asn Arg Ile Lys 105 Asp Ser Gln Gln Gly Asp Asn Lys Gly Ala Cys Ala Pro Tyr Arg Arg 120 125 Leu His Val Cys Asp Gln Asn Leu Glu Gln Ile Glu Pro Ile Lys Ile 130 135

	145					150					155					Phe 160
					165					170	ľ				175	Tyr
5				Pro 180					185					190	Phe	Ala
			195					200	ŀ				205	Gly	Asn	
10	Gln	Glu 210	Ile	Lys	Gln	Arg	Gln 215		Leu	Glu	Asn	Asn 220	Leu	ГÀЗ	Thr	Ile
	225					230					235	Glu	Ala			Gly 240
	Asn	Asp	Pro	Glu	Phe 245	Phe	Lys	Leu	Arg	Glu 250		Trp	Trp	Thr	Ala 255	Asn
15	Arg	Glu	Thr	Val 260	Trp	Lys	Ala		Thr 265		Asn	Ala	Trp	Gly 270	Asn	Thr
			275	Ala				280					285	Gly	Tyr	_
20		290		Asp			295					300				•
	305			Trp		310					315				_	320
05				Ile	325					330					335	
25				Asp 340					345					350		_
			355	Ala				360					365			
30		370		Ala			375					380				-
	385			Asp		390					395					400
35				Gly	405					410					415	_
<b>J</b> J				Thr 420					425					430		_
		•	435	Lys				440					445			_
40 '		450		Glu			455					460			-	_
	465			Phe		470					475					480
45				Val	485					490					495	
73				Pro 500					505					510	_	
			<b>51</b> 5	Trp				520					525		_	-
50		530		Pro			535					540				
	545			Lys		550					555				_	560
<b>5</b> 5				Lys	565					570		_	_		575	
33				Gly 580					585					590		
			595	Tyr				600					605		_	
60		61Ú		Glu -			615					62Ú				_
	625			Asn -		630					635		_			640
					645					650		•			655	•
65	Tyr	Trp	Val-	Ala	His	Met	Leu	Lys	Asp	Ser	Ile	His	Trp	Lys	Lys	Lys

										•						
	Leu	Gln	Arg	660 Cys		Gln	Asn	Gly	665 Asn		Ile	Lys	Cys	670 Gly	Asn	Asn
		Cys	<b>67</b> 5					680					685	_		
5		Asp					695					700				-
	705					710					715					720
40		Gly			725					730					735	_
10		Asp		740					745					750	_	
		Ser	755					760					765		_	
15		Glu 770					775		•			780				
	Asp 785	Asn	Asn	Gln	Glu	Ala 790	Ser	Val	Gly	Gly	Gly 795	Val	Thr	Glu	Gln	Lys 800
	Asn	Ile	Met	Asp	<b>L</b> ys 805	Leu	Leu	Asn	Tyr	Glu 810	Lys	Asp	Glu	Ala	Asp 815	Leu
20	Cys	Leu	Glu	Ile 820	His	Glu	Asp	Glu	Glu 825	Glu	Glu	Lys	Glu	Lys	Gly	Asp
	Gly	Asn	Glu 835	Cys	Ile	Glu	Glu	Gly 840	Glu	Asn	Phe	Arg	Tyr 845		Pro	Cys
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	Ala 865	Tyr	Gln	Met	His	His 870	Lys	Ala	Lys	Thr	Gln 875	Leu	Ala	Ser	Arg	Ala 880
	Gly	Arg	Ser	Ala	Leu 885	Arg	Gly	qaA	Ile	Ser 890		Ala	Gln	Phe	Lys 895	Asn
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	Asn	Tyr	Ser 915	Asn	Asp	Ser	Arg	Gly 920	Asn	Ser	Gly	Gly	Pro 925	Cys	Thr	Gly
35		Asp 930					935					940				_
	Ser 945	Asn	Ile	Glu	Gly	Lys 950	Lys	Gln	Thr	Ser	Tyr 955	Lys	Asn	Val	Phe	Leu 960
	Pro	Pro	Arg	Arg	Glu 965	His	Met	Cys	Thr	Ser 970	Asn	Leu	Glu	Asn	Leu 975	
40	Val	Gly	Ser	Val 980	Thr	Lys	Asn	Asp	Lys 985	Ala	Ser	His	Ser	Leu 990		Gly
		Val	995					1000	) .				100	5	_	_
45		Lys 1010	)				1015	5			-	1020	)			-
	1025					1030	)				1035	;				1040
		Asp			1045	5				1050	)				105	5
50		Asp		1060	)				1065	5	•			1070	<u> </u>	
		His	1075	5				1080	)				1089	5		
55		Lys 1090	•	•			1095	5				1100	) _			
	1105		•			1110	)				1115					1120
	Cys	Pro	Gly	Met	Pro 1125		Asp	Asp	Tyr	Ile 1130		Gln	Arg	Leu	Arg 113	_
60		Thr		1140	)				1145	5				1150	Ğlu )	īyr
	Asp	Lys	Leu 1155		Lys	Ile	Cys	Ala 1160		Суѕ	Met	Ser	Lys 116	Gly		Gly
65	Lys	Cys 1170		Gln	Gly	Asp	Val 1175		Cys	Gly	Lys	Cys 1180	Lys		Ala	Cys

•	Asp	Lys	Tyr	Lys	Glu	Glu	Ile	Glu	Lys	Trp			Gln	Trp	Arg	Lys
	118 Ile	Ser	Asp	Lys	Tyr	119 Asn		Leu	Tvr	Leu	119: Gln	5 Ala	Lvs	Thr	Thr	1200 ·
5		•			120	5				121	0				121	5
J		Asn		122	U				122	5				122	Λ	
	Gln	Met	Val	Asp	Phe	Leu	Thr	Pro	Ile	His	Lys	Ala		Ile	Ala	Ala
•	Arg	Val	123 Leu		Lys	Arg	Ala	124 Ala		Ser	Pro	Thr	124 Glu	5 Ile	Ala	Ala
10 .		125	0 '				125	5				126	0			
	126					127	0				1279	5				1280
	Glu	Ile	Gly	Tyr	Gly 128	Gly 5	Cys	Gln	Glu			Gln	Phe	Cys		
15	Lys	His	Gly	Ala			Thr	Ser	Thr	129 Thr	Lys	Glu	Asn	Lys	129 Glu	5 Tyr
		Phe		130	0				130	5				131	0	
			131	5				132	0	•			132	5		
20	Asn	Arg	Ser O	GIn	Thr	Glu	Glu 133	Pro 5	Lys	Lys	Lys	Glu 134	Glu n	Asn	Val	Glu
	Ser	Ala	Cys	Lys	Ile	Val	Glu	Lys	Ile	Leu		Gly	Lys	Asn	Gly	Arg
	134 Thr	o Thr	Val	Gly	Glu	135 Cys		Pro	Lys	Glu	1355 Ser	5 Tvr	Pro	Asp	·Trò	1360 Asp
25					1369	5				137	0				137	5
20		Lys		138	0				138	5				139	0	
	Arg	Arg	Gln 1399	Lys	Leu	Cys	Leu	Tyr 140	Tyr	Ile	Ala	His		Ser	Gln	Thr
20	Glu	Asn	Ile		Thr	Asp	Asp			Lys	Asp	Ala	140: Phe	Ile	Lys	Thr
30		1410 Ala	0				141	5				142	n			
	142	>				143	)				1435	;	•			1440
		Ser			1445	5				1450	)				145	5
35	Leu	Arg	Ser	Met 1460	Met	Tyr	Thr	Phe			Tyr	Arg	Asp		Cys	Leu
	Asn	Thr	Asp	Ile		Lys	Lys	Gln	1469 Asn	Asp	Val	Ala	Lys	1470 Ala	Lys	Asp
•		Ile	1475	•				148	0				1489	5		
40		1490	)				1499	5				1500	)		-	
	150	Ser	Arg	GIN	GIu	Trp	Trp )	Lys	Thr	Asn	Gly 1515	Pro	Glu	Ile	Trp	Lys 1520
	Gly	Met	Leu	Cys	Ala 1525	Leu	Thr	Lys	Tyr		Thr	Asp	Thr	Asp		Lys
45	Arg	Lys	Ile	Lys	Asn	Asp	Tyr	Ser	Tyr	1530 Asp	Lvs	Val	Asn	Gln	1539 Ser	Gln
		Gly		1540	)				1545	5				1550	)	
			1555	;				1560	)				1565	5		
50	Arg	Trp 1570	Met )	He	GIu	Trp	G1y 1575	Glu 5	Glu	Phe		Ala 1580		Arg	Gln	Lys
	Lys 1589	Glu	Asn	Ile	Ile	Lys	Asp	Ala	Cys		Glu	Ile	Asn	Ser		
		Cys	Asn	Asp	Ala	1590 Lys		Arg	Cys		1595 Gln		Cys	Arq	Ala	1600 Tvr
55		•			1605	5				1610	)				1615	5
		Glu		1620	)				1625	5				1630	)	
	Asn	Phe	Val 1635	Leu	Lys	Ala	Asn	Val 1640		Pro	Gln	Asp	Pro 1645		Tyr	Lys
00	Gly	Tyr	Glu		Lys	Asp	Gly	Val		Pro	Ile	Gln	Gly	, Asn	Glu	Tyr
60	Leu	1650 Leu		Lvs	Cvs	Asp	1655 Asn		Lvs	Cve	Ser	1660 Cvs	) Met	λαη	Glv	Δen
	1665	•				1670	)				1675		•			1680
	val	Leu	ser	val	Ser 1685	Pro	гуѕ	GIu	ьуs	Pro 1690		Gly	Lys	Tyr	Ala 1695	_
65	Lys	Tyr	Pro	Glu			qaA	Cys	Tyr			Lys	His	Val	Pro	Ser

				. 170					170	15				171	.0	
	Ile	Pro	Pro 171	Pro 5	Pro	Pro	Pro	Val 172		Pro	Gln	Pro	Glu 172		Pro	Thr
5	Val	Thr 173	Val	Asp	Val	Cys	Ser 173	Ile 5	Val	Lys	Thr	Leu 174	Phe	Lys	Asp	Thr
	Asn 174	Asn 5	Phe	Ser	Asp	Ala 175	Cys		Leu	Lys	Tyr 175	Gly	Lys	Thr	Ala	Pro 1760
	Ser	Ser	Trp	Lys	Cys 176		Pro	Ser	Asp	Thr 177	Lys	Ser	Gly	Ala	Gly 177	Ala
10	Thr	Thr	Gly	Lys 178	Ser 0	Gly	Ser	Asp	Ser 178	Gly		Ile	Cys	Ile 179	Pro	Pro
•	Arg	Arg	Arg 179		Leu	Tyr	Val	Gly 180		Leu	Gln	Glu	Trp 180	Ala	Thr	Ala
15		181	0	Gly			181	5				182	0		_	_
	182	5				1830	D				183	Glu 5	Thr			Leu 1840
				Tyr	184	5				185	0				185	Gln 5
20				Ser 186	0				186	5				187	Glu 0	Asp
			187					188	0				188	5		
25		189	0	Phe			189	5				190	0			
	190	5		Thr		1910	)				1915	5				1920
30				Leu	1925	5				193	0				193	5
				Ile 1940	)				194	5				195	D	
			195					196	0				196	5		
35		1970	0	Ile			197	5				198	n	_		
	T38;	>		Asp		1990	)				1995	5				2000
40				Val	2005	5				201	)			_	201	5
				Ser 2020	)				202	5				203	ם כ	-
			2035					204	0				204	5		
45		2050	)	Thr			2055	5				2060	)			-
	2065	5		Leu		2070	)				2075	;		_	_	2080
50					2085	5				2090	)				2095	Gly 5
<b>50</b>				2100	)				210	5	•			2110	)	Gly
•			2115					2120	0				2125	5		- •
55		2130	)	Pro			2135	5				2140	) '			_
	2145	5		Lys		2150	1				2155				-	2160
				Asp	2165	i				2170	)				2175	3
50				Thr 2180	)				2189	5				2190	)	_
			2195					2200	)				2205	5		
<b>3</b> 5	Asp	Asp 2210		Lys	Thr	Phe	Lys 2215	His	Thr	Lys	Asp	Cys 2220		Pro	Cys	Leu

	Lys 222	Phe 5	Ser	Val	Asn	Cys 223	Lys 0	Lys	Asp	Glu	Cys 223		Asn	Ser	Lys	Gly 2240
					224	5				225	Ala 0	Thr	_		225	Asn
5				226	0				226	Arg 5	Val			227	Ser	Lys
			227	5				228	0				228	Gly	Ala	Gly
10		Phe 229	U				229	5				230	0			
	230	5.				231	0				231!	5			_	Gly 2320
15		His			232	5				233	0				233	5
13		Phe		234	0				234	5				235	0	
		Asn	235	5				236	0				236	Glu 5	Lys	
20		Asp 237	0				237	5				238	0			_
	Asp 238	Gln 5	Tyr	Lys	Asn	Asp 239	Asn O	Ser	Asp	Asp	Asp 2399		Val	Arg	Ser	Phe 2400
•		Glu			240	Pro 5	Gln			241	Ala O	Asn		_	241	Lys 5
25	Val	Ile	Lys	Leu 242	Ser	Lys	Phe	Gly	Asn	Ser	Cys	Gly	Cys		Ala	Ser
	Ala	Asn	Glu 243	Gln		Lys	Asn	Gly 244	242: Glu	Tyr	Lys	Asp	Ala 244		Asp	Cys
30	Met	Leu 2450	Lys	Lys	Leu	Lys	Asp 245	Lys	Ile	Gly			Glu	Lys	Lys	His
	His 246	Gln		Ser	Asp	Thr 247	Glu		Ser	Asp	Thr 2475		Gln	Pro	Gln	Thr 2480
	Leu	Glu	Asp	Glu	Thr 248		Asp	Asp	Asp		Glu		Glu	Glu		Lys
35	Lys	Asn	Met	Met 2500	Pro		Ile	Cys	Glu 250			Leu	_	Thr 251		5 Gln
	Gln	Glu	Asp 2515	Glu		Gly	Cys	Val 2520	Pro		Glu	Asn	Ser 252	Glu	Glu	Pro
40	Ala	Ala 2530	Thr	Asp	Ser	Gly	Lys 2535		Thr	Pro			Thr	Pro	Val	Leu
	Lys 2545	Pro		Glu	Glu	Ala 2550	Val		Glu		Pro 2555		Pro	Pro	Pro	Gln 2560
		Lys	Ala	Pro	Ala 2565	Pro		Pro	Gln	Pro 2570	Gln	Pro	Pro	Thr	Pro 257	Pro
45	Thr	Gln	Leu	Leu 2580	Asp	Asn	Pro	His	Val 2585	Leu	Thr	Ala	Leu	Val 259	Thr	Ser
		Leu	2595	Trp	Ser	Val	Gly	Ile 2600	Gly )	Phe	Ala	Thr	Phe 2605	Thr	Tyr	
50	Tyr	Leu 2610		Lys	Lys	Thr	Lys 2615		Ser	Val	Gly	Asn 2620		Phe	Gln	Ile
	2625					2630	)				2635	Thr	Lys			2640
	Asn	Arg	Tyr	Ile	Pro 2645	Tyr	Thr	Ser	Gly	Lys 2650		Arg	Gly	Lys	Arg 265	
55		Tyr		2660	)				2665	Asp	Ser			2670	Asp	His
	Tyr	Ser	Asp 2675	Ile	Thr	Ser	Ser	Glu 2680		Glu	Tyr	Glu	Glu 2689		Asp	Ile
60	Asn	Asp 2690	Ile		Val	Pro	Gly 2695	Ser		Lys	Tyr	Lys 2700	Thr		Ile	Glu
	Val 2705	Val		Glu	Pro	Ser 2710	Gly		Asn		Thr 2715	Ala	Ser	Gly	Asn	Asn 2720
		Thr	Ala	Ser	Gly 2725	Asn		Thr	Thr	Ala 2730	Ser	Gly	Lys	Asn	Thr 273!	Pro
65	Ser	Asp	Thr	Gln			Ile	Gln	Asn '			Ile	Pro	Ser	Ser	Lys

																•
				274					274					275	0	
		Thr	275	5				276	0				276	5		
5		Leu 277		Ser	Glu	Pro	Asn 277	Thr	Glu	Pro	Asn	Met 278	Leu	Gly	Tyr	Asn
		Asp		Asn	Thr	His 279	Pro		Thr	Ser	His 279	His		Val	Glu	Glu 2800
		Pro	Phe	Ile	Met 280	Ser		His	Asp	Arg 281	Asn	Leu	Phe	Ser	Gly 281	Glu
10	Glu	Tyr	Asn	Tyr 282	Asp		Phe	Asn	Ser 282	Gly		Asn	Pro	Ile 283	Asn	Ile
	Ser	Asp	Ser 283	Thr		Ser	Met	Asp 284	Ser		Thr	Ser	Asn 284	Asn		Ser
15 .	Pro	Tyr 285	Asn			Asn	Asp 285	Leu		Ser	Gly	Ile 286	Asp	Leu	Ile	Asn
	Asp 286	Ala 5	Leu	Ser	Gly	Asn 2870	His		Asp	Ile	Tyr 2875	Asp	Glu	Met	Leu	Lys 2880
	Arg	Lys	Glu	Asn	Glu 288	Leu 5	Phe	Gly	Thr	Lys 289	His		Thr	Lys	His 289	Thr
20	Asn	Thr	Tyr	Asn 290	Val		Lys	Pro	Ala 290	Arg		Asp	Pro	Ile 291	Thr	
		Ile	291	5				292	Leu	Asp			292	Asp 5	Met	_
25		Lys 293	0				293	5				294	0	•		
	294	_				2950	)				2955	5				2960
		Asn			2965	5				2970	)			_	297	5
30		Pro		2980	)				2985	5				2990	0	_
		Ser.	2995	5				3000	)				300	5		
35		Tyr	0.				3015	5				302	0			
	302	_				3030	)				3035	5				3040
40		Thr	_		Asn 3045		Ąsp	Val	Pro	Thr 3050		Met	His	Ile	Glu 3055	
40	Asn	Ile	Val	Asn 3060	)											
	(2) INFO	RMAT.	ION I	FOR S	SEQ 1	D NC	):15	:								
45	(i) SE															
	· (B) '	LENG:	: nuc	cleid	aci	id	ırs									
50		STRAI TOPOI				ıgıe				•						
<b>3</b> U	(ii) MO								•							
	(iii) (iv) AN				_: NC	)										
55	(xi) SE	OUENC	E DE	SCRI	ידים:	N: 5	SEO 1	ים או	) • 15 •							

55 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

			•				
	CGTGGTAGTG	GCGGTAGTAG	TAGTGGTAAA	GGGAAGAAGG	ATACATCTGA	GTATATTTAT	600
	GTGAGCGATG	CTAAGGATCT	TTTGGATAGA	GTTGGAGAAA	AAGTGTACGA	AGAAAAAGTG	660
	AAAAATGGTG	ATGCTAAAAA	ATATATTGAG	GCGTTGAAAG	GAAATTTGAA	CACAGCAAAT	720
_	GGTCGTAGTT	CGGAAACAGC	TAGCAGTATT	'GAAACGTGCA	CCCTTGTAAA	AGAATATTAT	780
5	GAGCGTGTTA	ATGGTGATGG	TAAAAGGCAT	CCGTGCAGAA	AAGACGCAAA	AAATGAAGAT	840
	GTAAACCGTT	TTTCGGATAC	ACTTGGTGGC	CAATGTACAT	' ACAATAGGAT	AAAAGATAGT	900
	CAACAGGGTG	ATAATAAAGT	AGGAGCCTGT	GCTCCGTATA	GACGATTACA	TTTATGTGAT	960
	TATAATTIGG	AATCTATAGA	CACAACGTCG	ACGACGCATA	AGTIGTTGTT	AGAGGTGTGT	1020
10	AIGGCAGCAA	AATACGAAGG	AAACTCAATA	AATACACATT	ATACACAACA	TCAACGAACT	1080
10	CATATICAGGATT	CTGCTTCCCA	ATTATGTACT	GTATTAGCAC	GAAGTTTTGC	AGATATAGGT	1140
	CHIMICGIMA	GAGGAAAAGA	CAAACAMAM	GGTTATGATA	ATAAAGAAAA	AGAACAAAGA	1200
	AMMAMAI ING	AACAGAAATT CACAAGAACG	GAAAGATATT CTX CXTX CXT	TTCAAGAAAA	TACATAAGGA	CGTGATGAAG	1260
	AGAGAAGATT	GGTGGACGTC	CINCAIAGAI	ACACTA TOCA	GAGGAGATTT	TTTTCAATTA	1320
15	CCAAAAGAAG	CTAATTATTT	TATABARACA	CCCCCTCTA ATC	TACCALIAGE	ATGTCATGCA	1380
	CAATGCCATT	GCATTGGTGG	AGATGTTCCC	ACATATATIC	ATTATCTCC	CCACTATIGGT	1440
	CGCTGGTTCG	AGGAATGGGC	AGAAGACTTT	TGCAGGAAAA	ATTAIGIGCC	ACTACAAAAT	1500
	TTGCAAAAAC	AGTGTCGTGA	TTACGAACAA	AATTTATATT	СТАСТССТАА	TCCCTDCCDT	1620
	TGCACAAAAA	CTATATATAA	AAAAGGTAAA	CTTGTTATAG	GTGAACATTG	TACAAACTCT	1680
20	TCTGTTTGGT	GTCGTATGTA	TGAAACTTGG	ATAGATAACC	AGAAAAAAGA	ATTTCTTAAAA	1740
	CAAAAAAGAA	AATACGAAAC	<b>AGAAATATCA</b>	GGTGGTGGTA	GTGGTAAGAG	TCCTAAAAGG	1800
	ACAAAACGGG	CTGCACGTAG	TAGTAGTAGT	<b>AGTGATGATA</b>	ATGGGTATGA	AAGTAAATTT	1860
•	TATAAAAAAC	TGAAAGAAGT	TGGCTACCAA	GATGTCGATA	AATTTTTAAA	AATATTAAAC	1920
	Aaagaaggaa	TATGTCAAAA	ACAACCTCAA	GTAGGAAATG	AAAAAGCAGA	TAATGTTGAT	1980
25	TTTACTAATG	AAAAATATGT	AAAAACATTT	TCTCGTACAG	<b>AAATTTGTGA</b>	ACCGTGCCCA	2040
	TGGTGTGGAT	TGGAAAAAGG	TGGTCCACCA	TGGAAAGTTA	AAGGTGACAA	<b>AACCTGCGGA</b>	2100
	AGTGCAAAAA	CAAAGACATA	CGATCCTAAA	AATATTACCG	ATATACCAGT	ACTCTACCCT	2160
	GATAAATCAC	AGCAAAATAT	ACTAAAAAAA	TATAAAAATT	TTTGTGAAAA	AGGTGCACCT	2220
20	GGTGGTGGTC	AAATTAAAA	ATGGCAATGT	TATTATGATG	AACATAGGCC	TAGTAGTAAA	2280
30	AATAATAATA	ATTGTGTAGA	AGGAACATGG	GACAAGTTTA	CACAAGGTAA	ACAAACCGTT	2340
	TCCAACACAC	ATGTTTTTT	TTGGGATTGG	GTTCATGATA	TGTTACACGA	TTCTGTAGAG	2400
	1GGAAGACAG	AACTTAGTAA	GTGTATAAAT	AATAACACTA	ATGGCAACAC	ATGTAGAAAC	2460
	WHINATHWAT	GTAAAACAGA	TIGIGGTIGT	TTTCAAAAAT	GGGTTGAAAA	AAAACAACAA	2520
35	CTTATCCTAT	CAATAAAAGA TTAGTCCCTA	TOCA COTTOTO	CACCERTOCITE	ATATTGTCCA	ACAAAAAGGT	2580
	Саравтатта	AAGATGTTCA	TCCACATACA	CATCACATAA	A A CA CAMMA A	CARACTETTE	2640
	GATGAGGAAG	ACGCAGTAGC	ACTITIONALIACA	CCTCCCAACC	AACACATIAA	SAMACIGITG	2700
•	TTACTACAAC	ACGAAAAAGA	ACAAGCAGAA	CAATGCAAAC	ADADGCAGGA	WHITCHIMM	2020
	AAAAAAGCAC	AACAAGAAAG	TCGTGGTCGC	TCCGCCGAAA	CCCGCGAAGA	CCDDDCCDCD	2820
40	CAACAACCTG	CTGATAGTGC	CGGCGAAGTC	GAAGAAGAAG	AAGACGACGA	CGACTACGAC	2940
*	GAAGACGACG	AAGATGACGA	CGTAGTCCAG	GAGGAGGAAG	AGGGAAAGGA	GGAAGGAACG	3000
	GTCACAGAGG	TAACAGAGGT	AACAGAGGTC	<b>GTGGAAGAGA</b>	CGGTAACAGA	ACAGGAAGGG	3060
	GTGAAGCCAT	GTGACATAGT	GGGCAAACTA	TTTGAGGACG	ACAAAAGTCT	CAAAGAGGCA	3120
	TGTGGTCTAA	AATACGGTCC	AGGTGGAAAA	GAAAAATTCC	CCAATTGGAA	GTGTGTCACA	3180
45	CCAAGTGGTG	TCAGTACTGC	CACTAGTGGA	AAAGACGGCG	CTATATGTGT	GCCACCCAGG	3240
	AGACGACGAT	TATACGTAGG	TGGTTTATCA	CAATGGGCAA	GTCGTGGTGG	TGACGAGACC	3300
	ACGGAGGTGT	CGAGTGAAGC	CACTTCGGCG	CCGTCACAGT	CAGAAAGTGA	AAAACTACGT	3360
	ACTGCGTTTA	TTGAGTCCGC	TGCAATAGAG	ACGTTTTTTT	TGTGGCATAA	GTATAAAGAA	3420
EO	GAGAAAAAAC	CACCAGCAAC	ACAAGATGGA	GCGGGACTTG	GAGTATCACT	CCCAGAACCG	3480
50	TCACCACCGG	GAGAGGACCC	CCAAACACAA	TTACAACAAA	CTGGTGTTAT	ACCCCCGAT	3540
	TTTTTGCGTC	AAATGTTTTA	TACATTAGCA	GACTACAAAG	ACATATTATA	CAGTGGTAGT	3600
	AACGACACAA	GIGACACAAC	TGGTAAACAG	ACACCTAGTA	GTAGTAATGA	CAACCTCAAA	3660
	AATATTGTTC	TGGAAGCAAG	TGGTAGTACT	GAGCAGGAGA	AGGAGAAAAT	GAAACAAATA	3720
<b>5</b> 5	CAAGCGAAAA AATAGTGTCA	TAAAAAAAAT	TTTAAACGGT	GCCACATCTG	GIGICCCACC	TGTCACCAAA	3780
33	GCTATGGTAT	CTCCACTAAC	ACAMACCIGG	A A TICA COCCO	CACCGAAGGA	TATCTGGAAT	3840
	GAACAGAATA	ACCATTACA A	N N N C C C N C TTT	TCCCACCAAC	CCAACAAG	TGCCAAAATA	3900
	GAGAAATACC	AGGATIIGAA	TOTON	CAACATCAAA	CTCCTCCTCCT	AACCCCCATA	3960
	ACCATCCAAC	CCCCCDCCTTTT	TGICWWWCIC	CACCY Y YANY C	OTOGIGCCAY	MAGCAACGAC	4020
<b>RN</b>	CATGAGTGGG	CCCCCACGII	United States of	DIGGRAMIAC	CINCALILLE	ATTOUTING TIA	4080
	GAGTGTATGG	ATGAGGATGG	TGAAAAACAA	TATACTCCCC	ATTCCCCANTA	መተረጥረን አረን አ	4200
	ATTTTTAGTA	AGCAATATAA	ТСТТСТССТС	CATTTALCOU	CCACLLCCCC	TIGIONAGHA	42UU 42CA
	AGATTGTATA	AAACGTGGAT	AGAAAAAAA	AAAACAGAAT	ATGAGADACA	TURECTION	432N
	TATGAACAAC	AAAAAAGTAA	TTACGAAAAT	GAACAAAAAG	ACAAATGCCA	AACACAAAGT	4380
65	AATAATAATG	CTAATGAATT	TTCTAGAACA	CTAGGAGCGT	CCCCTACAGC	TGCAGAATTT	4440
	. =	<b></b>					

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TTACAAAAGT TAGGATCATG TAAAAATGAT AATGGATATG AGAATGGAGA GGATAATAAA 4500
     ATAGATTITA AAAATCCAGA TAAAACATTT AAGGAAGCAC ACAGTTGTGA TCCATGTCCT 4560
     ATAACTGGAG TTAAATGTCA AAATGGTCAT TGTGTGGGTT CTGCTAATGG AAAGGAGTGC 4620
     AAAAACAATA AGATTACTGC AGAAGATATT AAAAATAAGA CAGATCCTAA TGGAAACATA 4680
     GAAATGGTTG TCAGTGATGA CAGTACAAAT ACATTTGAAC ATTTAGGCGA TTGTAAAAGC 4740
     TCAGGTATCT TTAAAGGTAT CAGAAAAGAT GAATGGAAAT GCGCTAATGT ATGTGGTGTA 4800
     GATATATGTA CTCTGGAAAA AAAAATTAAG AATGGGCAAG AAGGTGATAA AAAATATATC 4860
     ACAATGAAAG AATTGCTTAA ACGATGGCTA GAATATTTTT TAGAAGATTA TAATAGAATT 4920
     AGAAAAAAA TAAAGCTATG TACGAAAAAG GAAGATGGAT GCAAATGTAT AAAAGGTTGT 4980
10
     ATAGAAAAAT GGGTACAAGA AAAAACGAAA GAATGGCAAA AAATAAACGA TACTTATCTT 5040
     GAACAATATA AAAATGATGA TGGTAATACT TTAACTAATT TTTTGGAGCA ATTCCAATAT 5100
     CGAACTGAAT TTAAAAACGC TATAAAACCT TGTGATGGTT TAGACCAGTT CAAGACTTCG 5160
     TGTGGTCTTA ATAGTACTGA TAATTCACAA AATGGTAATA ATAACGATCT TGTTCTATGT 5220
     TTGCTTAATA AACTTCAAAA AAAAATTAGT GAGTGTAAAG AACAACATAG TGGCCAAACC 5280
     CAAACACCGT GTGATAACTC TTCCCTTAGT GGTAAAGAAT CCACCCTCGT TGAAGACGTT 5340
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     GATGATTATG AGGAACAAAA CCCAGAAAAC AAAGTGGAAC AACCTAAATT TTGTCCAGAT 5400
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     AAAAAAAAG TGGAAGACAG TGTAATCGAA CAAAAAGAGG AAGAAGCAGC TAGTGCCCCA 5520
     GAGGAATCTC CTCCATTAAC CCCGGAAGCA CCAAAAAAAG AGGAAAATGT GGTACCAAAA 5580
     CCACCACCAC CACCAAAAAA ACGCCGAATC AAAACCCGTA ATGTGTTGGA CCACCCCGCT 5640
20
     GTCATACCCG CCCTCATGTC TTCTACCATC ATGTGGAGTA TTGGCATCGG TTTTGCTGCG 5700 TTCACTTATT TTTATCTAAA GAAAAAAACC AAATCATCTG TTGGAAATTT ATTCCAAATA 5760
     CTGCAAATAC CCAAAAGTGA TTATGATATA CCTACATTGA AATCAAGCAA TCGTTATATA 5820
     CCCTATGCAA GTGATAGACA TAAAGGCAAA ACATATATTT ATATGGAAGG AGATAGCAGT 5880
     GGAGATGAAA AATATGCATT TATGTCTGAT ACTACTGATA TAACTTCATC CGAAAGTGAG 5940
25
     TATGAAGAAT TGGATATTAA TGATATATAT GTACCAGGTA GTCCTAAATA TAAAACATTG 6000
     ATAGAAGTAG TACTTGAACC ATCAAAAAGA GATACACAAA ATGATATACA CAATGATATA 6060
     CCTAGTGATA TACCAAATAG TGACACCA CCACCCATTA CTGATGATGA ATGGAATCAA 6120
     TTGAAAAAG ATTTTATATC TAATATGTTA CAAAATACAC AAAATACGGA ACCAAATATT 6180
     TTACATGATA ATGTGGATAA TAATACCCAT CCTACCATGT CACGTCATAA TATGGACCAA 6240
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     AAACCTTTTA TTATGTCCAT ACATGATAGA AATTTATTTA GTGGAGAAGA ATACAATTAT 6300
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     AGTCTAACAA GTAACAACCA TAGTCCATAT AATGATAAAA ATGATTTATA TAGTGGTATC 6420
     GACCTAATCA ACGACGCACT AAGTGGTAAT CATATTGATA TATATGATGA AATGCTCAAA 6480
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35
     GTCGTTACCC AAACAAGTAG TGACGACCCT ATAACCAATC AAATAAATTT GTTCCATAAA 6600
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     AAATTGAAAG AATTGTGGGA AAATGAGACA CATAGTGGTG ACATAAATAG TGGTATACCT 6720
     AGTGGTAACC ATGTGTTGAA TACTGATGTT TCTATTCAAA TAGATATGGA TAATCCGAAA 6780
40
     ACAATGAATG AATTTACTAA TATGGATACA AACCCCGACA AATCTACTAT GGATACTATA 6840
     TTGGATGATC TAGAAAAATA TAACGAACCC TACTACTATG ATTTTTATAA ACATGATATC 6900
     TATTATGATG TAAATGATGA TAAAGCATCT GAGGATCATA TAAATATGGA TCATAATAAG 6960
     ATGGATAATA ATAATTCGGA TGTCCCCACT AACGTACAAA TTGAAATGAA TGTCATTAAT 7020
     AATCAGGAGT TACTACAAAA TGAATATCCT ATATCGCATA TGTAGGGAAT ATGAAAATAA 7080
     TAGATGTATA TATGTTTTTT TCTTTTTTTG TGTGTGTGCA GTTTATATTT TTTATTTGTA 7140
45
    TATATTTTT TTTTTGTGCA TTTGTCTATT TTTTATTTGT GCTTTATATA TATATATATT 7260
     TTATTCAGCT TGGACTTAAC CAGGCTGAAC TTGCT
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- 50 (2) INFORMATION FOR SEQ ID NO:16:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 2182 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
- 60 (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (v) FRAGMENT TYPE: N-terminal

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

	Met 1	Glu	Pro	Gly	Gly 5	Ser	Gly	Gly	Arg	Gly	Ser	Gly	Gly	Ser	Ser	Ser
5	Gly	Lys	Gly	Lys 20	Lys	Asp	Thr	Ser	Glu 25	Tyr	Ile	Tyr	Val	Ser 30	Asp	Ala
	Lys	Asp	Leu 35	Leu	Asp	Arg	Val	Gly 40		Lys	Val	Tyr	Glu 45	Glu	Lys	Val
10	Lys	Asn 50	Gly	Asp	Ala	Lys	Lys 55	Tyr	Ile	Glu	Ala	Leu 60		Gly	Asn	Leu
	65			Asn		70					75					80
45				Val	85					90					95	
15				Cys 100					105					110	_	
			115	Leu				120					125	_	_	
20		130		Asp			135				•	140			_	
	145			Asp		150					155					160
25				Leu Thr	165					170					175	
20				180 Leu					185					190	_	
			195	Arg				200					205	_		_
30		210		Arg			215					220	_		_	
	225			Lys		230					235					240
35				Ala	245					250					255	-
	•			260 Asn					265				_	270	_	-
			275	Ala				280					285			
40		290		Gly			295					300				
	305			Val		310					315					320
45				Arg	325					330				_	335	
				340 Tyr					345					350	-	
F0	Cys		355 Lys	Thr	Ile	Tyr		360 Lys	Gly	Lys	Leu	Val	365 Ile	Gly	Glu	His
50	Cys	370 Thr	Asn	Cys	Ser		375 Trp	Cys	Arg	Met	Tyr	380 Glu	Thr	Trp	Ile	Asp
	385 Asn	Gln	Lys	Lys	Glu	390 Phe	Leu	Lys	Gln		395 Arg	Lys	Tyr	Glu	Thr	400 Glu
55	Ile	Ser	Gly	Gly	405 Gly	Ser	Gly	Lys		410 Pro	Lys	Arg	Thr	Lys	415 Arg	Ala
	Ala	Arg		420 Ser	Ser	Ser	Ser		425 Asp	Asn	Gly	Tyr	Glu	430 Ser	Lys	Phe
60			435 Lys	Leu	Lys	Glu		440 Gly	Tyr	Gln	Asp		445 Asp	Lys	Phe	Leu
<del>uu</del>	Lys	450 Ile	Leu	Asn	Lys		455 Gly	Ile	Cys	Gln		460 Gln	Pro	Gln		
	465 Asn	Glu	Lys			470 Asn	Val	Asp	Phe		475 Asn	Glu	Lys	Tyr	Val	480 Lys
65	Thr	Phe	Ser		485 Thr	Glu	Ile	Cys	Glu	490 Pro	Cys	Pro	Trp	Cys	495 Gly	Leu

				E00					<b>50</b> 5							
	Glu	Lys	Gly 515	500 Gly	Pro	Pro	Trp	Lys 520		Lys	Gly	Asp	Lys 525		Cys	Gly
5	Ser	Ala 530	Lys	Thr	Lys	Thr	Tyr 535	qaA	Pro	Lys	Asn	Ile 540		Asp	Ile	Pro
	Val 545	Leu	Tyr	Pro	Asp	Lys 550	Ser	Gln	Gln	Asn	Ile 555			Lys	Tyr	Lys 560
	Asn	Phe	Cys	Glu	Lys 565	Gly	Ala	Pro	Gly	Gly 570		Gln	Ile	Lys	Lys 575	Trp
10	Gln	Cys	Tyr	Tyr 580	Asp	Glu	His	Arg	Pro 585	Ser	Ser	Lys	Asn	Asn 590	Asn	Asn
	Cys	Val	Glu 595	Gly	Thr	Trp	Asp	Lys 600	Phe	Thr	Gln	Gly	Lys 605	Gln	Thr	Val
15		Ser 610					615					620	_			
	625	Ser				630					635					640
		Asn			645					650					655	_
20		Cys		660					665					670		
		Lys	675					680					685		_	_
25	•	Ile 690					695					700				
	705	Asn				710					715	_	_		_	720
20		Lys			725					730					735	
30		Leu	•	740					745					750		
		Lys	755					760					765		_	
35		Tys 770					775					780				
	785	Glu Glu				790					795	_				800
40		Gln			805					810			_	_	815	
		Glu		820					825					830		
		Lys	835					840					845			
45		850 Lys					855					860		_		
•	865	Pro				870					875					880
50		Gly			885					890					895	
		Val		900					905		•			910		
		Glu	915					920					925			
55		930 Lys					935					940				
	945	Leu				950					955					960
δû		GTA			965					970					975	
		Asp		980			•	•	985					990		_
		Leu	995					1000	)				100	5		_
65		1010					1019				-	1020				

	Tyr Ser	Gly Se	r Asn	Asp		Ser	Asp	Thr	Thr 103		Lys	Gln	Thr	
		Ser As	n Asp 104	Asn		Lys	Asn	Ile 105	Val		Glu	Ala		
5	Ser Thi	Glu Gl			Glu	Lys	Met 106	Lys		Ile	Gln	Ala 107		
•	Lys Lys	Ile Le 1075		Gly	Ala	Thr	Ser		Val	Pro	Pro	Val	Thr	Lys
10	Asn Ser	Val Ly	s Thr	Pro	Gln 109	Gln		Trp	Trp	Glu 110	Asn		Ala	Lys
		Trp As	n Ala	Met 111	Val		Ala	Leu	Thr	Tyr	Lys	Glu	Asn	_
		Gly Th	r Ser 112	Ala		Île	Glu	Gln 113	Asn	Lys	Asp	Leu		
15	Ala Leu	Trp As	p Glu		Asn	Lys		Thr		Ile	Glu			Gln
	Tyr Thr	Asn Va 1155		Leu	Glu			Ser	Gly	Ala			Asn	Asp
20	Thr Ile	Gln Pr	o Pro	Thr				Phe	Val				Thr	Phe
20	Phe Arg	Trp Le	u His	Glu	117! Trp		Asn	Ser				Glu	Arg	
	1185 Lys Arg	Leu Al				His	Glu				Glu	Asp	Gly	1200 Glu
25	Lys Gln	Tyr Se	120 r Gly	-	Gly	Glu	Tyr	1210 Cys			Ile		121: Ser	
	Gln Tyr	12 Asn Va		Gln	Asp	Leu	122! Ser		Ser	Cys	Ala	123 Lys		Cys
	Arg Leu	1235				124	0	•			124	5		_
30	125 Gln Gln	0			125	5				1260	)	_		_
	1265	:		1270	)				1275	, -				1280
	Lys Asp		128	5				1290	)				129	5
35	Arg Thr	13	00				1309	5				1310	o ¯	
	Gly Ser	1315				1320	כ				132	5		-
40	Ile Asp 133	0			1335	5				1340	)			-
	Asp Pro 1345	Cys Pro	o Ile	Thr 1350		Val	Lys	Суѕ	Gln 1355		Gly	His	Cys	Val 1360
	Gly Ser	Ala Ası	1 Gly 136		Glu	Cys	Lys	Asn 1370		Lys	Ile	Thr	Ala 137	
45	Asp Ile	138	30				1385	5				1390	)	
	Ser Asp	Asp Ser 1395	Thr	Asn	Thr	Phe 1400		His	Leu	Gly	Asp 140	_	Lys	Ser
50	Ser Gly		. Lys	Gly	Ile 1415	Arg	Lys	Ąsp	Glu	Trp 1420		Cys	Ala	Asn
	Val Cys 1425	Gly Va	l Asp	Ile 1430	Cys		Leu	Glu	Lys 1435		Ile	Lys	Asn	Gly 1440
	Gln Glu	Gly Asp	Lys 144		Tyr	Ile	Thr	Met 1450	Lys		Leu	Leu	Lys 1459	Arg
55	Trp Leu	Glu Ty		Leu	Glu	Asp	Tyr 1465	Asn		Ile	Arg	Lys 1470	Lys	
	Lys Leu			Lys	Glu	Asp 1480	Gly		Lys	Cys	Ile 1489	Lys		Cys
00	Ile Glu 149		Val	Gln	Glu 1495	Lys		Lys	Glu	Trp 1500	Gln		Ile	Asn
	Asp Thr 1505		Glu	Gln 1510	Tyr		Asn	Asp	Asp 1515	Gly		Thr	Leu	Thr 1520
	Asn Phe	Leu Glu	Gln 1529	Phe		Tyr	Arg	Thr 1530	Glu		Lys	Asn .	Ala 1539	Ile
65	Lys Pro	Cys Asp			Asp	Gln	Phe			Ser	Cys	Gly		

				154					154	5				155	0	
	•	Thr	155	5				156	0				156	5		
5		Leu 157	0				157	5				158	0			
	158					159	0				159	5				1600
••		Ser			160	5				161	0				161	5
10		Asn		162	0				162	5				163	O	
		Lys	163	5,				164	0				164	5		
15		Lys 165	0				165	5				166	0			
	166					1670	)				167	5				1680
20		Glu			1683	5				169	0				169	5
20		Ile		170	0				170	5				171	0	
		Met	171	5				172	0				172	5		
25		Thr 1730	0				173	5				174	0			
•	174	Phe 5 Lys				1750	)				1755	5				1760
30		Lys			1769	5				1770	0			_	177	5 <sup>*</sup>
		Ala		1780	)				178	5				179	0	
		Glu	179	5				180	0				180	5		
35		1810 Lys	)				181	5				182	0			
	182	Asn				1830	)				1835	5				1840
40		Pro			1845	5				1850	)				185	5
		Ile	Ser	1860 Asn	)			Asn	1869 Thr	5				1870	)	_
AE	Leu	His	1875 <b>A</b> sp		Val	Àзр				His	Pro	Thr	1885 Met	5 Ser	Arg	His
45	Asn	1890 Met		Gln				Ile	Met	Ser	Ile	1900 His	) Asp	Arg	Asn	
	Phe	Ser	Gly	Glu	Glu	1910 Tyr			Asp	Met		Asn	Ser	Gly		
50	Pro	Ile	Asn	Ile 1940	1925 Ser		Ser	Thr				Asp	Ser			Ser
	.Asn	Asn	His 1955	Ser		Tyr	Asn	Asp			Asp	Leu			Gly	Ile
<b>55</b>	Asp	Leu 1970	Ile		Asp	Ala	Leu 1975	Ser		Asn	His	Ile 1980		Ile	Tyr	Asp
	Glu 1989	Met		Lys		Lys 1990	Glu		Glu		Phe 1995	Gly		Gln	His	His 2000
		Lys	Asn	Ile		Ser		Arg	Val		Thr		Thr	Ser	Ser 2019	Asp
en en	Àŝp	Prô	Ile	Thr 2020	ĀSII		Ile	ĀSII	Դeu 2029	Phe		гуs	Trp	Eeu 2030	Āsp	Arg
	His	Arg	Asp 2035	Met		Glu	Lys	Trp 2040	Lys		Asn	His	Glu 2045	Arg		Pro
65	Lys	Leu 2050	Lys		Leu		Glu 2055	Asn		Thr		Ser 2060	Gly		Ile	Asn

	Ser Gly Ile Pro Ser Gly Asn His Val Leu Asn Thr Asp Val Ser Ile 2065 2070 2075 2080
	Gln Ile Asp Met Asp Asn Pro Lys Thr Met Asn Glu Phe Thr Asn Met 2085 2090 2095
5	Asp Thr Asn Pro Asp Lys Ser Thr Met Asp Thr Ile Leu Asp Asp Leu 2100 2105 2110
	Glu Lys Tyr Asn Glu Pro Tyr Tyr Tyr Asp Phe Tyr Lys His Asp Ile 2115 2120 2125
10	Tyr Tyr Asp Val Asn Asp Asp Lys Ala Ser Glu Asp His Ile Asn Met 2130 2135 2140
	Asp His Asn Lys Met Asp Asn Asn Ser Asp Val Pro Thr Asn Val 2145 2150 2155 2160
	Gln Ile Glu Met Asn Val Ile Asn Asn Gln Glu Leu Leu Gln Asn Glu 2165 2170 2175
15	Tyr Pro Ile Ser His Met 2180
	(2) INFORMATION FOR SEQ ID NO:17:
20	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs
	(B) TYPE: nucleic acid (C) STRANDEDNESS: single
25	(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO
	(iv) ANTISENSE: NO (v) FRAGMENT TYPE:
30	(vi) ORIGINAL SOURCE:
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:
35	ATCGATCAGC TGGGAAGAAA TACTTCATCT 30
	(2) INFORMATION FOR SEQ ID NO:18:
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs
4D-	(B) TYPE: nucleic acid
	(C) STRANDEDNESS: single (D) TOPOLOGY: linear
45	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO
10	(iv) ANTISENSE: NO
	(v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:
	ATCGATGGGC CCCGAAGTTT GTTCATTATT 30
55	(2) INFORMATION FOR SEQ ID NO:19:
<b>J</b> J	(i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 30 base pairs (B) TYPE: nucleic acid
	(C) STRANDEDNESS: single
ซีบิ	(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO
or.	(iv) ANTISENSE: NO
65	(v) FRAGMENT TYPE:

	(vi) ORIGINAL SOURCE:	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:	
5	TCTCGTCAGC TGACGATCTC TAGTGCTATT	30
	(2) INFORMATION FOR SEQ ID NO:20:	
10	<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 30 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
15	<pre>(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:</pre>	
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
	ACGAGTGGGC CCTGTCACAA CTTCCTGAGT	30
25	(2) INFORMATION FOR SEQ ID NO:21:	
30	<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 17 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
35	<pre>(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:</pre>	
40	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:	,
	AGACCTCAAT TTCTAAG  (2) INFORMATION FOR SEQ ID NO:22:	17
45	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 18 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single	
50	(D) TOPOLOGY: linear	
	<pre>(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE:</pre>	
55	(vi) ORIGINAL SOURCE:	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	4.0
60	AATCGCGAGC ATCATCTG	18
	(2) INFORMATION FOR SEQ ID NO:23:	
65	<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 20 base pairs</li><li>(B) TYPE: nucleic acid</li></ul>	

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(C) STRANDEDNESS: single
             (D) TOPOLOGY: linear
           (ii) MOLECULE TYPE: cDNA
 5
           (iii) HYPOTHETICAL: NO
           (iv) ANTISENSE: NO
           (v) FRAGMENT TYPE:
           (vi) ORIGINAL SOURCE:
10
           (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:
     CCRAGRAGRC AARAAYTATG
                                                               20
             (2) INFORMATION FOR SEO ID NO:24:
15
           (i) SEQUENCE CHARACTERISTICS:
             (A) LENGTH: 18 base pairs
             (B) TYPE: nucleic acid
             (C) STRANDEDNESS: single
(D) TOPOLOGY: linear
20
           (ii) MOLECULE TYPE: cDNA
           (iii) HYPOTHETICAL: NO
           (iv) ANTISENSE: NO
25
           (v) FRAGMENT TYPE:
           (vi) ORIGINAL SOURCE:
           (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:
30
     CCAWCKKARR AATTGWGG
                                                               18
             (2) INFORMATION FOR SEQ ID NO:25:
           (i) SEQUENCE CHARACTERISTICS:
35
            (A) LENGTH: 291 amino acids
            (B) TYPE: amino acid
            (C) STRANDEDNESS: single
            (D) TOPOLOGY: linear
40
          (ii) MOLECULE TYPE: peptide
          (iii) HYPOTHETICAL: NO
          (iv) ANTISENSE: NO
          (v) FRAGMENT TYPE: internal
          (vi) ORIGINAL SOURCE:
45
          (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:
     10
                                                      15
50
     Xaa Xaa Xaa Val Cys Ile Pro Asp Arg Arg Tyr Gln Leu Cys Met Lys
               20
                                 25
                                                  30
     35
                             40
     55
                          55
                                           60
     70
                                        75
     Xaa Asp Phe Cys Lys Asp Ile Arg Trp Ser Leu Gly Asp Phe Gly Asp
                   85
                                    90
CO
     The The Met Gly Thr Asp Met Glu Gly The Gly Tyr Ser Lys Kaa Kaa
               100
                                 105
                                                  110
     Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Thr Asp Glu Lys Ala Gln Gln
                             120
                                              125
     Arg Arg Lys Gln Trp Trp Asn Glu Ser Lys Ala Gln Ile Trp Thr Ala
65
                          135
```

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	145	Met				150					155					160
	Cys	Xaa	Xaa	Xaa	Xaa 165	Xaa	Xaa	Xaa	Xaa	Glu 170	Pro	Gln	Ile	Tyr	Arg 175	Trp
5	Ile	Arg	Glu	Trp 180	Gly	Arg	Asp	Tyr	Val 185	Ser	Glu	Leu	Pro	Thr 190	Glu	Val
	Gln	Lys	Leu 195	Lys	Glu	Lys	Cys	Xaa 200	Xaa	Xaa	Xaa	Xaa	Xaa 205	Xaa	Xaa	Xaa
10	Xaa	Xaa 210	Cys	Xaa	Val	Pro	Pro 215	Cys	Gln	Asn	Ala	Cys 220	Lys	Ser	Tyr	Asp
	Gln 225	Trp	Ile	Thr	Arg	Lys 230	Lys	Asn	Xaa	Xaa	Xaa 235	Xaa	Xaa	Xaa	Xaa	Xaa 240
	Xaa	Xaa	Xaa	Xaa	Xaa 245	Xaa	Xaa	Xaa	Xaa	Xaa 250	Xaa	Xaa	Xaa	Xaa	Xaa 255	Xaa
15	Xaa	Xaa	Xaa	Xaa 260	Xaa	Xaa	Xaa	Xaa	Xaa 265	Xaa	Xaa	Xaa	Xaa	Xaa 270	Xaa	Xaa
	Xaa	Xaa	Xaa 275	Xaa	Xaa	Xaa	Xaa	Xaa 280	Xaa	Xaa	Xaa	Xaa	Xaa 285		Xaa	Xaa
20	Cys	Xaa 290	Cys													

#### (2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS: 25
  - (A) LENGTH: 271 amino acids (B) TYPE: amino acid

  - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide (iii) HYPOTHETICAL: NO 30

35

- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

	Cys 1	Xaa	Xaa	Xaa	Xaa 5	Xaa	Xaa	Xaa	Xaa	Xaa 10	Xaa	Cys	Xaa	Xaa	Xaa 15	Xaa
40	Xaa	Xaa	Xaa	Xaa 20	Xaa	Val	Cys	Ile	Pro 25	Asp	Arg	Arg	Ile	Gln 30		Суѕ
	Ile	Val	Asn 35	Leu	Xaa	Xaa	Xaa	Xaa 40	Xaa	Xaa	Xaa	Xaa	Xaa 45	Xaa	Xaa	Xaa
45	Xaa	Xaa 50	Xaa	Xaa	Xaa	Xaa	<b>Xaa</b> 55	Xaa	Xaa	Xaa	Xaa	Xaa 60	Xaa	Xaa	Xaa	Xaa
	Xaa 65	Xaa	Xaa	Xaa	Xaa	Xaa 70	Xaa	Xaa	Lys	Phe	Cys 75	Asn	Asp	Leu	Lys	Asn 80
	Ser	Phe	Leu	Asp	Tyr 85	Gly	His	Leu	Ala	Met 90	Gly	Asn	Asp	Met	Asp 95	Phe
50	Gly	Gly	Tyr	Ser 100	Thr	Xaa	Xaa	Xaa	Xaa 105	Xaa	Xaa	<u>X</u> aa	Xaa	Xaa 110		Xaa
	Xaa	Xaa	Xaa 115	Xaa	Xaa	Xaa	Ser	Glu 120	His	Lys	Ile	Lys	Asn 125		Arg	Lys
55	Glu	Trp 130	Trp	Asn	Glu	Phe	Arg	Glu	Lys	Leu	Trp	Glu 140		Met	Leu	Ser
	Glu 145	His	Xaa	Xaa	Xaa	Xaa 150	Xaa	Xaa	Cys	Xaa	Xaa 155	Xaa	Xaa	Xaa	Xaa	Glu 160
	Leu	Gln	Ile	Thr	Gln 165	Trp	Iļe	Lys	Glu	Trp 170	His	Gly	Glu	Phe	Leu 175	
ĐŨ	GIu	Arg	Asp	Asn 180	Arg	ser	Lys	Leu	Pro 185	Lys	Ser	Lys	Cys	Xaa 190		Xaa
	Xaa	Xaa	Xaa 195	Xaa	Xaa	Cys	Xaa	Glu 200	Lys	Glu	Cys	Ile	Asp 205		Cys	Met
65	Lys	Tyr 210		Asp	Trp	Ile	Ile 215		Ser	Lys	Phe	Xaa 220		Xaa	Xaa	Xaa

## (2) INFORMATION FOR SEQ ID NO:27:

```
10 (i) SEQUENCE CHARACTERISTICS:
```

- (A) LENGTH: 277 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

·15

- (ii) MOLECULE TYPE: peptide (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- 20 (vi) ORIGINAL SOURCE:

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

25	Cys	Xaa		Xaa	Xaa	Xaa	Xaa	Cys	Xaa							
25	1	35	••		5	••	••	••		10	_		_		15	
				20	Xaa				25					30	_	_
	Gln	Glu	Leu 35	Cys	Leu	Gly	Asn	Ile 40	Xaa	Xaa	Xaa	Xaa	Xaa 45	Xaa	Xaa	Xaa
30	Xaa	Xaa 50	Xaa	Xaa	Xaa	Xaa	Xaa 55	Xaa	Xaa	Xaa	Xaa	Xaa 60	Xaa	Xaa	Xaa	Xaa
	Xaa 65	Xaa	Xaa	Xaa	Xaa	Xaa 70	Xaa	Xaa	Xaa	Xaa	<b>X</b> aa 75	Xaa	Glu	Val	Cys	Lys 80
35	Ile	Ile	Asn	Lys	Thr 85	Phe	Ala	Asp	Ile	Arg	Asp	Ile	Ile	Gly	Gly 95	
				100	Asp		•		105					110	Xaa	
			115		Xaa			120					125	_		
40		130			Trp		135					140	_			
	145				Xaa	150					155					160
45					Phe 165					170				_	175	_
	Gln	Asp	Lys	Thr 180	Lys	Met	Ile	Glu	Thr 185	Leu	Lys	Val	Glu	Cys 190	Xaa	Xaa
	Xaa	Xaa	Cys 195	Xaa	Asp	Asp	Asn	Cys 200	Lys	Ser	Lys	Cys	Asn 205	Ser	Tyr	Lys
50	Glu	Trp 210	Ile	Ser	Lys	Lys	Lys 215	Lys	Xaa	Xaa	Xaa	Xaa 220	Xaa	Xaa	Xaa	Xaa
	Xaa 225	Xaa	Xaa	Xaa	Xaa	Xaa 230	Xaa	Xaa	Xaa	Xaa	Xaa 235	Xaa	Xaa	Xaa	Xaa	Xaa 240
55	Xaa	Xaa	Xaa	Xaa	Xaa 245	Xaa	Xaa	Xaa	Xaa	Xaa 250	Xaa	Xaa	Суѕ	Xaa	Xaa 255	Xaa
	Xaa	Xaa	Xaa	Xaa 260	Xaa	Xaa	Xaa	Xaa	Xaa 265	Xaa	Xaa	Xaa	Xaa	Xaa 270		Xaa
	Xaa	Cys	Xaa 275	Xaa	Cys											
60						,										

#### (2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 282 amino acids
- 65 (B) TYPE: amino acid

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-79-
           (C) STRANDEDNESS: single
           (D) TOPOLOGY: linear
         (ii) MOLECULE TYPE: peptide
5
         (iii) HYPOTHETICAL: NO
         (iv) ANTISENSE: NO
         (v) FRAGMENT TYPE: internal
         (vi) ORIGINAL SOURCE:
10
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:
    Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Val Cys Gly Pro Pro Arg Arg
15
    Gln Gln Leu Cys Leu Gly Tyr Ile Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                          40
    20
    70
                                   75
    Ala Ile Leu Gly Ser Tyr Ala Asp Ile Gly Asp Ile Val Arg Gly Leu
                                90
    Asp Val Trp Arg Asp Ile Asn Thr Asn Xaa Xaa Xaa Xaa Xaa Xaa
25
             100
                             105
    Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Lys Lys Gln Asn Asp Asn
                         120
                                         125
    Asn Glu Arg Asn Lys Trp Trp Glu Lys Gln Arg Asn Leu Ile Trp Ser
       130
                       135
30
    Ser Met Val Lys His Ile Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa
                   150
                                   155
    Xaa Xaa Xaa Ile Pro Gln Phe Leu Arg Trp Leu Lys Glu Trp Gly
                165
                                170
    Asp Glu Phe Cys Glu Glu Met Gly Thr Glu Val Lys Gln Leu Glu Lys
35
             180
                             185
    Ile Cys Xaa Xaa Xaa Cys Xaa Glu Lys Lys Cys Lys Asn Ala Cys
                          200
                                         205
    Ser Ser Tyr Glu Lys Trp Ile Lys Glu Arg Lys Asn Xaa Xaa Xaa Xaa
                      215
                                      220
40
    230
                                   235
    245
                               250
    45
             260
                            265
    Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
         275
                          280
           (2) INFORMATION FOR SEQ ID NO:29:
         (i) SEQUENCE CHARACTERISTICS:
          (A) LENGTH: 324 amino acids
           (B) TYPE: amino acid
           (C) STRANDEDNESS: single
```

55 (D) TOPOLOGY: linear

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- (ii) MOLECULE TYPE: peptide
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (V) FRAGMENT TYPE: internal
  - (vi) ORIGINAL SOURCE:
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:
- 65 Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa

	1				5					10					15	•
	Xaa	Xaa	Xaa	Xaa 20	Xaa	Xaa	Xaa	Ala	Cys 25	Ile	Pro	Pro	Arg	Arg 30	Gln	Lys
5			35		Tyr			40				•	45			
		50			Xaa		55					60				
	65				Xaa	70					75					80
10					Xaa 85					90			_		95	
				100	Asp				105					110		
15			115		Thr			120					125			
		130			Xaa		135					140	_	_	_	
	145				Asn	150					155	_			-	160
20					Xaa 165					170					175	
				180	Xaa				185					190		
25			195		Xaa			200					205			
		210			Glu		215					220			_	_
20	225				Leu	230					235					240
30					Cys 245					250					255	_
				260	Ser				265			_		270		
35			275		Xaa			280					285			
,		290			Xaa		295					300				
40	305				Xaa	Xaa 310	Суѕ	Xaa	Xaa	Xaa	Xaa 315	Xaa	Xaa	Xaa	Xaa	Cys 320
40	Xaa	Xaa	Xaa	Cys												

#### (2) INFORMATION FOR SEQ ID NO:30:

45 (i) SEQUENCE CHARACTERISTICS:

50

55

- (A) LENGTH: 362 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

WO 96/40766 PCT/US96/09508

-81-

```
Ala Arg Ser Phe Ala Asp Ile Gly Asp Ile Val Arg Gly Lys Asp Leu
             70
                        75
   Tyr Leu Gly Tyr Asp Asn Lys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
5
           85
                      90
   100
                   105
   120
                            125
10
   Phe Phe Gln Leu Arg Glu Asp Trp Trp Thr Ser Asn Arg Glu Thr Val
               135
   Trp Lys Ala Leu Ile Cys His Ala Xaa Xaa Xaa Xaa Xaa Xaa Xaa
             150
                        155
   15
           165
                      170
   . 180
                   185
   Arg Trp Phe Glu Glu Trp Ala Glu Asp Phe Cys Arg Lys Lys Lys
       195
                 200
                           205
20
   Lys Leu Glu Asn Leu Gln Lys Gln Cys Xaa Xaa Xaa Xaa Xaa Xaa Cys
               215
                          220
   230
                        235
   Thr Asn Cys Ser Val Trp Cys Arg Met Tyr Glu Thr Trp Ile Asp Asn
25
           245
                     250
   260
                   265
   275
                 280
                            285
30
   295
                         300
   310
                       315
   Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
35
           325
                      330
   345
   Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
40
```

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 411 amino acids

(B) TYPE: amino acid

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

55 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

```
85
                        90
    Met Leu Ala Arg Ser Phe Ala Asp Ile Gly Asp Ile Val Arg Gly Arg
 5
           100
                      105
                                  110
    Asp Leu Tyr Leu Gly Asn Pro Gln Glu Xaa Xaa Xaa Xaa Xaa Xaa
         115
                    120
                                125
    140
10
    Xaa Xaa Xaa Xaa Xaa Xaa Asn Asp Pro Glu Phe Phe Lys Leu Arg
               150
                           155
    Glu Asp Trp Trp Thr Ala Asn Arg Glu Thr Val Trp Lys Ala Ile Thr
             165
                         170
                                    175
    Cys Asn Ala Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa
15
                      185
                                  190
    195
                    200
    Xaa Xaa Xaa Xaa Val Pro Gln Tyr Leu Arg Trp Phe Glu Glu Trp Ala
                             220
    Glu Asp Phe Cys Arg Lys Lys Asn Lys Lys Ile Lys Asp Val Lys Arg
20
               230
                           235
    245
                        250
    25
                      265
           260
                                  270
    Xaa Xaa Xaa Xaa Cys Ile Ser Cys Leu Tyr Ala Cys Asn Pro Tyr
        275
                    280
                                285
    Val Asp Trp Ile Asn Asn Gln Lys Glu Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                  295
                             300
30
    310
                           315
    325
                        330
    35
           340
                      345
    360
                               365
    375
                           380
40
    390
                           395
    Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
             405
45
         (2) INFORMATION FOR SEQ ID NO:32:
       (i) SEQUENCE CHARACTERISTICS:
        (A) LENGTH: 411 amino acids
         (B) TYPE: amino acid
50
        (C) STRANDEDNESS: single
        (D) TOPOLOGY: linear
       (ii) MOLECULE TYPE: peptide
       (iii) HYPOTHETICAL: NO
55
       (iv) ANTISENSE: NO
       (v) FRAGMENT TYPE: internal
       (vi) ORIGINAL SOURCE:
       (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:
6N
    10
```

```
40
    Xaa Xaa Val Phe Leu Pro Pro Arg Arg Glu His Met Cys Thr Ser Asn
                   55
    5
                70
                             75
    85
                          90
    105
10
    Xaa Xaa Xaa Xaa Xaa Xaa Xaa Ala Met Cys Arg Ala Val Arg Tyr
                      120
    Ser Phe Ala Asp Leu Gly Asp Ile Ile Arg Gly Arg Asp Met Trp Asp
                   135
                                140
    15
                150
                             155
    165
                          170
    Xaa Xaa Xaa Xaa Lys Lys Pro Ala Tyr Lys Lys Leu Arg Ala Asp
                        185
20
    Trp Trp Glu Ala Asn Arg His Gln Val Trp Arg Ala Met Lys Cys Ala
         195
                     200
    Thr Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Ile Pro
      210
                   215
                               220
    Gln Arg Leu Arg Trp Met Thr Glu Trp Ala Glu Trp Tyr Cys Lys Ala
25
                230
                             235
   Gln Ser Gln Glu Tyr Asp Lys Leu Lys Lys Ile Cys Xaa Xaa Xaa
              245
                          250
   Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Cys Gly
           260
                        265
                                     270
30
   Lys Cys Lys Ala Ala Cys Asp Lys Tyr Lys Glu Glu Ile Glu Lys Trp
        275
                     280
                                  285
   Asn Glu Gln Trp Arg Lys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                   295
                               300
   35
                310
                             315
   325
                          330
   340
                        345
40
   355
                     360
                                  365
   Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                   375
                               380
   45
                390
                             395
   Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
              405
         (2) INFORMATION FOR SEQ ID NO:33:
50
       (i) SEQUENCE CHARACTERISTICS:
         (A) LENGTH: 311 amino acids
         (B) TYPE: amino acid
         (C) STRANDEDNESS: single
```

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO

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- (v) FRAGMENT TYPE: internal
  - (vi) ORIGINAL SOURCE:
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:
- Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa

```
10
    Xaa Xaa Xaa Xaa Xaa Ala Cys Met Pro Pro Arg Arg Gln Lys Leu
           20
    5
         35
                     40
                                  45
    55
    70
                             75
10
    Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Gln Phe Leu Arg Ser Met Met
              85
                          90
    Tyr Thr Phe Gly Asp Tyr Arg Asp Ile Cys Leu Asn Thr Asp Ile Ser
           100
                        105
                                    110
    Lys Lys Gln Asn Asp Val Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
15
         115
                     120
    Xaa Xaa Xaa Xaa Ser Lys Ser Pro Ser Gly Leu Ser Arg Gln Glu
                   135
                               140
    Trp Trp Lys Thr Asn Gly Pro Glu Ile Trp Lys Gly Met Leu Cys Ala
    145
                150
                            155
20
    165
                          170
    180
                        185
    Xaa Xaa Xaa Xaa Xaa Lys Pro Gln Phe Leu Arg Trp Met Ile Glu
25
        195
                     200
                                  205
    Trp Gly Glu Glu Phe Cys Ala Glu Arg Gln Lys Lys Glu Asn Ile Ile
                   215.
                               220
   230
                             235
30
   Lys His Arg Cys Asn Gln Ala Cys Arg Ala Tyr Gln Glu Tyr Val Glu
                          250
                                       255
   260
                        265
   35
        275
                     280
                                  285
   295
                               300
   Xaa Xaa Xaa Cys Xaa Cys
40
         (2) INFORMATION FOR SEQ ID NO:34:
       (i) SEQUENCE CHARACTERISTICS:
         (A) LENGTH: 7 amino acids
45
         (B) TYPE: amino acid
         (C) STRANDEDNESS: single
         (D) TOPOLOGY: linear
       (ii) MOLECULE TYPE: peptide
50
       (iii) HYPOTHETICAL: NO
       (iv) ANTISENSE: NO
```

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Pro Arg Arg Gln Xaa Leu Cys

ΰû (2) INFORMATION FOR SEQ ID NO:35:

(vi) ORIGINAL SOURCE:

(i) SEQUENCE CHARACTERISTICS:

(v) FRAGMENT TYPE: N-terminal

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single

```
(D) TOPOLOGY: linear
             (ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
 5
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
             (vi) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEO ID NO:35:
10
      CCRAGRAGRC AARAAYTATG
                                                                           20
                (2) INFORMATION FOR SEQ ID NO:36:
15
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 base pairs
               (B) TYPE: nucleic acid
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
20
             (ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
25
             (vi) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:
      CCSMGSMGSC AGCAGYTSTG
                                                                          20
30
                (2) INFORMATION FOR SEQ ID NO:37:
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 7 amino acids
35
               (B) TYPE: amino acid
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
             (ii) MOLECULE TYPE: peptide
40
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE: N-terminal
             (vi) ORIGINAL SOURCE:
45
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:
      Phe Ala Asp Xaa Xaa Asp Ile
50
                (2) INFORMATION FOR SEQ ID NO:38:
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 base pairs
               (B) TYPE: nucleic acid
55
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
            (ii) MOLECULE TYPE: cDNA
            (iii) HYPOTHETICAL: NO
60
            (iv) ANTISENSE: NO
            (v) FRAGMENT TYPE:
            (vi) ORIGINAL SOURCE:
            (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:
```

	IIIGCWGAIW WWSGWGAIAI	20
	(2) INFORMATION FOR SEQ ID NO:39:	
5	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 20 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
10	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO	
15	(v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:	
20	TTCGCSGATW WCSGSGACAT  (2) INFORMATION FOR SEQ ID NO:40:	20
25	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 6 amino acids</li> <li>(B) TYPE: amino acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
30	<ul> <li>(ii) MOLECULE TYPE: peptide</li> <li>(iii) HYPOTHETICAL: NO</li> <li>(iv) ANTISENSE: NO</li> <li>(v) FRAGMENT TYPE: N-terminal</li> <li>(vi) ORIGINAL SOURCE:</li> </ul>	
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:	
	Pro Gln Phe Xaa Arg Trp 1 5	
40	(2) INFORMATION FOR SEQ ID NO:41:	
45	<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 18 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
50	<pre>(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:</pre>	
55	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:	
	CCAWCKKARR AATTGWGG	18
	(2) INFORMATION FOR SEQ ID NO:42:	
<del>0</del> 0	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single	
65	(D) TOPOLOGY: linear	

```
(ii) MOLECULE TYPE: cDNA
              (iii) HYPOTHETICAL: NO
              (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
 5
             (vi) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:
       CCASCKGWAG AWCTGSGG
                                                                           18
10
                (2) INFORMATION FOR SEQ ID NO:43:
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 7 amino acids
15
               (B) TYPE: amino acid
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
             (ii) MOLECULE TYPE: peptide
20
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE: N-terminal
             (vi) ORIGINAL SOURCE:
25
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:
      Glu Trp Gly Xaa Xaa Xaa Cys
                        5
30
                (2) INFORMATION FOR SEQ ID NO:44:
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 base pairs
               (B) TYPE: nucleic acid
35
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
             (ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
40
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
             (vi) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:
45
      CAAWAWTCWT CWCCCCATTC
                                                                           20
                (2) INFORMATION FOR SEO ID NO:45:
50
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 base pairs
               (B) TYPE: nucleic acid
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
55
             (ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
60
             (VI) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:
      CAGWASTCST CSCCCCACTC
                                                                           20
65
```

#### WE CLAIM:

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- A composition comprising a nucleotide sequence of the DBL gene family, wherein said nucleotide sequence is selected from the group consisting of the ver-1, var-2, var-3 and var-7 genes.
- 2. The composition of Claim 1, wherein the nucleotide sequence of the *var-1*, *var-2*, *var-3* or *var-7* gene encodes a cysteine-rich domain homologous to a cysteine-rich domain of a Duffy Antigen Binding Protein (DABP) derived from *Plasmodium vivax* and a Sialic Acid Binding Protein (SABP) derived from *Plasmodium talciparum*.
- 3. The composition of Claim 1, wherein the nucleotide sequence of the *var-1*, *var-2*, *var-3* or *var-7* gene encodes a cysteine-rich interdomain region between a first domain and a second domain.
- 4. The composition of Claim 1, wherein the nucleotide sequence is derived from a coding region of SEQ ID NO:13 or SEQ ID NO:15.
  - 5. A composition comprising a polypeptide encoded by a nucleotide sequence of the *DBL* gene family, wherein said polypeptide is encoded by a *var-1*, *var-2*, *var-3* or *var-7* gene.
  - 6. The composition of claim 5, wherein the polypeptide comprises a sequence of amino acid residues homologous to cysteine-rich domains of a Duffy Antigen Binding Protein (DABP) derived from *Plasmodium vivax* and a Sialic Acid Binding Protein (SABP) derived from *Plasmodium falciparum*.
  - 7. The composition of claim 5, wherein the polypeptide comprises a sequence of about 300 to 400 amino acid residues occurring in the cysteine-rich interdomain region between a first domain and a second domain of a polypeptide encoded by the var-1, var-2, var-3 or var-7 gene.
  - 8. The composition of claim 5, wherein the polypeptide comprises a sequence of amino acid residues of SEO ID NO:14 or SEO ID NO:16.
  - 9. The composition of claim 5, wherein the polypeptide comprises a sequence of about 50 to about 325 amino acid residues of SEQ ID NO:14 or SEQ ID NO:16.
  - 10. The composition of claim 5, wherein the polypeptide comprises a sequence of about 75 to about 300 amino acid residues of SEQ ID NO:14 or SEQ ID NO:16.
  - 11. The composition of claim 5, wherein the polypeptide comprises a sequence of about 100 to about 250 amino acid residues of SEQ ID NO:14 or SEQ ID NO:16.
  - 12. The composition of claim 5, further comprising a pharmaceutically acceptable carrier and an isolated Duffy Antigen Binding Protein (DABP) binding domain polypeptide, a Sialic Acid Binding Protein (SABP) binding domain polypeptide, or a combination thereof, in an amount sufficient to induce a protective immune response to *Plasmodium* merozoites in a mammal.
  - 13. The composition of any of the preceding claims for use in inducing a protective immune response to *Plasmodium* merozoites in a mammal.
  - 14. Use of the composition of any one of claims 1-12 in the preparation of a medicament for inducing a protective immune response to *Plasmodium* merozoites in a mammal.
  - 15. A method of inducing a protective immune response to Plasmodium merozoites in a mammal, comprising administering to a mammal an immunologically effective amount of a pharmaceutical composition

comprising a pharmaceutically acceptable carrier and an isolated cysteine-rich polypeptide encoded by a var gene selected from the group of genes consisting of var-1, var-2, var-3 and var-7 genes.

16. The method of claim 15, further comprising administering to said mammal an immunologically effective amount of a Duffy Antigen Binding Protein (DABP) binding domain polypeptide, a Sialic Acid Binding Protein (SABP) binding domain polypeptide, or a combination thereof.

DMEGIGYSK-X <sub>11</sub> - DMDFGGYST-X <sub>17</sub> - DYWNDLSNR-X <sub>15</sub> - DVWRDINTN-X <sub>17</sub> -	DLYLGYDNK-X37- DLYLGNPQR-X30- DMWDEDKSS-X32- DISKKQNDV-X15-	XVSELPTEVQKLKEKCX <sub>11</sub> ELLERDNRSKLPKSKCX <sub>8</sub> - KCQDKTKMIETLKVECX <sub>4</sub> - ?CEEMGTEVKQLEKICX <sub>4</sub> -	ECRKKKKLENLQKQCX6- ECRKKNKKIKDVRRNCX12 CCKAQSQEYDKLKKICX11 CCABRQKKENIIKDACX8	FIG. 1	
G-X <sub>12</sub> -G-X <sub>5</sub> VCIPDRRYQLCMKEL-X <sub>4</sub> 7- DFCKDIRWSLGDFGDIIMGTDMEGIGYSK-X <sub>11</sub> -C-X <sub>10</sub> -C-X <sub>10</sub> -VCIPDRRIQLCIVNL-X <sub>3</sub> 6- KFCNDLKNSELDYGHLAMGNDMDFGGYST-X <sub>1</sub> 7-C-X <sub>13</sub> -C-X <sub>10</sub> -VCVPPRRQELCLGNI-X <sub>3</sub> 6- EVCKIINKTEADIRDIIGGTDYWNDLSNR-X <sub>15</sub> -C-X <sub>11</sub> -VCGPPRRQCLCLGYI-X <sub>3</sub> 6- KICNAILGSXADIGDIVRGLDVWRDINTN-X <sub>17</sub> -	C-X15-C-X15-ACAPYRRLHLCDYNL-X43-QLCTVLARSEADIGDIVRGKDLYLGYDNK-X37-C-X15-ACAPYRRLHVCDQNL-X45-QICTMLARSEADIGDIVRGRDLYLGNPQE-X30-C-X17-C-X31-VFLPPRREHMCTSNL-X55-AMCRAVRYSEADLGDIIRGRDMWDBDKSS-X32-C-X10-C-X10-ACMPPRRQKLCLYYI-X52-QFLRSMMYTEGDYRDICLNTDISKKQNDV-X15-C-X10-C-X11-ACIPPRRQKLCLHYL-X51-DFKRQMFYTEADYRDICLGTDISSKKDTS-X15-	TDEKAQQRRKQWWNBSKAQIWTAMMYSV-X <sub>11</sub> -C-X <sub>8</sub> ePQIYRWIREWGRDYVSELPTEVQKLKEKCX <sub>11</sub> C-X <sub>1</sub> SEHKIKNFRKEWMNBFREKLWRAMLSEH-X <sub>6</sub> C-X <sub>6</sub> eLQITQMIKEWHGEELLERDNRSKLPKGKCX <sub>8</sub> C-X <sub>0</sub> NKKNDKLFRDEWWKVIKKDVWNVISWVF-X <sub>5</sub> C-X <sub>7</sub> IPQFFRWFSEWGDDYCQDKTKMIETLKVECX <sub>4</sub> C-X <sub>1</sub> - KKQNDNNERNKWWEKQRNLIWSSMVKHI-X <sub>5</sub> C-X <sub>8</sub> IPQFLRWLKEWGDEECEEMGTEVKQLEKICX <sub>4</sub> C-X <sub>5</sub>	KGGDFFQLREDMMTSNRETVWKALICHA-X <sub>11</sub> -C-X <sub>23</sub> -VPQYLRWFEEWAEDECRKKKKKLENLQKQCX <sub>6</sub> C-X <sub>15</sub> NDPBFPRLREDMMTANRETVWKAITCNA-X <sub>9</sub> C-X <sub>23</sub> -VPQYLRWFEEWAEDECRKKNKKIKDVKRNCX <sub>12</sub> C-X <sub>22</sub> KKPAYKKLRADMWEANRHQVWRAMKCAT-X <sub>4</sub> C-X <sub>8</sub> IPQRLRWTEWAEWACKAQSQBYDKLKKICX <sub>11</sub> C-X <sub>6</sub> - SKSPSGLSRQEWWKTNGPBIWKGMLCAL-X <sub>3</sub> 7KPQFLRWMIEWGEBECABRQKKENIIKDACX <sub>8</sub> C-X <sub>3</sub> -	MITRKKN-X56CXC HIIRSKP-X41-C-X7CXC HISKKKK-X36-C-X20CXX-C MIKERKN-X38-C-X19CXX-C	CTNCSVWCRMXET MIDNQKK-X68-C-X30CXX-C CISCLYACNPYVD HINNQKE-X69-C-X40CXX-C CGRCKAACDKYKEBIBKENEQWRK-X73-C-X6-C-X30-CXX-C KHRCNQACRAXQB XVENKKK-X43-C-X4CXC CVACKDQCKQXHS MIGIWID-X42-C-X8CXXXC
G-X <sub>12</sub> -G-X <sub>5</sub> VGI C-X <sub>10</sub> -C-X <sub>9</sub> VCI C-X <sub>13</sub> -C-X <sub>10</sub> -VCV C-X <sub>12</sub> -C-X <sub>11</sub> -VCG	C-X15-G-X15-ACA C-X17-C-X31-VFL C-X10-C-X10-ACM C-X10-C-X11-ACM	Tdekaqqrrkomenes Sehkiknfrremene Nkkndklfrdemevi Kkondnnernkmerk	KGGDFFQLREDMMTSN NDPRPPKLREDMMTAN KKPAYKKLRADMMEAN SKSPSGLSRQEMMKTN KISNSIRYRKSMMETN	VPPCQNACKSYDQ BKECIDPCMKYRD DDNCKSKCNSYKE EKKCKNACSSYEK	CTNCSVWCRMXET CISCLYACNPYVD CGKCKAACDKYKBEIBI KHRCNQACRAYQB CVACKDQCKQYHS
DABP F1 SABP F1 SABP F2 EBL-e1	HBL-e2 Proj3 F1 Proj3 F2 Proj3 F3	DABP SABP F1 SABP F2 EBL-e1	EBL-e2 Proj3 F1 Proj3 F2 Proj3 F3 E31a	Dabp Sabp F1 Sabp F2 Ebl-el	ENL-e2 Proj3 F1 Proj3 F2 Proj3 F3 Eila
Family 1	Family 2	Family 1 Cont'd	Family 2 Cont'd	Family 1 Cont'd	Family 2 Cont'd

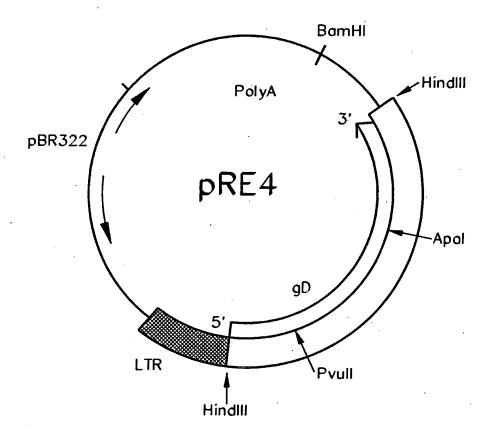


FIG. 2

3/5

# FIG. 3

Concensus amino acid sequences and the synthetic oligonucleotide primers designed from them.

UNIEBP5 and 5A: PRRQ K/E L C

UNIEBP5, for A+T biased codon usage: CC(A/G)-AG(G/A)-AG(G/A)-CAA-(G/A)AA-(C/T)TA-TG

UNIEBP5A, for G+C biased codon usage: CC(C/G)-(C/A)G(C/G)-(C/A)G(C/G)-CAG-CAG-(C/T)T(C/G)-TG

UNIEBP5 B and C: F A D I/Y G/R D I

UNIEBP5B, for A+T biased codon usage: TTT-GC(A/T)-GAT-(A/T)(A/T)-(G/C)G(A/T)-GAT-AT

UNIEBP5C, for G+C biased codon usage: TTC-GC(G/C)-GAT-(A/T)(A/T)C-(G/C)G(G/C)-GAC-AT

UNIEBP3 and 3A: P Q F L/F R W

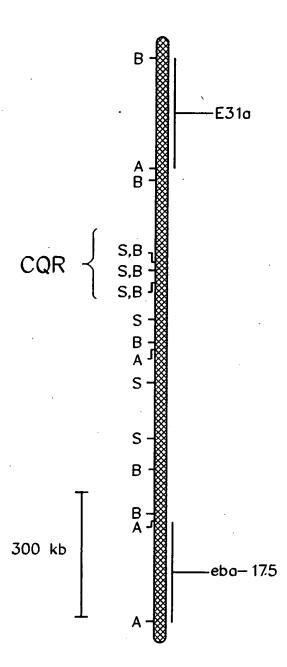
UNIEBP3, for A+T biased codon usage: CCA-(A/T)C(T/G)-(T/G)A(A/G)-(A/G)AA-TTG-(A/T)GG

UNIEBP3A, for G+C biased codon usage: CCA-(C/G)C(G/T)-G(A/T)A-GA(A/T)-CTG-(C/G)GG

UNIEBP3 B and C: E W G D/E D/E Y/F C

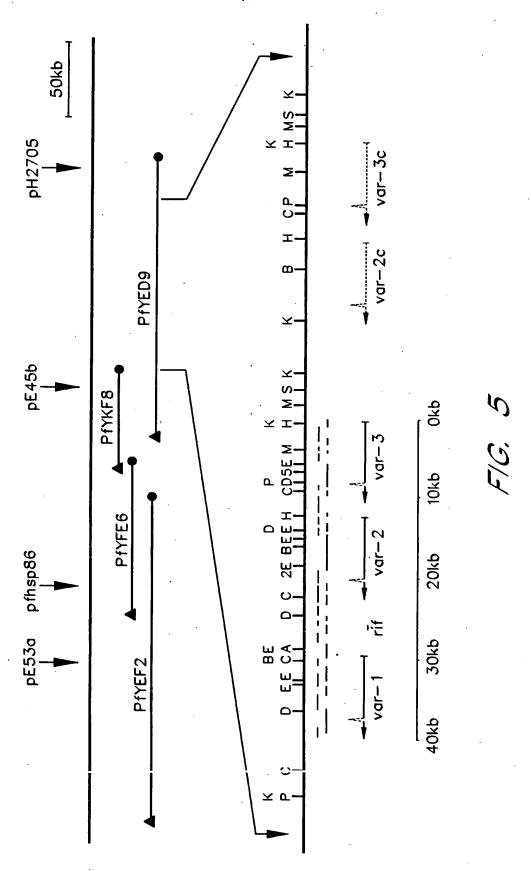
UNIEBP3B, for A+T biased codon usage: CA-A(A/T)A-(A/T)TC-(A/T)TC-(A/T)CC-CCA-TTC

UNIEBP3C, for G+C biased codon usage: CA-G(A/T)A-(G/C)TC-(G/C)TC-(G/C)CC-CCA-CTC G+C Biased



F/G. 4

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